



Physiological Aspects of Arabian Sand Gazelle Semen and its Cryopreservation

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ABSTRACT

*The distribution of Arabian Sand Gazelles (*Gazella subgutturosa marica*) has declined dramatically during recent decades, apparently due to excessive hunting and habitat degradation. The Saudi Wildlife Commission has a research centers for some endangered species, one of them is the sand gazelle. Also the activities includes captive breeding, habitat protection, and reintroduction. There are no studies of the semen characteristics and its cryopresercation and artificial insemination of the Arabian Sand Gazelle, so this study was focused to described semen characteristics and Cryopreservation by using Triladyl and Tris Diluents as extender for semen cryopreservation of the Arabian Sand Gazelle Males. Information on the Physiological Aspects of The Arabian Sand Gazelle semen and its cryopresercation is very limited in this respect and references on other species will be used where available.*

Key words: Sand Gazelle, (*Gazella subgutturosa marica*), Semen, Cryopreservation, Triladyl, Tris Diluent, Glycerol

INTRODUCTION

The Saudi Wildlife Commission in Saudi Arabia make huge efforts in keeping and breeding endangered species, with a view to preserve them from extinction and ideally to reintroduce them back in their natural habitats.

The distribution of sand gazelles (*Gazella subgutturosa marica*) has declined dramatically during recent decades, apparently due to excessive hunting and habitat degradation. Today, sand gazelles survive only in small numbers in a few isolated parts of their former range. The Saudi Wildlife Commission has a research centers for some endangered species, one of them is the sand gazelles. Also the activities includes captive breeding, habitat protection, and reintroduction [14].

Little is known about the reproduction of large mammals inhabiting arid environments, and less is known about the mammals of the Arabian Peninsula, especially the Arabian sand gazelles [1,2,3,4,5]. There are no studies of the semen characteristics, its cryopresercation, and artificial insemination of the Arabian Sand Gazelle except that published by [2,3,4]. That study was focused to described Semen characteristics and Cryopreservation by using Triladyl and Tris Diluents as extender for semen cryopreservation of the Arabian sand Gazelle Males.

The order Artiodactyla, the even-toed ungulates, is comprised of the major families *Bovidae* (antelope, cattle, bison, buffalo, sheep, and goats), *Cervidae* (deer), *Suidae* (pigs), *Tayassuidae* (peccaries), *Hippopotamidae* (hippopotamuses), *Giraffidae* (giraffe and okapi), *Camelidae*, and the smaller families of *Antilocapridae* (pronghorn antelope), and *Tragulidae* (chevrotains). According to the 2007 IUCN Red List, of 214 species of Artiodactyla, described, two are listed as extinct in the wild, 14 are critically endangered, 26 are endangered, and 35 are considered vulnerable [3,4]. For the purpose of this review, the term non-domestic bovids refers to members of the *Bovidae* family, but excludes those that can be considered domestic livestock such as cattle, buffalo, bison, sheep, and goats [19].

Semen storage technology was revolutionized approximately 50 years ago by the discovery that glycerol could act as a cryoprotectant. The discovery of glycerol as a cryoprotectant marked a

quantum advance in semen cryopreservation, but subsequent research has only made relatively small improvements to the basic techniques established in the early 1950s.

Glycerol concentrations of less than 2% were necessary to achieve good fertility also after freezing of boar semen, although maximum sperm motility was seen at a concentration of 7% glycerol [8, 10, 15, 18].

In 1957, Canadians Barker and Gandier reported the first foaling following insemination with frozen epididymal spermatozoa. During the next decade a dozen more successful reports with ejaculated semen followed from Germany, Japan, Russia, and the U.S. Sugar-containing diluents were most commonly used. A study by Garde, J., *at al.*, [12] on Sperm Cryopreservation in Three Species of Endangered Gazelles (*Gazella cuvieri*, *G. dama mhorh*, and *G. dorcas neglecta*) were used five diluents to compared cryopreservation for these three species by using Glycerol concentration between 6- 8 %.

This quantitative difference between species is an important determinant of the fertility of cryopreserved semen and means that development of successful freezing procedures necessarily involves more than the identification or application of novel cryoprotectants and additives [12].

Information on the Physiological Aspects of The Arabian Sand Gazelle semen and its cryopreservation is very limited in this respect and references on other species will be used where available .

PRINCIPLES OF CRYOPRESERVATION

Freezing rate

During the cryopreservation process, cells are exposed to cryoprotective agents such as glycerol, ethylene glycol or dimethylsulfoxide in an extender. The addition of 'cryoprotectant' agents exposes the cells to a hypertonic environment, this causes the cells to initially shrink, but they regain their normal volume as the cryoprotectant enters the cells. Cooling also changes the cell membrane. During freezing, cells again shrink as water flows out of the cell and ice forms in the extracellular space. Depending upon the cooling rate, ice may form inside the cell as well [22].

