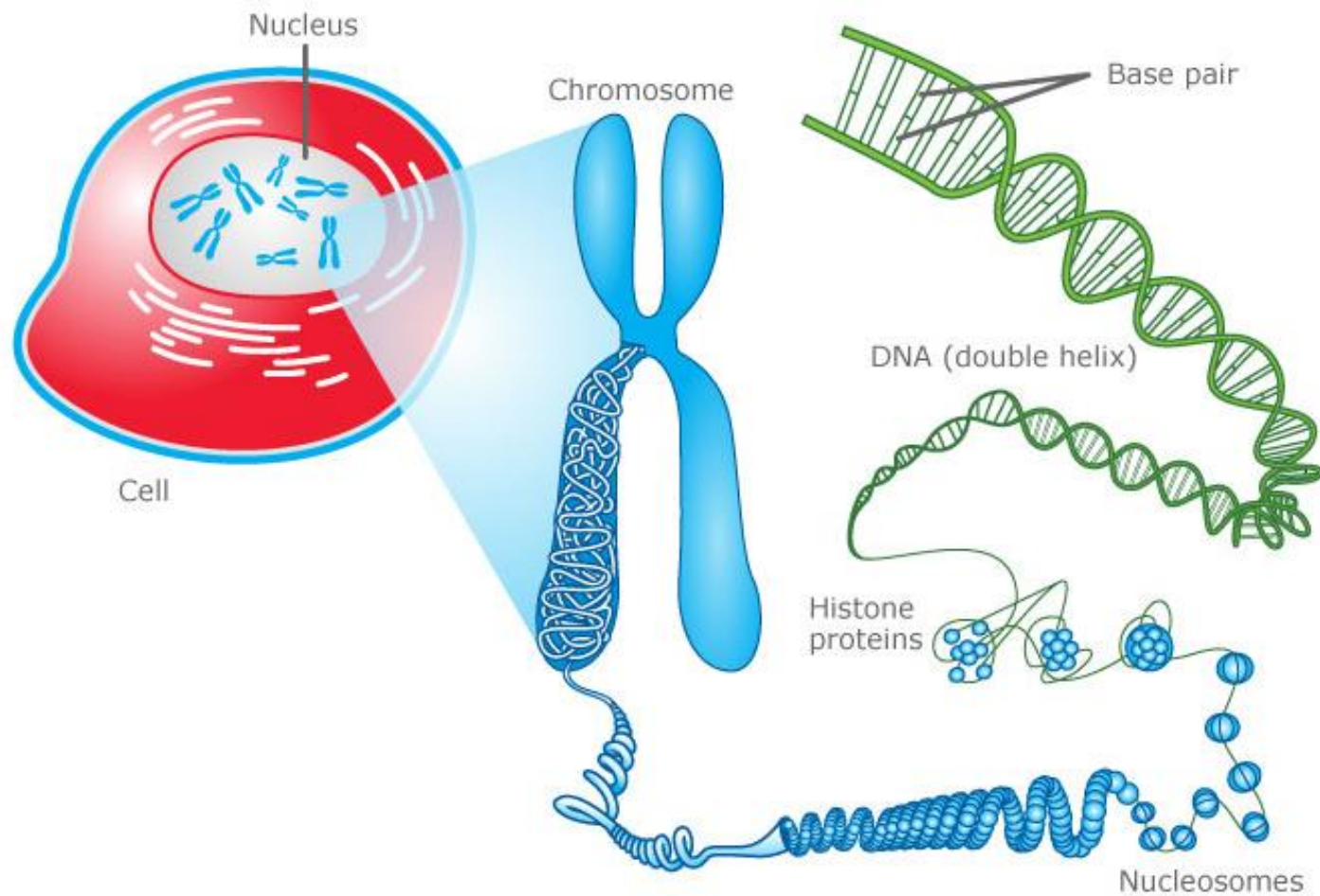


# **SPECTRAL CHARACTERIZATION OF DNA**

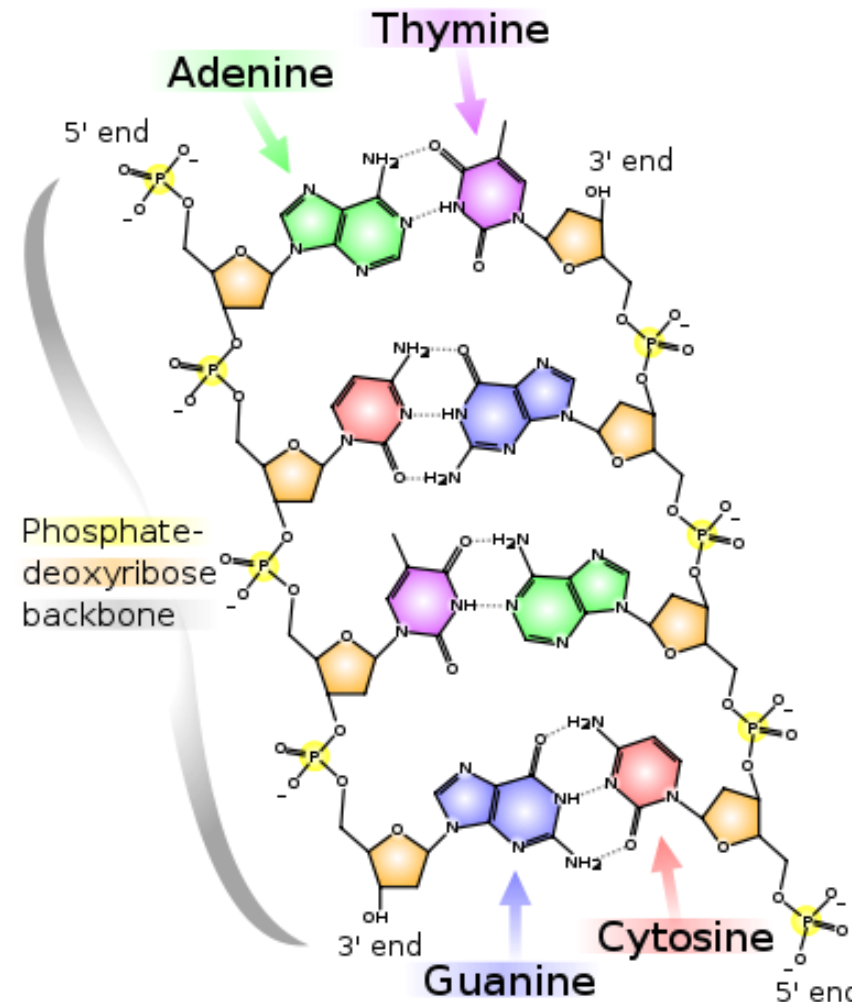
# DNA

## 'DEOXY RIBONUCLEIC ACID'

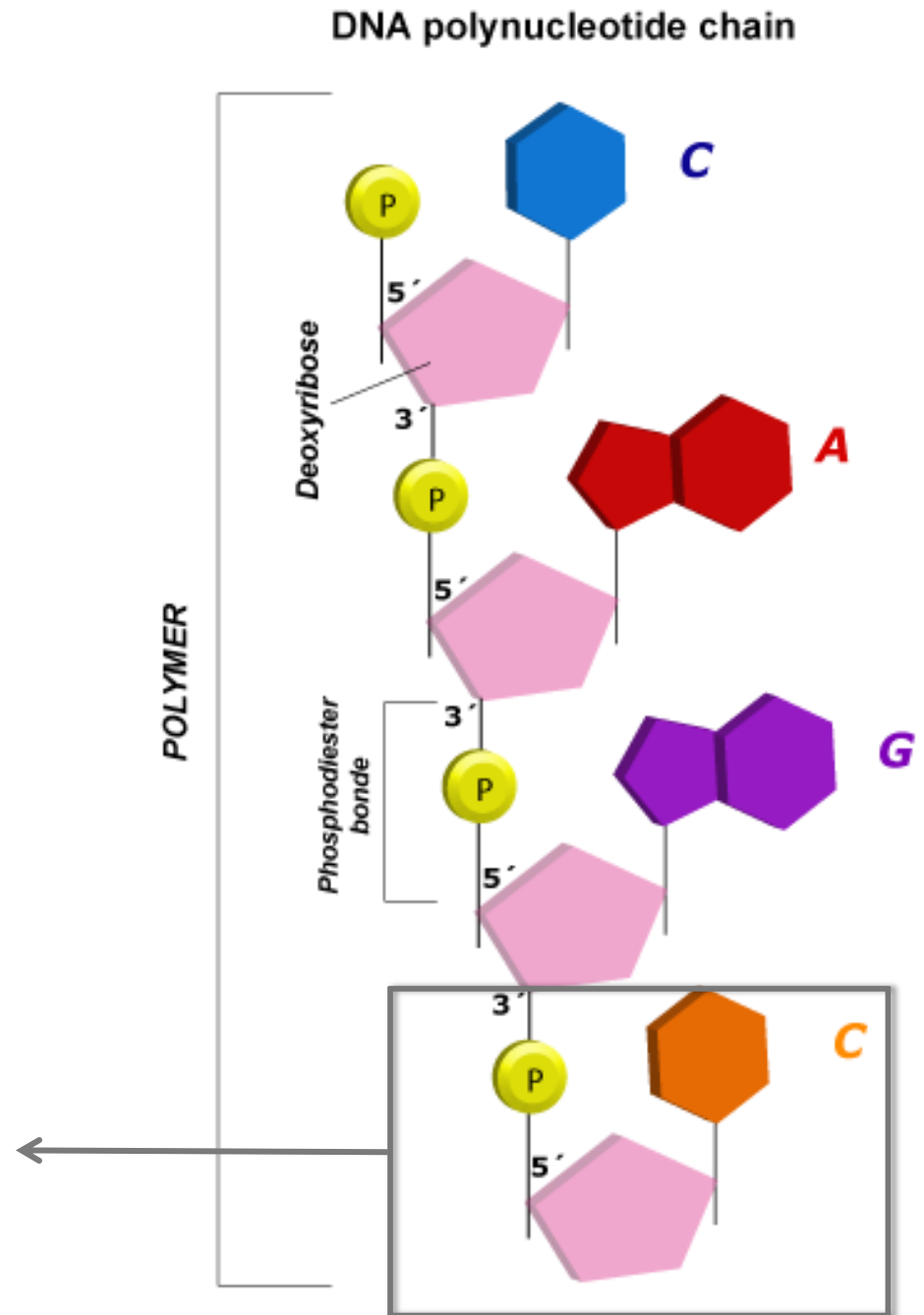
