

BCH 462

**Sodium Dodecyl Sulfate -PolyAcryl amide Gel
Electrophoresis
[SDS-PAGE]**



Objectives:

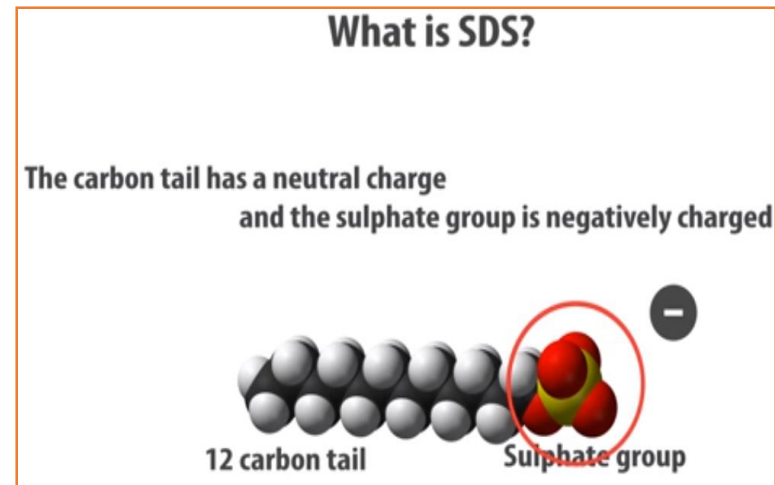
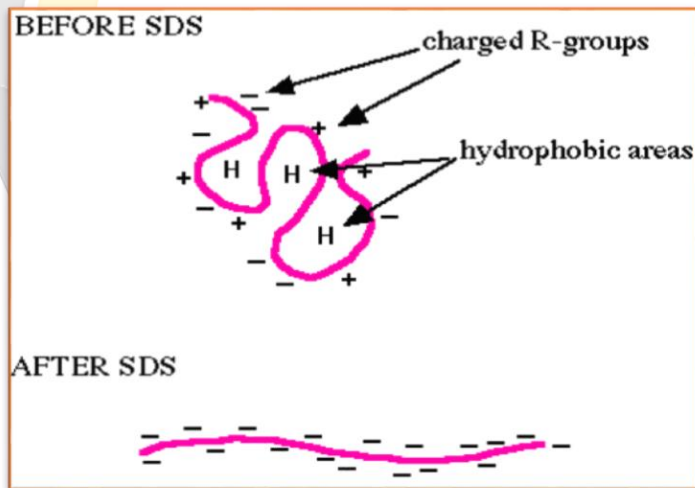
- Separation of bacterial proteins using SDS-PAGE.

SDS-Polyacrylamide Gel Electrophoresis

- Sodium Dodecyl Sulfate-Polyacrylamide gel Electrophoresis (SDS-PAGE), is a technique widely used in biochemistry ,forensics, genetics and molecular biology to separate and identify proteins according to their molecular weight.
- This method separates proteins based primarily on their molecular weights.

Principle:

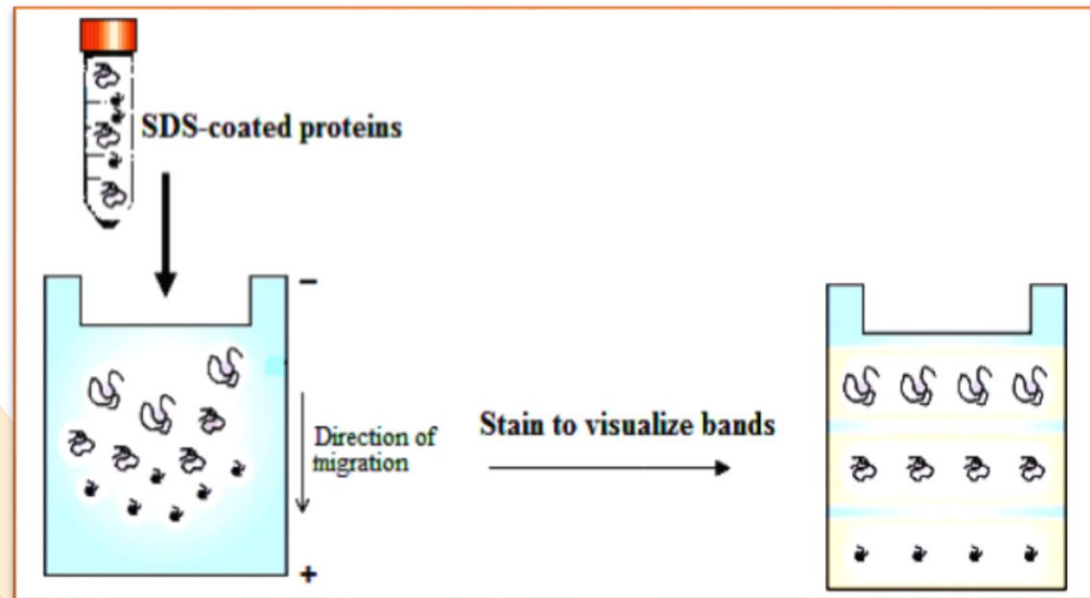
-Sodium Dodecyl Sulfate [SDS]: is a detergent which denature proteins by binding to the hydrophobic regions, **all non-covalent bonds will disrupted** and the **proteins acquire a negative net charge**.



