

## ORGANOCHLORINE RESIDUES IN CUTTLEFISH FROM THE ARABIAN GULF

A. El-Gendy, S. Al-Farraj, S. Al Kahtani, M. El-Hedeny<sup>2</sup>

<sup>1</sup>Department of Zoology, Faculty of Science, Alexandria University, Alexandria (EGYPT)

<sup>2</sup>Department of Sciences, College of Teachers, King Saud University,  
Riyadh (KINGDOM OF SAUDI ARABIA)

E-mails: mmelhedeny@gmail.com, melhedeny@ksu.edu.sa

### ABSTRACT

Contaminations of persistent organic pollutants (POPs) such as, dichlorodiphenyl trichloroethane (DDT), hexachlorocyclohexane (HCH) and chlordane (CHLs) were examined in the edible mantle tissues of the commercial cuttlefish *Sepia pharaonis* Ehrenberg 1831, collected from the marine water of the Arabian Gulf. The mean concentrations of DDT, CHLs and HCH were in the ranges of 29.4 - 56 ng/g, 47.4 - 100 ng/g and 1 - 4 ng/g, respectively. Among the POPs analyzed, HCH showed the lowest concentrations ranging between 1 to 5 ng/g lipid wt. However, concentrations of DDT, CHLs and HCH, detected in this study, were generally comparable or lower than those found in studies of similar cephalopod species from other areas subject to a high anthropogenic impact. Relationships between total body lengths and/or dorsal mantle lengths of the organisms and the concentration values of the studied POPs were also considered. Compared with recommendations of the international organizations, there are no potential risks associated with consuming the studied cuttlefish species.

**Key words:** cuttlefish, *Sepia pharaonis*, organochlorine, DDT, CHLs, HCH, Arabian Gulf

### 1. INTRODUCTION

Organochlorinated substances are listed as Persistent Organic Pesticides (POPs) by the United Nations Environment Program (UNEP) in the 1995 Stockholm Convention. Twelve POPs were identified of which, nine are pesticides (aldrin, endrin, dieldrin, heptachlor, chlordane (CHL), mirex, toxaphene, Dichlorodiphenyltrichloroethane (DDT) and hexa chlorobenzene (HCB) (Wei et al. 2007). Although the usage of technical different types of POPs in several countries as Japan, United States and the former Soviet Union were officially banned more than 20 years ago, continuing usage of these compounds for agricultural or public health purpose in these countries have been reported (Nakata et al., 1995; Depeng et al., 1996; McConnell et al., 1996; MAFFJ., 2001). In the last two decades, POPs have shifted from industrialized countries of Northern Hemisphere to developing countries in tropical and sub-tropical regions including India and China (Wong et al., 2005). Many countries have now implemented the Stockholm Convention on POPs (2001), which suggests ending commercial use of 12 POPs and reducing or eliminating their emission into the environment.

POPs are highly persistent, they may have a half-life of years or decades in soil/sediment and several days in the atmosphere (Jones & Voogt, 1999). In the aquatic environment, they partition strongly to organic phase and accumulate in the lipid rich tissues in animals and become soluble in fatty tissues, rather than entering into the aqueous milieu of cells. Because of this, these chemicals are persistent in biota and biomagnify in food chains causing severe injury to the nervous, endocrine, reproductive and immune systems in birds, fish, and mammals (Ratcliffe, 1967; Metcalf, 1977; Kelce et al., 1995; Skaare et al., 2000; Toft et al., 2003).

The Arabian Gulf is represents a highly stressful environment due to a combination of both prevailing natural conditions and development pressures along its coastline (Tolosa et al, 2005). The countries that located along its coast undergone considerable development, and consequently urbanization, industrialization. The major sources of POPs in the coastal zone in the Gulf are exports of agricultural chemicals from coastal catchments, heavy manufacturing effluent discharge, municipal and industrial sewage.

