Histological study on the hazardous effects of ethanol on liver and spleen in Swiss albino mice

Badr Abdullah Aldahmash¹, Doaa Mohamed El-Nagar²

¹ Medical Laboratory Department, College of Health Sciences, King Saud University, Riyadh, Kingdom of Saudi Arabia,
² Faculty of Sciences and Humanities, University of Salman bin Abdul Aziz, Kingdom of Saudi Arabia.

Abstract

Considering that ethanol caused number of health problems in the world, the present study was initiated to investigate the histological hazardous effects of ethanol on the liver and spleen. Animals were divided into 4 groups, the first group served as a control group, the second, third and fourth received 1.2 and 6 ml/kg/bw of 70% ethanol respectively. At the day 5 post treatment, the liver and spleen sections were prepared for the histological study. Ethanol intake induced marked histological alterations in the liver and spleen that correlated with the dose taken, low dose induced liver and spleen injury mainly as cytoplasmic degeneration in liver and abnormal structure of spleen, whereas, high doses of ethanol resulted in fibrosis in liver and splenomegaly.

Key words: Ethanol, mice, liver, spleen.

Introduction

Ethanol has been a part of the human diet for centuries. However, its consumption in excess can result in a number of health problems, most notably liver damage. Ethanol cannot be excreted and must be metabolized, primarily by the liver (Berg et al., 2002). Alcoholic Liver Disease (ALD) is a blanket term in which conditions related specifically to the liver and alcohol use fall under. The most prevalent types of Alcoholic Liver Disease, or ALD, are fatty liver, alcoholic hepatitis, and cirrhosis. Often, as people continue to drink heavily, they progress from fatty liver to hepatitis to cirrhosis. All three of the disorders can occur together (Robert et al., 2004). Fatty Liver, which occurs after acute alcohol ingestion, is generally reversible with abstinence and is not thought to predispose to any chronic form of liver disease if abstinence and/or moderation are maintained (Kyrsten, 2004). It is estimated by the National Institutes of Health that about 20 percent of alcoholics and heavy drinkers develop fatty liver, or steatosis (Robert et al., 2004). Other sources estimate that 90-100 percent of patients with heavy drinking will develop this disease (Ismail and Riely, 2006). The condition can lead to death if alcohol consumption is not reduced or stopped. Alcohol can be a factor in several other forms of liver disease not specifically attributed to it, and may interact with risk factors for other forms of liver disease. An example of this is people with alcohol-related cirrhosis are at a higher risk of developing liver cancer. Those with Hepatitis B or C accompanied with heavy drinking are at a much higher risk of cirrhosis than with heavy drinking alone (Robert et al., 2004). Clinical evidence supports a correlation between excessive alcohol consumption and certain bacterial infections. For example, alcoholics who have developed cirrhosis of the liver are predisposed to spontaneous bacterial peritonitis. Phagocytes are an important defense against infection in this part of the body and defects in phagocytic cell function, observed in many alcoholics, may predispose these individual to peritoneal infection (Roselle, 1992). There is considerable evidence indicating that ethanol consumption alters immune system function and leads to increased susceptibility to infections and neoplastic diseases (Nath and Szabo, 2009; Lau et al., 2009; Nava-Aguiera et al., 2009; Szabo and Mandrekar, 2009).

Materials and Methods

Animals and experimental design

Male swiss mice weighed 25 to 30 g were obtained from King Saud university animal house. Mice were provided with water and balanced diet ad libitum. 20 adult swiss mice were randomly divided into 4 groups of 5 mice in each as following:
- Group 1: control group (normal mice without treatment).
- Group 2: mice treated with 1ml/kg/bw of 70% ethanol orally by stomach gavage for consecutive 5 days.
- Group 3: mice treated with 2ml/kg/bw of 70% ethanol orally by stomach gavage for consecutive 5 days.
- Group 4: mice treated with 6ml/kg/bw of 70% ethanol orally by stomach gavage for consecutive 5 days.

**Histological examination**

Mice were sacrificed in the sixth day of the beginning of the experiment, livers and spleens were collected and cut into small pieces and fixed 10% neutral buffered formalin, then embedded in paraffin and sectioned. The sections were stained with Hx&E stain for histopathological examination and photographing.

**Results**

1. **Liver**

   The liver of normal mice (untreated) consists of a vast interanastomosing network of hepatocytes arranged in single-cell thick plates separated by vascular sinusoids. The hepatocytes along with vascular channels form organized micro structures which serve as structural and functional units. The liver is composed of innumerable lobules, each of which is a hexagonal structure consisting of a central vein surrounded by radiating hepatocyte plates. However, another concept of a functional unit defines an acinus as the functional unit in relation to terminal portal branches and terminal hepatic venule. Portal tracts surround the classical lobules. An interlobular portal vein is also shown (Figure 1).