-Treatment with a disulfide reducing agent such as β -mercaptoethanol or DTT (dithiothreitol), which further denatures the proteins **by reducing disulfide linkages**, thus overcoming some forms of tertiary protein folding, and breaking up quaternary protein structure

So, the proteins samples are having uniformed structure and charge(-ve) → the separation will depend on their molecular weight only.

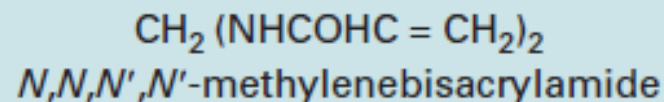
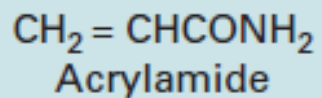
-Small proteins migrate faster through the gel under the influence of the applied electric field, whereas large proteins are successively retarded, due to the sieving effect of the gels



<http://www.youtube.com/watch?v=3CrzY7jb9fQ>

Polyacrylamide gel (Acrylamide stock):

- The polyacrylamide gel is formed by co-polymerization of acrylamide and a cross-linking By N,N'-methylene-bis-acrylamide "bis-acrylamide".
- To polymerize the gel a system, ammonium persulfate (**initiator**) and tetramethylene ethylene diamine (TEMED) [**catalyst**], are added.