Cephalopoda, one of the most important groups of marine invertebrates, are consumed throughout the world, both as food and as feed supplement and have great commercial value (Navarro & Villanueva, 2001 and Koueta & Boucaud-Camou, 2001). Demand of cephalopods, especially small ones is increasing. Indeed, they are actually much appreciated as a protein source and because of their gustative quality.

Very little studies have been established to investigate the distribution and analysis of organic and inorganic substances in cephalopod tissues (e.g. Miramand & Bentley, 1992; Butty & Holdway, 1997; Weisbrod et al., 2000, 2001; Ueno et al., 2003; Storelli et al., 2006; Won et al., 2009). According to Danisa, et al. (2005) and Won et al. (2009) more researches are needed in this field. In addition to the rare studies concerning the use of cephalopods as bioindicator in the Arabian Gulf water of the Kingdom of the Saudi Arabia, there are no researches to determine the residue levels of (POPs) in soft tissues of cephalopods there .

The broadly distributed neretic demersal cephalopod species *Sepia pharaonis* Ehenberg, 1831, is one of the most common edible cuttlefish species available in the Arabian Gulf and the Oman Sea (Tehranifard & Dastan, 2011).

In the light of this concern, our study was carried out to determine some POPs residues in mantle of the cuttlefish *Sepia pharaonis* to ascertain whether the concentrations exceed the levels fixed by legislation undertaken worldwide in order to assess human health risk. Besides, to find out if there is any relationship between the whole body length and/or the dorsal mantle length and the POPs concentration levels.

## 2. MATERIALS AND METHODS

### 1-Sampling

*Sepia pharaonis*, a common edible cuttlefish species, were caught by local professional fishermen from the Arabian Gulf water of Saudi Arabia coast (Fig. 1).



**Fig. 1.** Map of the Arabian Gulf, locality of the studied cuttlefish.

After capture, care was taken to avoid any contamination of the specimens that were immediately packed in ice and sent to the laboratory. The collected specimens were identified accordingly (Silas, 1985; Aoyama & Nguyen, 1989 and Graham, 1994). The total length (TL) and dorsal mantle length (DML) were measured (Table 1). After that, specimens were divided into 8 groups (each contains 5 animals) according to the dorsal mantle length. The collected animals were dissected and the edible mantle, the contaminant storage tissues, were obtained and homogenized. All the homogenized samples were stored at -20 C until chemical analysis was performed.

**Table 1.** Total Length (TL), Dorsal mantle length (DML), mean concentrations, standard deviation (S.D.) and standard error (S.E.) for the eight groups of the edible cuttlefish *Sepia pharaonis* collected from the Arabian Gulf of Saudi Arabia.

Group	TL	DML	CHL			HCH			DDT		
			Mean	S.D.	S.E.	Mean	S.D.	S.E.	Mean	S.D.	S.E.
1	10	6	80	24.49	10.95	1.8	0.83	0.37	55	13.22	5.91
2	11	7	75	17.16	7.67	3	1.58	0.70	40.4	11.10	4.96
3	13	8	70.6	19.04	8.51	1	1.00	0.44	29.4	9.52	4.26
4	14.5	9	89.2	14.25	6.37	5	1.58	0.70	56	15.57	6.96
5	16	10	47.4	10.85	4.85	1	1.00	0.44	40.4	11.10	4.96
6	17	11	100	12.24	5.47	2	1.22	0.54	52	10.81	4.83
7	17.5	12	71.6	18.78	8.40	4	1.58	0.70	44	17.10	7.64
8	19	13	55	13.22	5.91	3	1.58	0.70	48	15.65	7.00

### 2-Chemical analysis

Organochlorine insecticides, chlordane compounds (CHLs) and hexachlorobenzene (HCB) were analyzed using a Hewlett Packard HP5890- GC with split/splitless injector and a 25 m x 0.3 mm fused silica capillary with a chemically bonded gum phase SE54 sodium chloride and sodium sulfate that were kiln fired at 450° C overnight and cooled in a greaseless desiccator. Silica gel, used for column chromatography, was solvent extracted with n-hexane in a flask cartridge inserted into an extraction apparatus, as described by Ehrhardt (1987).