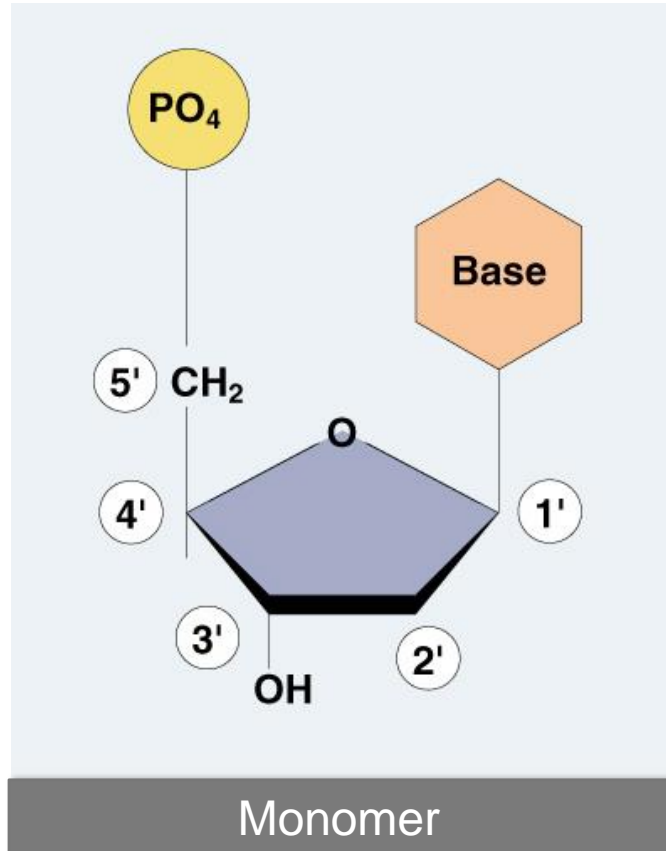


# DEOXY RIBONUCLEIC ACID (DNA)

- DNA is made of **2 polynucleotide chains** which run in opposite direction.”antiparallel ”
- DNA has a **double helical structure.**
- Each polynucleotide chain of DNA consists of monomer units.
- A monomer unit consists of 3 main components that are:
  1. **A sugar,**
  2. **a phosphate,**
  3. **a nitrogenous base.**