+
Free radical
catalyst

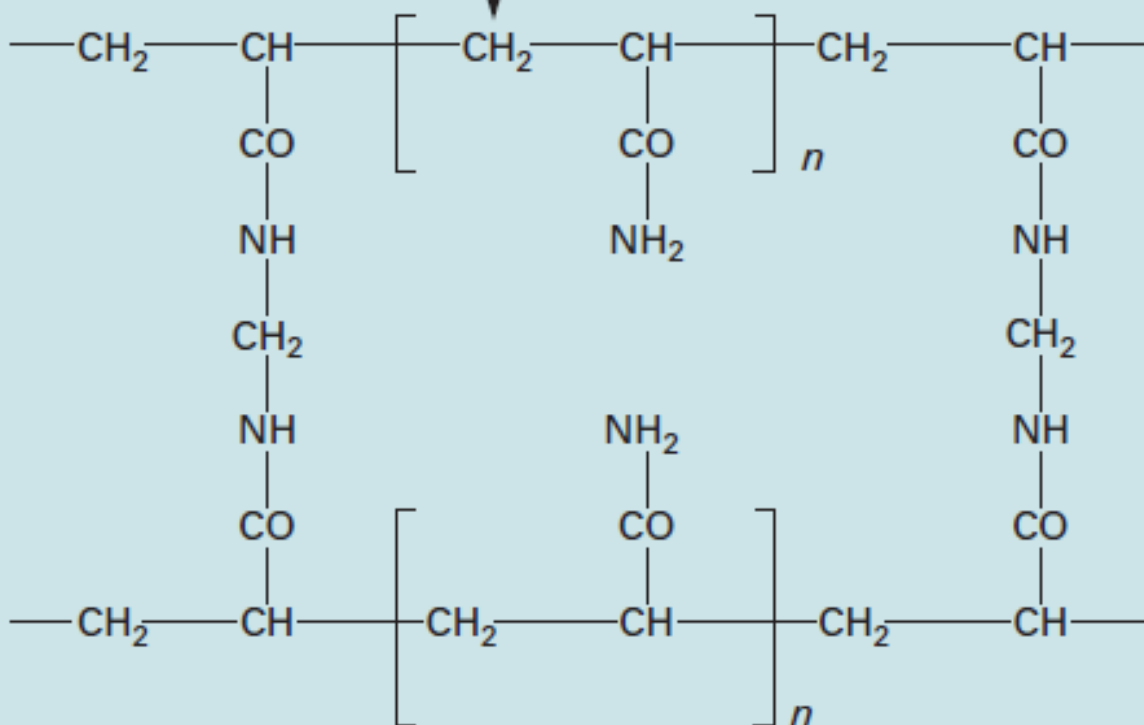
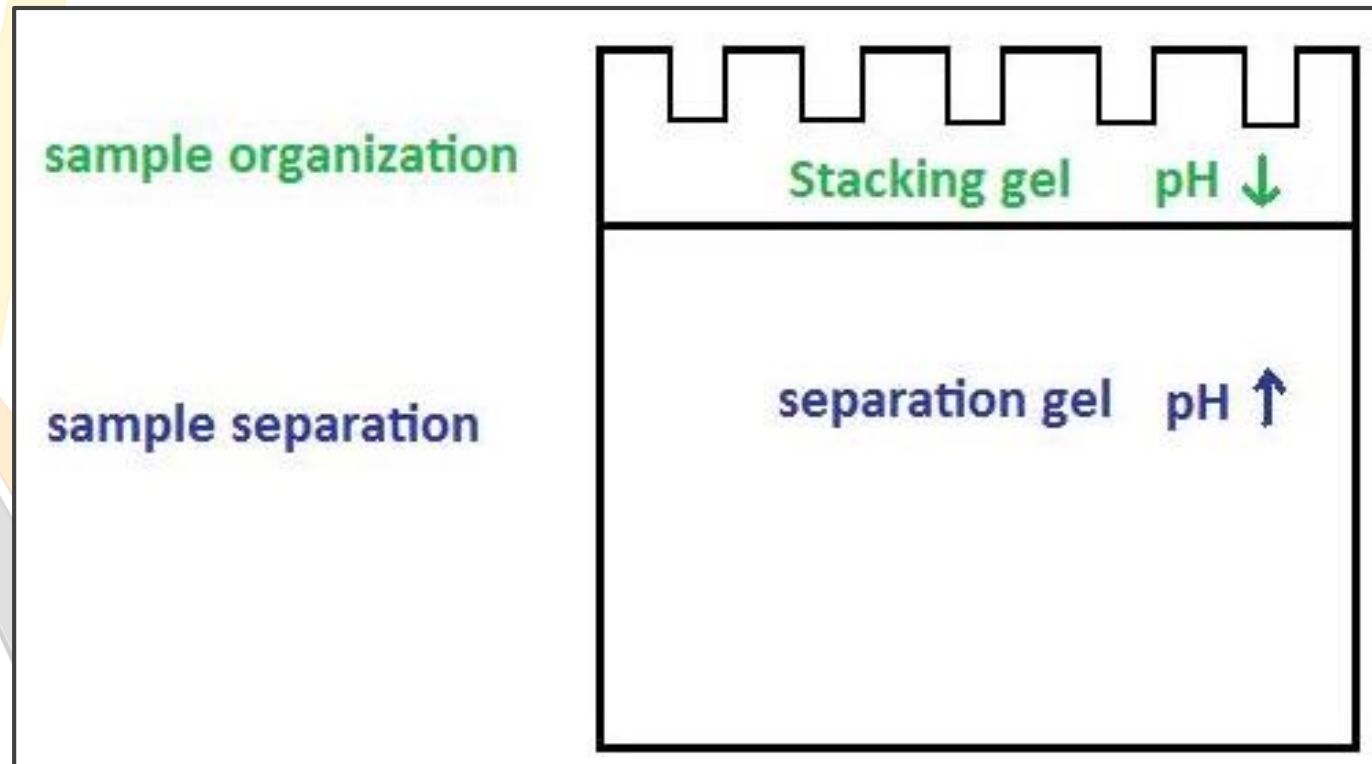


Fig. 10.5 The formation of a polyacrylamide gel from acrylamide and bis-acrylamide.



SDS-Polyacrylamide Gel Electrophoresis

SDS-Polyacrylamide Gel Electrophoresis preparations:

1-Sample Preparation:

-40µl of protein sample + 10 µl of disruption buffer → boil the mixture 3minets at 99°C.