After extraction, the silica gel was first dried in the same cartridge by passing ultra-pure nitrogen through; and was then activated by heating the cartridge in an electric tube oven to 200° C for 6 h with nitrogen stream reduced to a few ml per minute

The extraction method was based upon that Wade et al., (1988). A total 5g of dried tissues was Soxhlet-extracted with methyl chloride and concentrated in Kuderna-Danish tubes. The extracts were fractionated by alumina: silic gel (80-100 mesh) chromatography. The extracts were sequentially eluted from the column with 50 ml of pentane (aliphatic fraction) and 200 ml of 1:1 pentane-dichloromethane (POPs fraction) and concentrated for GC

analysis. POPs were chromatography in the split/splitless mode, using an electron capture detector (ECD). A 30 m x 0.32 mm i.d. fused-silica capillary column was used for this purpose with a chemically bonded gum phase SE54 (J&W Scientific, Inc.).

### 3-Recovery and Quality assurance

Recovery standards were added at the start of the procedure and carried through the extraction, cleanup, and instrumental analysis steps to determine the recovery of the analysis. The recoveries of DDT, CHLs and HCH in this analytical procedure were 94.5-7.6% for the studied pesticides.

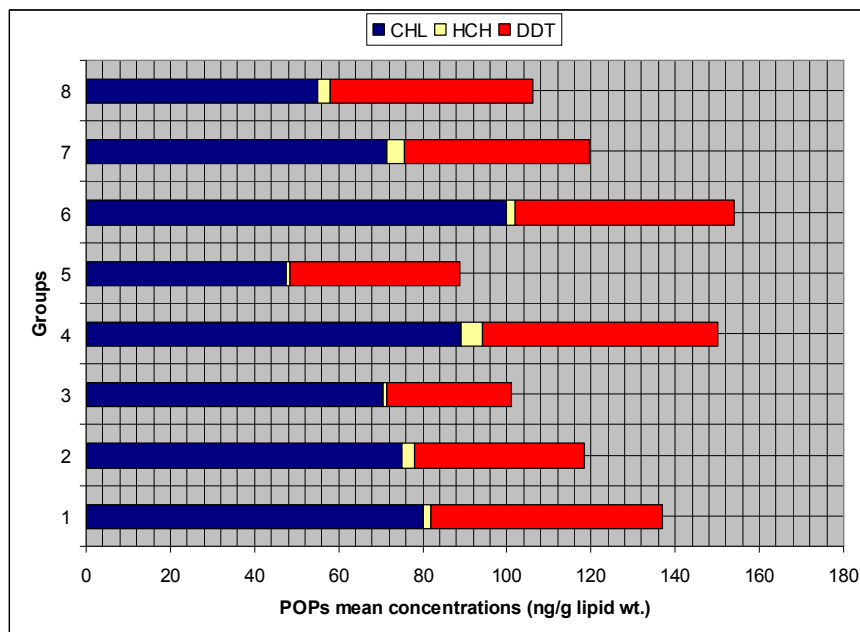
The specific quality assurance steps were used to ensure measurement accuracy and precision: one procedural blank, one matrix spike, one duplicate sample and one standard reference material were run with each batch. Quality assurance for the measurement of POCs was confirmed by analyzing Standard Reference Materials and the results agreed well with the certified values. Concentrations of POCs were not corrected for the recoveries and were given as nanograms per gram on a lipid weight basis.

### 4-Statistical analysis

An independent t test analysis was applied to the observed data to check the level of significance site-wise. Pearson's Correlation Coefficient Statistical functions on Microsoft Excel were used to test the relation between the body length and/or the dorsal mantle length and the POPs concentration levels.