# NUCLEOTIDE



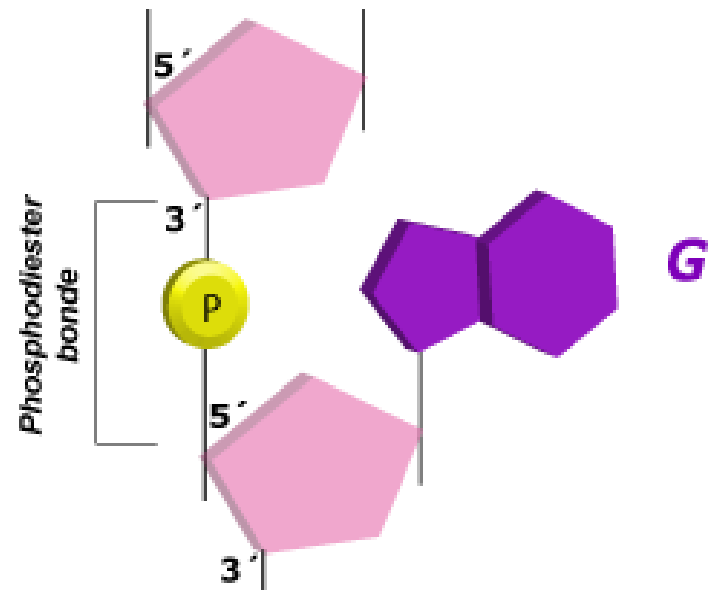
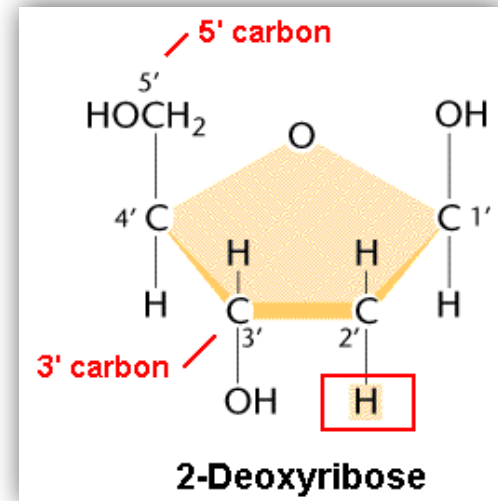
# DNA STRUCTURE

## 1. Deoxyribose sugar:

- Is a monosaccharide 5-Carbon Sugar, Its name indicates that it is a deoxy sugar, meaning that →  
[ it is derived from the sugar ribose by loss of an oxygen atom ].

## 2. Phosphate Group:

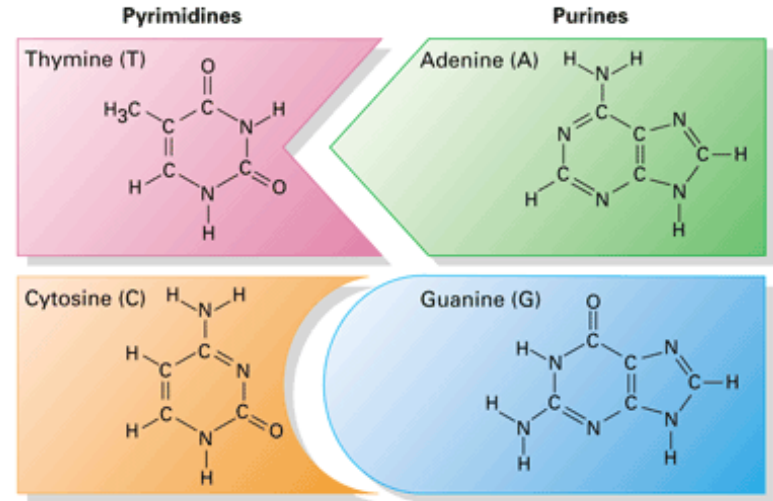
- The sugars are joined together by phosphate groups that form phosphodiester bonds between the third and fifth carbon atoms of adjacent sugar rings.