-Disruption buffer [loading buffer] contain:

- | | |
|----------------------|-----|
| -10% (w/v) SDS | [?] |
| -1M Tris/HCl, pH 6.8 | |
| -Glycerol | [?] |
| -β-Mercaptoethanol | [?] |
| -Bromophenol blue | [?] |

2- Polyacrylamide Gel Preparation :

Acrylamide stock should be prepared first :

-Cross-linked polyacrylamide gels are formed from the polymerisation of acrylamide monomer in the presence of smaller amounts of N,N'-methylene-bisacrylamide (some time referred to as 'bis'-acrylamide).

A-Separation gel preparation:

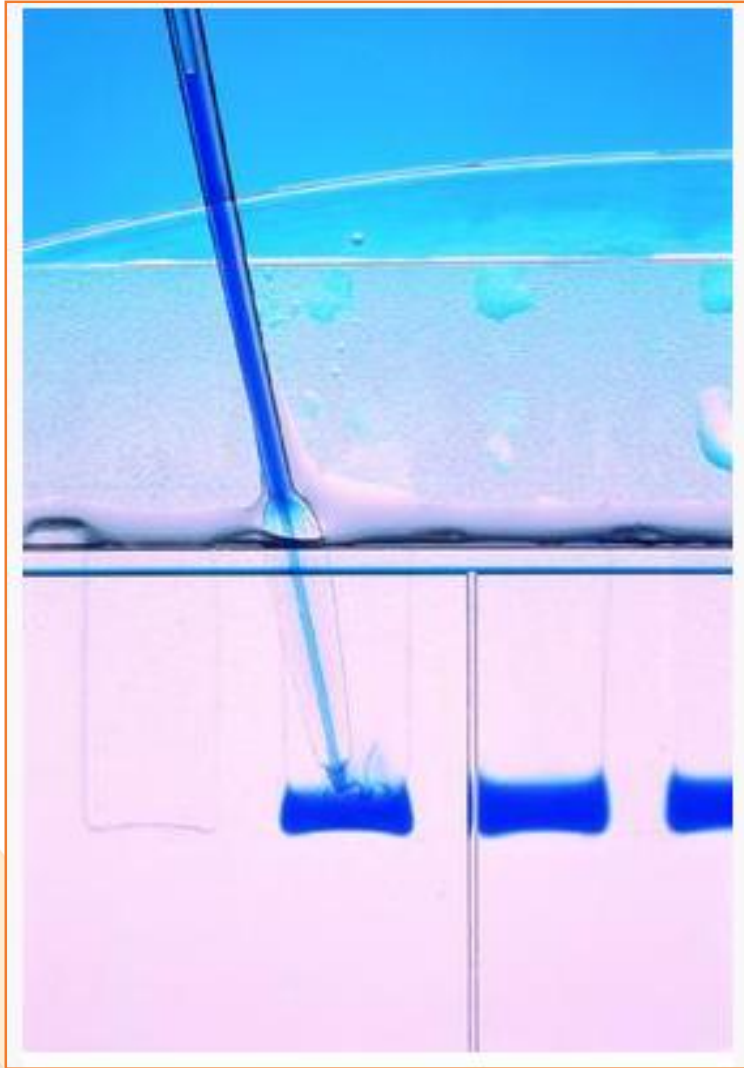
Stock solutions	Volume of stock solution required to make 12% polyacrylamide gel
1.5 M Tris/HCl, <u>pH 8.8</u>	2.0 ml
<u>Acrylamide stock</u>	<u>3.2 ml</u>
Water	2.8 ml
10% SDS	80 μ l
<u>10% Ammonium persulphate (fresh)</u>	100 μ l
<u>TEMED</u>	10 μ l

B-Stacking gel preparation:

Stock solutions	Volume of stock solution required to make 7% polyacrylamide gel
0.5M Tris/HCl, pH6.8	1.0 ml
<u>Acrylamide stock</u>	<u>1.0 ml</u>
Water	3.0 ml
10% SDS	80 µl
<u>10% Ammonium persulphate (fresh)</u>	50 µl
<u>TEMED</u>	5 µl

http://www.youtube.com/watch?v=EDi_n_0NiF4

3-Loading the samples:



4-Running the gel using , Running buffer 1x pH 8.3:

It is contain:

- Tris-HCl .
- Glycine.
- SDS.