   Histopathological examination of liver of mice received 1ml/kg/bw of 70% ethanol orally for 5 days showed hazardous effects on liver represented by appearance of many necrotic foci (N), vacuolar degeneration (vc) in the cytoplasm of hepatocytes, some cells showed abundant nuclei, others showed pyknotic (P) nuclei where others appeared without nuclei. Dilatation in blood sinusoids with kupffer (K) cells besides to existence of lymphocytic infiltration foci were abundant (L) (Figures 2a&2b). Sections stained by masson’s trichrome technique showed wide dilated and branched central vein (CV) (Figure 2c), moreover, Figure 2d revealed congested and dilated central vein with destructed wall and surrounded by necrotic areas.

   Examination of liver sections of mice received oral administration of 2ml/kg/bw of 70% ethanol for 5 days showed more complicated alterations remarked by destruction in the vein wall surrounded by aggregations of lymphocytes and necrotic areas (N), in addition, pyknotic nuclei were abundant in some cells, and other cells showed complete degeneration in its nuclei, moreover, sections revealed fusion of cells due to degeneration of cell walls (Figures 3a&3b). Whereas, trichrome stained sections (Figures 3c&3d) displayed wide necrotic areas were filled with erythrocytic exudates, in addition to, wide dilated blood sinusoids with kupffer cells, dilated portal vein appeared surrounded by concentric layers of collagenous fibers with lymphocytes in between.

   Livers of mice received oral administration of 6ml/kg/bw of 70% ethanol revealed the worst hazardous features in liver tissues compared with previous doses manifested by cytoplasmic and nuclear degeneration in hepatocytes, some cells showed pyknotic nuclei (p), dilatation in blood sinusoids in addition to appearance of kupffer cells in the sinusoids, presence of necrotic areas besides to blood exudates among liver cells and appearance of blood congestion in dilated central vein (Figure 4a), moreover, Figure 4b showed some aggregations of lymphocytes in liver tissue. Concentrated
layers of collagenous fibers were deposited in the tissue accompanied by lymphocytic infiltration and blood exudates, moreover, dilated central vein surrounded by thick concentric layers of collagenous fibers besides to presence of necrotic areas were seen (Figures 4c&4d).

2. Spleen

Spleen is a large lymphoid organ that contains two main compartments known as “the white pulp and the red pulp”. Spleen is covered with a fibrous capsule from which trabeculae enters into the parenchyma. The fibrous trabeculae ramify throughout the spleen and form supportive sheathing around blood vessels. The white pulp consists of lymphoid follicles. Each B-lymphoid follicle contains a distinct marginal zone of lymphocytes (outer rim of lymphocytes) around the mantle zone of the follicle (inner rim of lymphocytes). The marginal zone consists of loosely arranged lymphocytes. This ‘perifollicular zone’ is the boundary between white and red pulp and serves as the area where macrophages are abundant. The macrophages serve as the ‘custom officers’ for the newly entered red and white blood cells and particulate material. The red pulp is the area of spleen in between white pulp and consists of open sinuses and cellular cords. Splenic sinuses are open vascular spaces lined by a discontinuous layer of endothelial cells and supported by a fenestrated basal lamina and reticular fibers. The surrounding cellular splenic cords provide a tissue framework maintaining the network of sinuses.

Figure 2. Liver sections treated with 1ml/kg bw of ethanol showed marked changes (2a mag.x400 H&E) necrotic areas (N), vacuolar degeneration (VC), kupffer cells (K) and blood sinusoids (S) (2b mag.x400 H&E) lymphocytic infiltration (L) and pyknotic nuclei (P). (2c mag.x400 M.Tr.) dilated central vein, (2d mag.x400 M.Tr.) dilated central vein with erythrocytic congestion surrounded by necrotic foci (N).
Examination of mice spleen sections received 1 ml/kg/bw oral administration of 70% ethanol for 5 days showed marked changes in the spleen tissue represented by expansion of red pulp due to venous congestion, presence of number of resident macrophages to get rid of abnormal red blood cells and cellular inclusions due to the venous congestion (Figures 6a & 6b). Dilatation in venous sinuses that lined with endothelial cells besides to necrotic foci appeared in Figures 6c and 6d.