# DNA STRUCTURE

## 3. Nitrogenous bases:

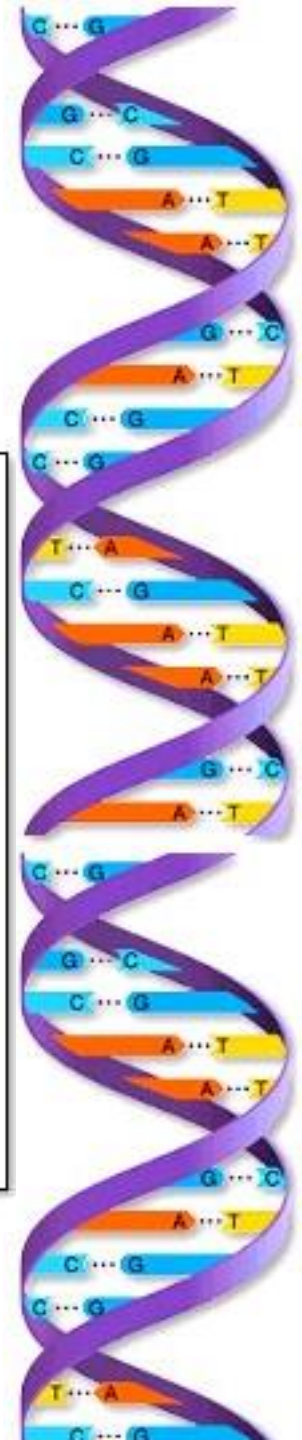
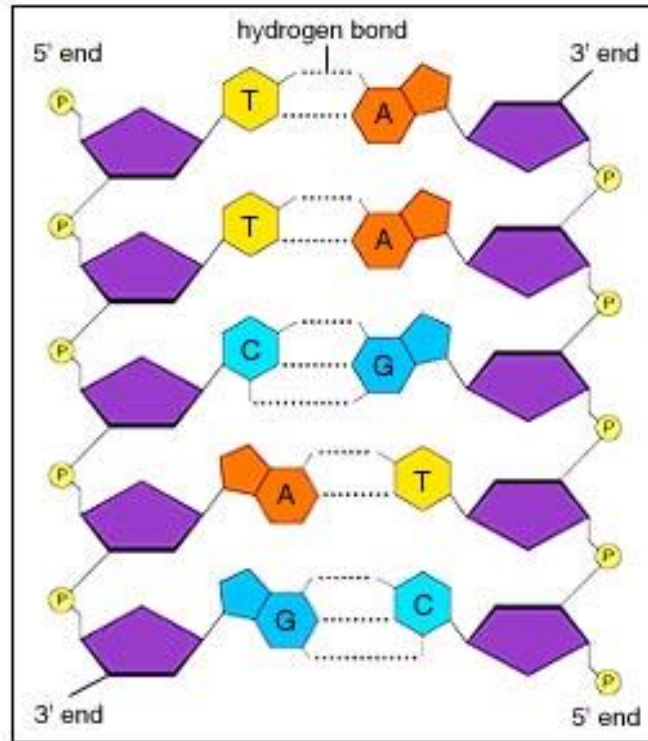
- is a nitrogen-containing organic molecule having the chemical properties of a base
- They are classified as the derivatives of two parent compounds,
  1. Purine.
    - [ Adenine, Guanine ]
  2. Pyrimidine.
    - [ Cytosine, Thymine ]



# DNA STRUCTURE

## 4. Hydrogen bond:

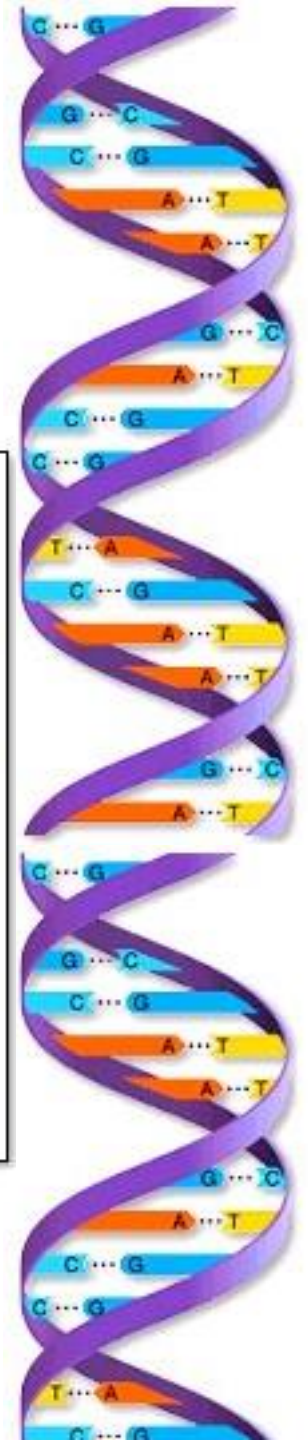
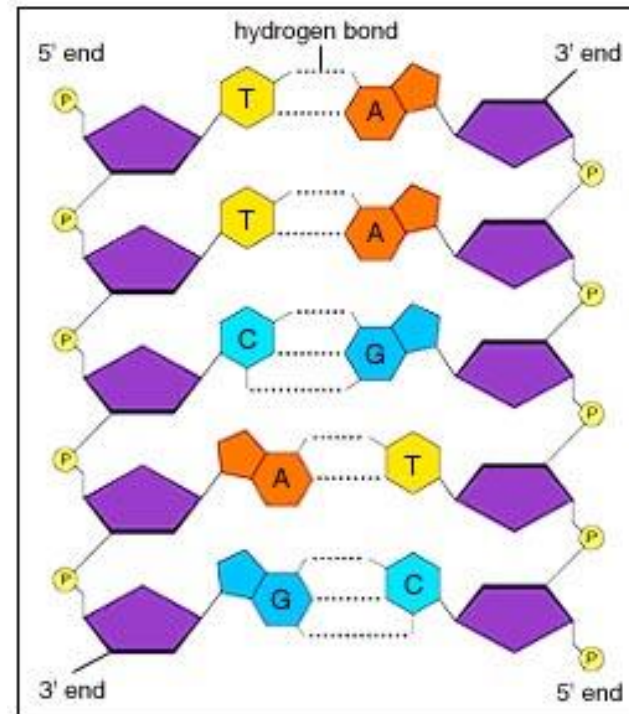
- The H-bonds form between base pairs of the antiparallel strands.
- The base in the first strand forms an H-bond only with a **complementary** base in the second strand.
- Those two bases form a *base-pair* (H-bond interaction that keeps strands together and form double helical structure).