Formation of intracellular ice usually results in damage to the cell membrane, it loses its semi-permeable properties, resulting in death of the cell. During thawing, the cells undergo similar changes in volume as water moves back into the cell. The thaw rate and cell survival are dependent upon the cooling rate used. As stated by a researcher considering the difficulties in semen cryopreservation of other species compared to the bull, it causes one to marvel at successful survival rather than to despair over the difficulties of its implementation to the preservation of a variety of other sperm cell types [9, 15, 17].

Supercooling and seeding

The temperature of the semen will invariably cool below the freezing point before freezing occurs (super cooling). A high degree of super cooling is generally considered unwanted because it will cause a rapid upward and downward temperature fluctuation. Seeding at a minimum temperature below the freezing point will initiate ice crystal formation and avoid the temperature drop. Seeding may be initiated by vibration or touching of the straw with an instrument colder than the package as is done in embryo freezing. Seeding will otherwise be initiated in the straw where it touches the freezing rack. Most heat is conducted away from the straw through the metal rack if the rack is in contact with the liquid nitrogen and therefore colder than the vapor. Freezing racks offering multiple contact points have been used for maxi straws (2.5–5 ml) but there is no data indicating an advantage for such racks.

Cryoprotectants

Life is a complex chemical process that happens in water. Without liquid water, there is no life, or at least no life process. Cryoprotectants are chemicals that protect living things from being injured by water freezing during exposure to cold [6].

Water expands when it freezes, but contrary to popular belief it is not expansion of water that causes injury. It is the purification of water during freezing that causes injury. Water freezes as a pure substance that excludes all else. It is this exclusion process that causes injury. Instead of remaining a solvent that allows the molecules of life to freely mix within it, water that freezes gathers itself up into crystals pushing everything else out. This is illustrated in Figure 1. (a, b).

Freezing causes damage by two distinct mechanisms. The first is mechanical damage as the shape of cells is distorted by ice crystals. The second is damage caused by chemical and osmotic effects of

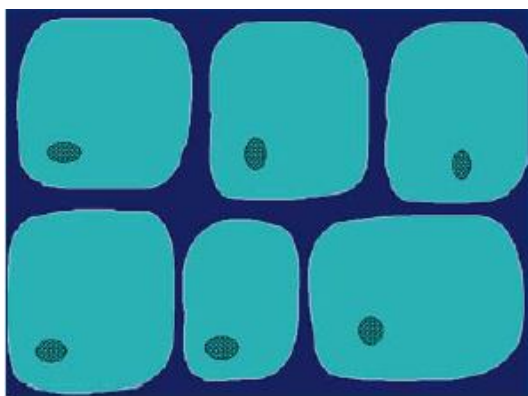


Figure 1a. Cells before freezing

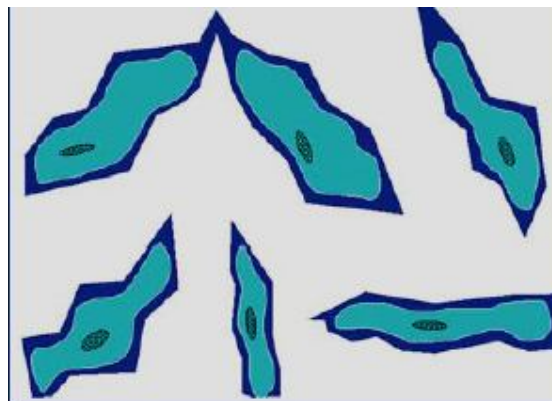


Figure 1b. Cells after freezing.