Spleen sections of mice received oral administration of 2 ml/kg/bw of 70% ethanol revealed abnormal architecture of spleen tissue remarked by distorted lymphoid follicles (white pulp) with resident macrophages in the marginal zone of the follicles, increasing the expansion of red pulp due to the increasing of venous congestion in addition to the dilatation in the venous sinuses (Figures 7a, 7b & 7c), whereas, Figure 7d showed wide necrotic areas in the lymphoid follicle due to the degeneration of B and T lymphocytes.

By increasing the dose of 70% ethanol to 6ml/kg/bw spleen sections showed very hazardous effects on the spleen represented by splenomegaly due to severe congestion that venous sinuses completely disappeared because of filling of red blood cells, in other side, distorted white pulp appeared with small sized lymph follicles surrounded by necrotic areas (Figures 8a & 8b).

**Discussion**

Ethanol, which is a weak anesthetic, is toxic for both humans and animals, and exerts important negative effects upon the liver, brain, heart, skeletal muscle, pancreas, hematological and immune systems, gastrointestinal apparatus and endocrine system (Rottenberg, 1992, Rodés et al.,1990 and...
Figure 4. Liver sections treated with 6ml/kg/bw of ethanol showed hazardous features (4a mag.x400 H&E) necrotic area (N), pyknotic nuclei (P), hepatocytic degeneration (d) (4b mag. X400 H&E) aggregations of lymphocytes (L) (4c mag.x400 M.Tr.) collagenous fibers (F) stained blue in the tissue, lymphocytic infiltration (L) and blood exudates (EX). (4d mag.x400 M.Tr.) dilated central vein (CV) surrounded by collagenous fibers (F) and wide necrotic areas (N).

Figure 5. Untreated spleen sections showed normal architecture (5a mag.x100 H&E) white pulp (WP) and red pulp (RP) (5b mag.x400 H&E) lymphoid follicle consists of lymphocytes (L), macrophages (M)
Bujanda et al., 1999). Alcohol can cause liver damage in the form of steatosis or fatty liver, fibrosis and liver cirrhosis. In general, the amount and duration of alcohol abuse correlate with the presence and severity of liver damage, at least as regards the initial stage of fatty liver (Becker et al., 1996). The present experimental study runs in full agreement with the previous studies that ethanol intake exerts hazardous effects upon liver and spleen, moreover, the amount of ethanol received correlates with severity damage of liver and spleen, it was observed that liver and spleen of mice received high doses of 2.6 ml/kg/bw of ethanol showed severe damage in the tissues compared to that received 1ml/kg/bw of ethanol.

Many studies proved that fatty changes, necrosis and inflammation were prominent in the livers of rats fed on ethanol (Gouillon et al., 2000). reported that Wistar rats livers exposed to 10% v/v ethanol for 12 weeks showed important changes in hepatic trabecular structure and increased hepatocytes with cytoplasmatic vacuoles (Brzóska et al., 2003 and Ito et al., 2007). The characteristic Liver Alcoholic Disorder (steatosis and fibrosis is not observed when ethanol concentrations are low (Harrison & Burt, 1993). In accordance with the previous results, the present study proved that administration of low doses of ethanol affected mice liver histology represented by necrosis and cytoplasmic vacuoles in liver, but there is no fibrosis was detected in the liver tissue, whereas, sections treated with high doses of ethanol showed accumulation of fibers that increased by increasing the dose, the appearance of fibrosis might be due to increasing of hepatocytic degeneration which replaced by fibrosis. Moreover, low doses of ethanol caused harmful effects on the spleen manifested by expansion of red pulp and increasing number of macrophages to get rid of cellular inclusions as the dose increased, the expansion and congestion of red pulp increased whereas, the white pulp decreased and distorted which resulted in splenomegaly.
Figure 7. Treated spleen sections with 2ml/kg/bw revealed more changes (7a mag.x400 H &E) distorted white pulp (WP), increasing number of macrophages (M), (7b mag.x400 H &E) small lymphoid follicle (WP), expansion of red pulp (RP) (7d mag.x400 H &E) abnormal white pulp (WP), congested red pulp (RP) (7d mag.x400 M.Tr.) necrotic foci (N).

Figure 8. Treated spleen sections with 6ml/kg/bw of ethanol showed severe alterations (8a mag.x400 H &E) severe congestion in red pulp (RP), shrunken lymphoid follicle (LF), necrosis (N) (8b mag. x400 H&E) distorted lymphoid follicle(LF), necrosis (N), trabeculae (T).
References


Corresponding Author
Badr Abdullah Aldahmash,
Medical Laboratory Department,
College of Health Sciences,
King Saud University,
Riyadh,
Kingdom of Saudi Arabia,
E-mail: dr_badr211@hotmail.com