# DNA STRUCTURE

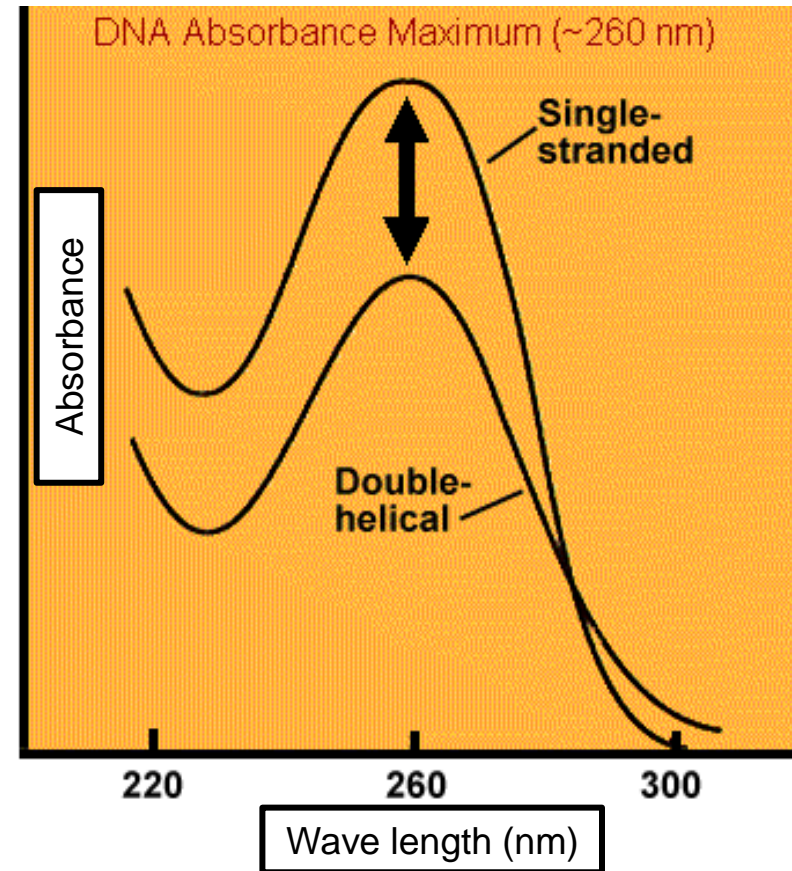
- The hydrophobic bases are inside the double helix of DNA, give the **hydrophobic effect to stabilize the double helix**.
- while **sugars and phosphates** are located **outside** of the double helical structure.