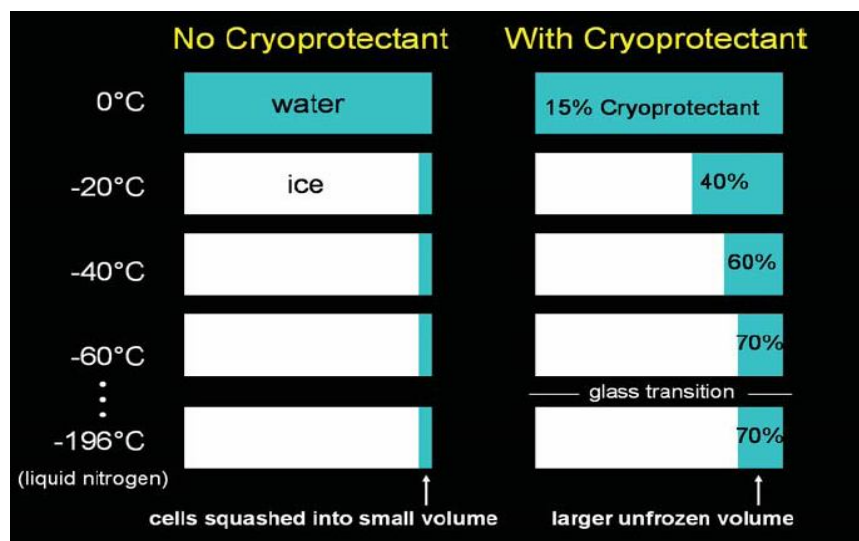


Figure 2. Water when frozen without and with added cryoprotectant.

Without cryoprotectant, almost the entire water volume freezes during cooling. Only salts and other dissolved molecules prevent water from freezing completely. With cryoprotectant, the percentage of cryoprotectant present in solution increases as ice grows. At any given temperature, ice growth stops when the cryoprotectant becomes concentrated enough to make the melting point equal to the surrounding temperature. Eventually the cryoprotectant reaches a concentration that cannot be frozen. No more ice can grow as the temperature is lowered, and there is more room for cells to survive between ice crystals. Below approximately -100°C, the remaining unfrozen liquid pocket solidifies into a glass, permitting storage for practically unlimited periods of time. Cells survive freezing by existing inside the glassy solid between ice crystals. The larger the starting cryoprotectant concentration, the larger the unfrozen volume will be at the end of freezing.

concentrated solutes in the residual unfrozen water between ice crystals. This is so-called “solution effects” injury [6, 7, 21, 23].

On the other hand many of The Cryoprotectants used for semen cryopreservation of gazelles (like *Gazella gazella* and *Gazella dorcas*) and one subspecies (*Gazella gazelle acaiae*) as mentioned by Joseph Saragusty, *et al*, also Cuvier gazelle (*Gazella cuvieri*) by Garde, J., *et al* in 2003 and 2005, and sand gazelles (*Gazella subgutturosa marica*) as mentioned by (Al-eissa, M.S., *et al*, b,c) [16], [12, 13], [3,4].

How Cryoprotectants Protect Cells

Cryoprotectants are chemicals that dissolve in water and lower the melting point of water. For applications outside cryobiology, such chemicals are sometimes called “antifreeze”. Common examples are glycerol, ethylene glycol, propylene glycol, and dimethylsulfoxide (DMSO). A cryoprotectant concentration of about 5% to 15% is usually all that is required to permit survival of a substantial fraction of cells after freezing and thawing from liquid nitrogen temperature.

Figure 2. shows the essential concept of cryoprotection during cell freezing. Growing ice compacts cells into smaller and smaller pockets of unfrozen liquid as the temperature is lowered. The presence of cryoprotectants makes these pockets larger at any given temperature than they would be if no cryoprotectant were present. Larger unfrozen pockets for cells reduces damage from both forms of freezing injury, mechanical damage from ice and excessive concentration of salt [6].

Diluents and Dilution

Freezing of undiluted semen with some success is possible only with human semen [24]. In early studies with bull semen, the expression “glycerol equilibration time” was coined [20]. It is possible that the long time necessary between dilution and freezing in bull semen had very little to do with glycerol penetration of the spermatozoa. Also, the dilution effect may represent instability in the phospholipid bilayer of the sperm membrane, possibly caused by movement of the surface proteins [10, 11].