5- Stain the gel using staining buffer :

It is contain:

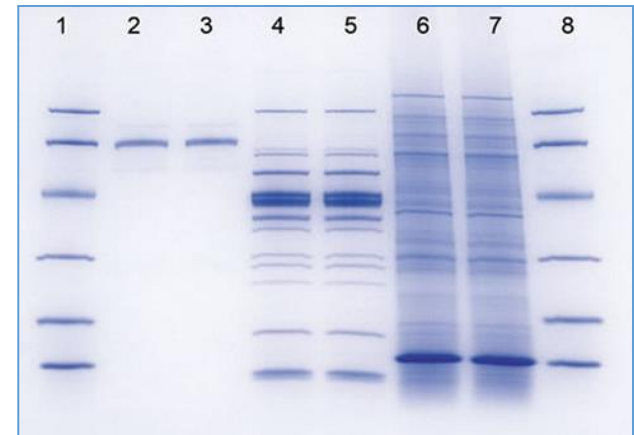
- Glacial acetic acid
- Methanol
- Coomassie brilliant blue 250-R**



6- De-stain the gel using De-staining buffer:

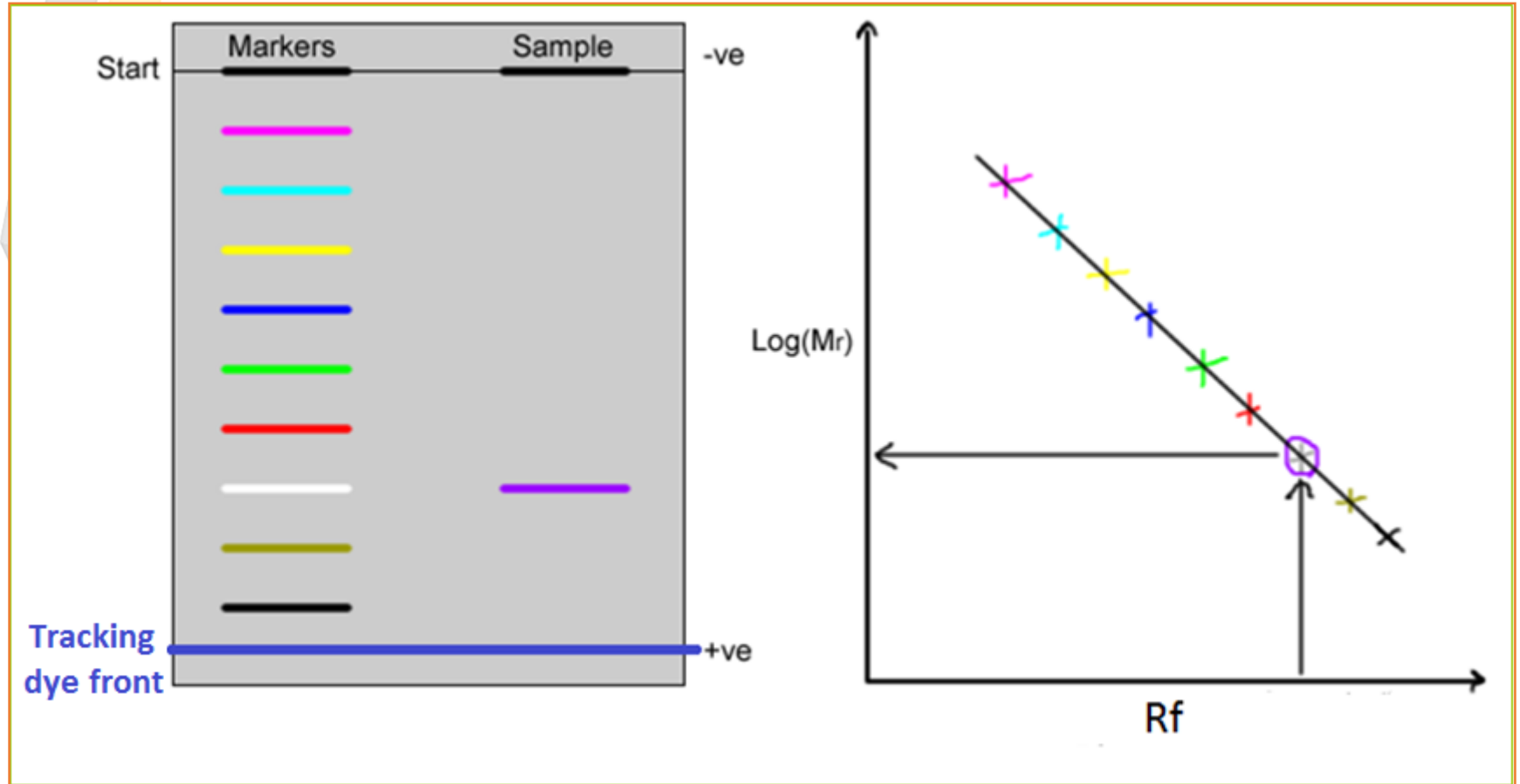
It is contain:

- Glacial acetic acid
- Methanol

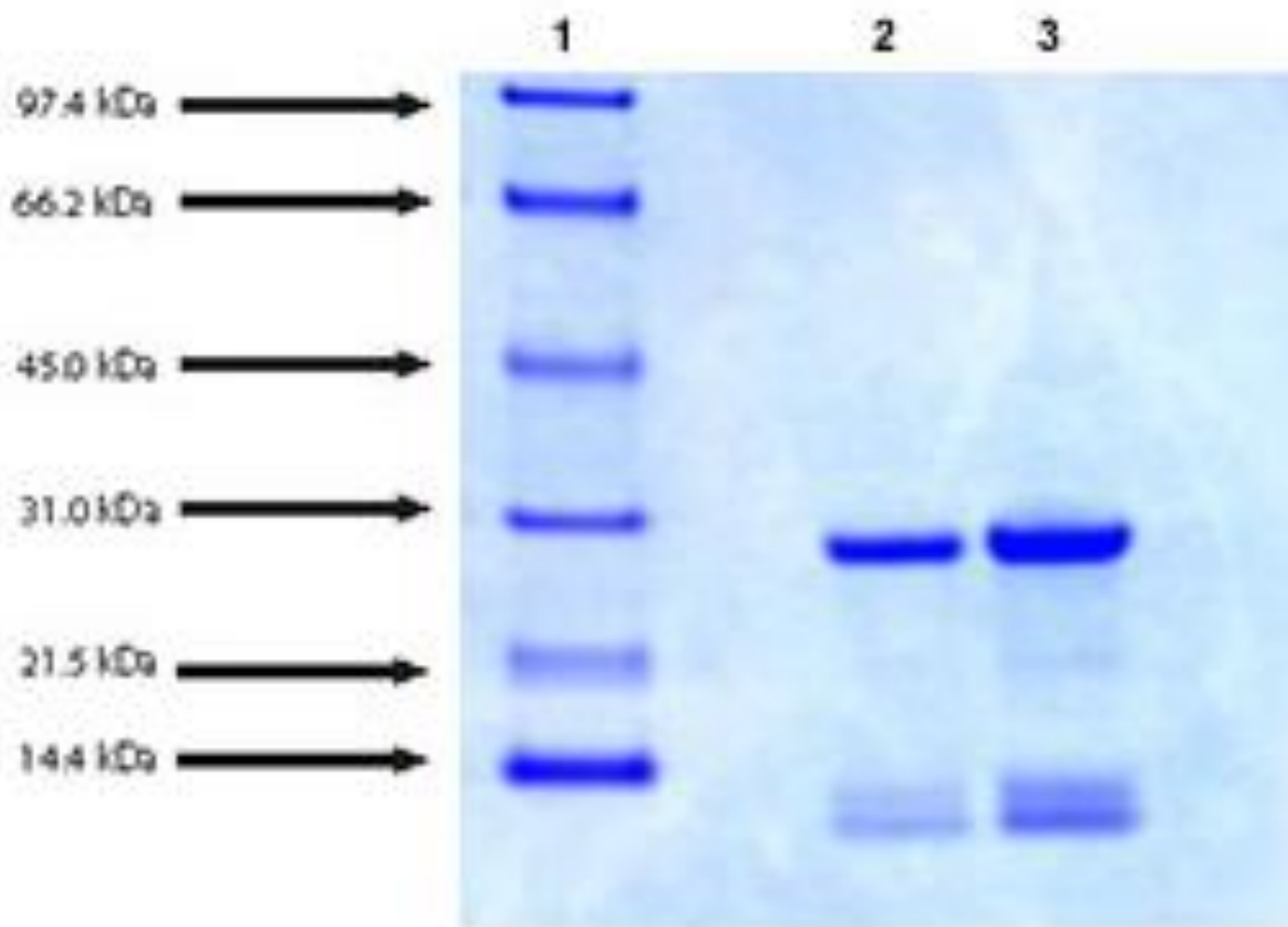


7-Analysis:

For Molecular weight Determination.



$$- R_f = \frac{\text{Distance of migration of sample}}{\text{Distance moved by tracking dye}}$$



Applications:

1. To detect the purity of the protein.
2. Determine of protein molecular weight.
3. Determine the presence of certain protein.