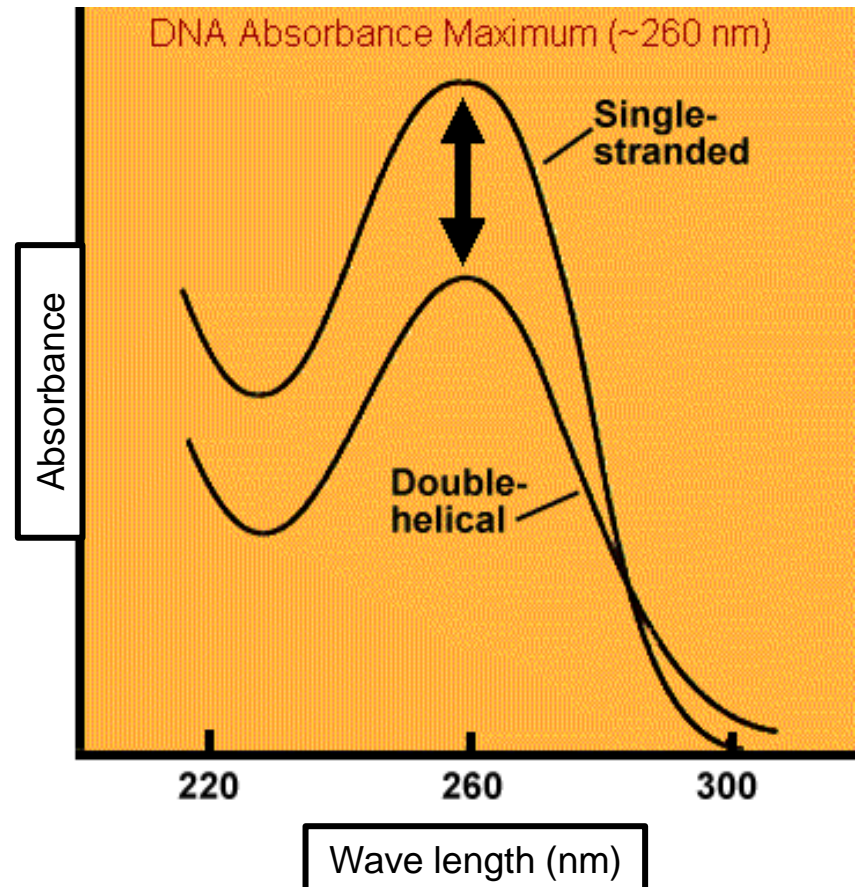
# OPTICAL DENSITY OF DNA

- **Nucleic acid** would be expected to have maximum **absorbance** at 260.
- It absorbs at this wavelength because of the nitrogenous bases (A, G, C and T) of DNA.
- In a spectrophotometer, a sample is exposed to ultraviolet light at **260 nm**, and a photo-detector measures the light that passes through the sample.



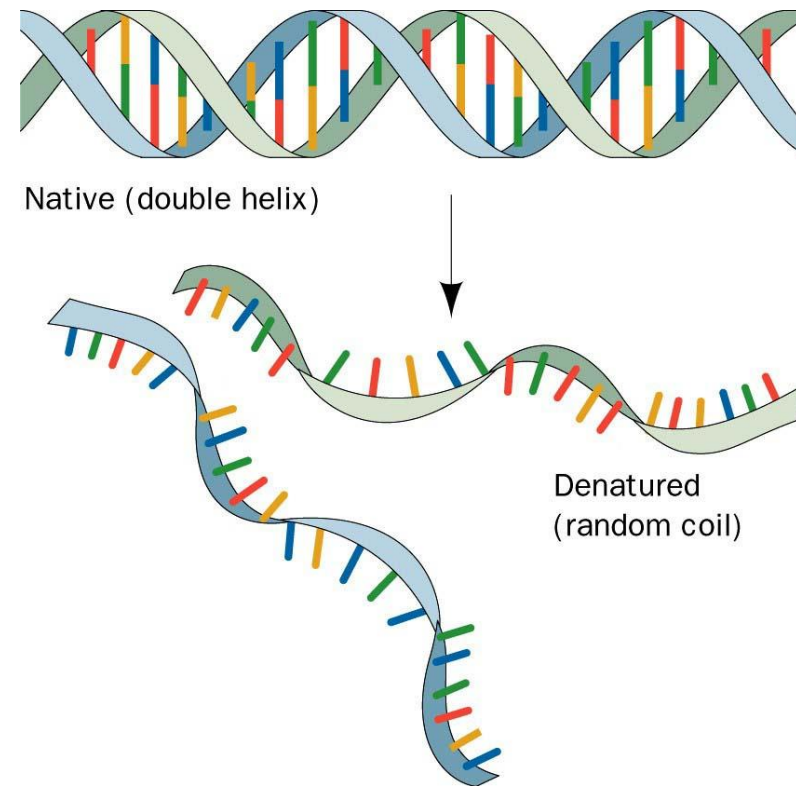
# HYPERCHROMICITY

- The **increase of absorbance** (*optical density*) of a material.
- The most famous example is the **hyperchromicity of DNA** that occurs when the DNA duplex is denatured.
- The opposite, a decrease of absorbance is called **hypochromicity**.