REFERENCES

- [1] Al-Eissa, M.S., and Alhomida, A.S. (1997). A study of the distribution of total, free, short-chain acyl and long-chain acyl carnitine in whole blood and plasma of arabian sand gazelles (*Gazella subgutturosa marica*). *J. Comparative Haematology International*, 7 (2): 65-69.
- [2] Al-Eissa, M.S. (2007a). "Assessment of reproductive efficiency of the Arabian Sand Gazelle males (*Gazella subgutturosa marica*) and semen cryopreservation". PhD Thesis, King Saud University, College of Science, Department of Zoology.
- [3] Al-Eissa, M.S., Al-Hamidi, A.R. and Kandeal, S. (2007b). Semen cryopreservation using triladyl and tris diluents of the Arabian Sand Gazelle males (*Gazelle Subgutturosa marica*). *Arabian Gulf Journal of Scientific Research*, 25 (4):199-206.
- [4] Al-Eissa, M.S., Al-Hamidi, A.R. and Kandeal, S. (2007c). Assessment of Reproductive Efficiency of The Arabian Sand Gazelle Males (*Gazella subgutturosa marica*). *Saudi Journal of Biological Sciences*, Vol. 15, No (3), 2008.
- [5] Al-Eissa, M.S., Mohamed Abd El Kader Sandouka, Saad Hussin Alkahtani, and Ashraf Ahmed Qurtam. (2009). Reproductive characteristics of The Arabian Sand Gazelles (*Gazelle Subgutturosa marica*). *J. Saudi Soc. for Agric. Sci.*, Vol. 9, No.1a.
- [6] Alcor Life Extension Foundation. (2007). The Science of cryonics. Special ed. *Cryonics 3rd Quarter*, (28):3.
- [7] Anderson, K. and Amdal, J. (1972). Artificial Insemination with frozen semen in sheep in Norway. *World review of animal production*, 8:77-79.
- [8] Asher, G.W., Berg, D.K. and Evans, G. (2000). Storage of semen and artificial insemination in deer. *Review. Anim. Reprod. Sci.*, 62 (1-3):195-211.
- [9] Bruce, E. Eilts. (2005). Theoretical aspects of canine semen cryopreservation. *Theriogenology*, 64: 692–697.
- [10] Crabo, Bo. G., DVM. (2001). Physiological Aspects of Stallion Semen Cryopreservation. *IVIS*, 47:292, proceedings.
- [11] Foote, R.H. (2001). The History of Artificial Insemination: Selected notes and notables. *American Society of Animal Science*.
- [12] Garde, J., Soler, A.J., Cassinello, J., Crespo, C., Malo, A.F., Espeso, G., Gomendio, M. and Roldan, E.R.S. (2003). Sperm Cryopreservation in three species of endangered gazelles (*Gazella cuvieri*, *G. dama mhor*, and *G. dorcas neglecta*). *Biology of reproduction*, 69:602-611.
- [13] Garde, J., Gomendio, M., Espeso, G. and Roldan, E.R. (2006). Live birth of Mohor Gazelle (*Gazella dama mhor*) calf following intrauterine insemination of mother with frozen- thawed semen. *Reproduction Fertility and Development*.18.1/2,218.
- [14] Haque, M.N., Smith, T.R. (1996). Reintroduction of Arabian sand gazelle (*Gazella subgutturosa marica*) in Saudi Arabia. *Biological Conservation*, Volume (76): 2,203-207(5).
- [15] Holt, W.V. (2000). Fundamental aspects of sperm cryobiology: the importance of species and individual differences. *Theriogenology*, 53:47-58.
- [16] Joseph, S., Gacitua, H., King, R. and Arav, A. (2006). Post-mortem semen cryopreservation and characterization in two different endangered gazelle species (*Gazella gazella* and *Gazella dorcas*) and one subspecies (*Gazella gazella acaiae*). *Theriogenology*, 66, 4, 775-784.
- [17] Mazur, P., Leibo, S.P., and Chu, E.H.Y. (1972). A two-factor hypothesis of freezing injury: evidence from Chinese hamster tissue-culture cells. *Exp. Cell Res.*, 71:345-355.
- [18] Monfort, S.L., Asher, G.W., Wildt, D.E., Wood, T.C., Schiewe, M.C., Williamson, L.R., Bush, M. and Rall, W.F. (1993). Successful intrauterine insemination in Eld's deer (*Cervus eldi thamin*) with frozen-thawed spermatozoa. *Journal of Reproduction and Fertility*, 99:459-465.

- [19] Morrow, C.J., Penfold, L.M., Wolfe, B.A. (2009). Artificial insemination in deer and non-domestic bovids. *Theriogenology*, 71:149–165.
- [20] Polge, C. Sperm freezing: Past, present and future. In Johnson LA, Larsson K, eds, (1985). *Deep freezing of boar semen*, Beltsville, MD: USDA,.167–173.
- [21] Purdy, P.H. (2005). A review on goat sperm cryopreservation. *small Ruminant Research*, 63:215-225.
- [22] Rapatz, G.L. (1966). What happens when semen is frozen?. *Proc.1st Tech.Conf. on A.I. and Bovine Reprod.*, Assoc.Anim. Breeders, Columbia, MO,.45.
- [23] Salamon, S. and Visser, D. (1972). Effect of composition of tris-based diluent and of thawing solution on survival of ram spermatozoa frozen by the pellet method . *Aust J Biol Sci.*, 25(3):605-18.
- [24] Sherman, J.K. (1978). Banks for frozen human semen: Current status and prospects. In *The Integrity of Frozen Spermatozoa*. Washington, DC: The National Research Council, 78–91.