# DENATURATION OF DNA

- Many different substances or environmental conditions can denature DNA, such as:
  - **strong acids, organic solvent**
  - **heating**
  - **Exposure to Radiation/ UV light**



# SPECTRAL CHARACTERIZATION OF YEAST DNA

## Objective:

- To establish the wave length that represent the maximum absorbance for DNA.
- To establish the hyperchromic effect on DNA.

## Principle:

- The double helix of DNA are bound together mainly by hydrogen bonds and hydrophobic effect between the complementary bases. .
- When DNA in solution is heated above its **melting temperature** (usually more than 80 °C), the double-stranded DNA unwinds to form single-stranded DNA.

# SPECTRAL CHARACTERIZATION OF YEAST DNA

## Principle:

- In single stranded DNA the bases become unstacked and can thus absorb more light.
- In their native state, the bases of DNA absorb light in the 260-nm wavelength region.
- When the bases become unstacked, the wavelength of maximum absorbance does not change, but the amount absorbed increases by 30-40%.
- a double strand DNA dissociating to single strands produces a sharp cooperative transition.

# **SPECTRAL CHARACTERIZATION OF YEAST DNA**

## **Materials:**

- **DNA concentrated sample( extracted from yeast).**
- **1X saline solution ( NaCl with Tri Sodium Citrate).**
- **Quartz Cuvtte.**
- **Spectrophotometer.**

# SPECTRAL CHARACTERIZATION OF YEAST DNA

## Method:

- **Set and label 6 test tube : D1, D2, D3,D4,D5,D6**
  - ✓ 1. In D1 pipette 0.5 ml of isolated DNA (extracted from Yeast) and add to it 4.5ml of 1X saline-citrate. Mix it very well.
- **Measure the absorbance of D1 at 260nm if it is  $> 3$  :**
  - ✓ 2. In D2 pipette **0.5 ml of D1** ,add to it 4.5ml of 1X saline-citrate. Mix it very well.
- **Measure the absorbance of D2 (if the absorbance is greater than 1,dilute the solution until you obtain A<sub>260</sub> of 1 or slightly less).**



# SPECTRAL CHARACTERIZATION OF YEAST DNA

## Method:

- When the absorbance of solution ( $A_{260} \approx 1.0$ ) is obtained read the absorbance of the solution at the following wave lengths:  
**(240,245,250,255,260,265,270,275,280)**
- **using 1X saline as a blank.**

# SPECTRAL CHARACTERIZATION OF YEAST DNA

## Method:

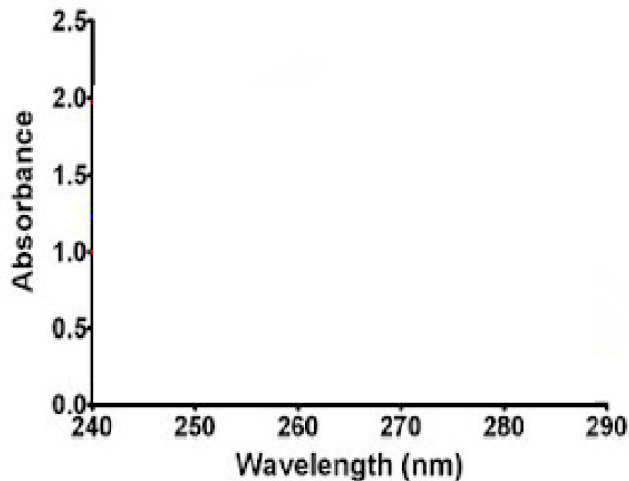
- Now take the dilution tube which give an absorbance=1 ,cover the tube and put it in boiling water bath for 15 min
- **Immediately measure the absorbance at the following wave lengths:**  
**(240,245,250,255,260,265,270,275,280)**
- using 1X saline as a blank.

## EXPERIMENT OF DAY

# SPECTRAL CHARACTERIZATION OF YEAST DNA

## Results:

- ✓ Plot The absorption spectra of the native DNA solution and the denatured DNA against wave lengths.
- ✓ Record Your result and write your comment in the discussion.



Wave length (nm)	Absorbance of isolated DNA	Absorbance of heated DNA
240		
245		
250		
255		
260		
265		
270		
275		
280		