## 3. RESULTS AND DISCUSSION

The persistent organochlorine pesticide residues in the mantle tissues of the common cuttlefish, *Sepia pharaonis*, from the Arabian Gulf water of the Kingdom of the Saudi Arabia are shown in Table 1 and Fig. 2. The accumulation order of the different contaminants was CHLs > DDT > HCH.



**Fig. 2.** Concentrations ( $\text{ng g}^{-1}$  lipid wt.) of organochlorine compounds in squid samples from Arabian Gulf of Saudi Arabia.

### DDT

In the present study, DDT mean values ranged between 29.4 - 55 ng/g lipid weight in the mantle tissues of *S. pharaonis*. These values are remarkably higher than those detected in the same species collected from the Red sea of Yemen ( $1.8 \pm 0.02$  ng/g) and the Gulf of Aden ( $1.5 \pm 0.02$  ng/g) (Al-Shwafi et al. (2009). Moreover, our mean values were higher than that recorded in the edible parts of *Sepia officinalis* (9.1 ng/g) from the Gulf of Naples, Southern Italy (Naso et al, 2005).

However, our values were lower than the Maximum Residue Limits (MRL) (1 mg/kg) for some foods of animal origin (Decreto Ministeriale 19 maggio, 2000). They were also below the maximum acceptable limit (MAL) (0.3 mg/g) proposed by the Food and Culture Organization FAO (1983).

On the other hand, the highest mean concentration (118-698 ng/g) of DDTs, so far, recorded in squid collected from offshore Korean waters (Yellow Sea) (Won, et al., 2009)

A preliminary survey of marine samples revealed the presence of DDT Derivatives in the oysters (*Pinctada margaritifera*) collected from Kuwait waters (Andrlini et al, 1981).

Douabul et al. (1987) proved the presence of DDT residues in 13 commercially important fish species collected from the North West of the Arabian Gulf, although, DDT has been officially banned in Iraq country. Organochlorine pesticides such as DDT are generally higher in the sediments of northwestern Gulf and Shatt Al-Arab than in the more southern sediments in Saudi Arabia, Qatar and AUE, suggesting the influence of Shatt Al-Arab discharge into the Gulf (Fowler, 2002).

There are 31 countries requesting public health exemptions for DDT under the Stockholm Convention, the Kingdom of the Saudi Arabia is one of them, those lists these additional countries as current users of DDT for vector control (WHO, 2001). Probably, this elucidates the compositions and the elevated residue concentrations of DDT found in cuttlefish samples in this study. This is confirmed by the work of Al Saleh et al. (2009) who mentioned that, although DDT is banned for agricultural purpose in Saudi Arabia, it is occasionally used to control vector-borne diseases in certain regions of the country. Naso et al. (2005) explain that the presence of the highest levels of DDT in (benthic and neritic) marine species clearly points to local sources of contamination mainly located along the marine coasts.

### CHL

The current study showed that the mean concentration values of CHL compounds ranged from 47.4 to 100 ng/g lipid wt. These values represent the maximum attainable concentration among the POPs analyzed. The peak concentration values of CHL in the present study may reflect the recent continuing illegal input of technical CHL from the countries located along the Arabian Gulf. Naso et al., (2005) found that the strong interaction with sea bed and its sediments may have an important influence on the degree of contamination detected in the benthic and neritic cephalopods.

To safeguard public health, CHL concentration standards in food has been established by various Organizations. In 1970, the FAO/WHO Joint Meeting on Pesticide Residues (JMPR) evaluated chlordane and established tolerances for residues in food of 0.02-0.5 mg/kg (FAO/WHO, 1971). In 1986, an acceptable daily intake (ADI) in food of 0-0.0005 mg/kg bw was established (FAO/WHO, 1987). In 1994, the ADI was changed to a provisional tolerable daily intake (PTDI) value at the same level, 0.0005 mg/kg bw (WHO, 1999).

CHL concentrations in the mantle of squid from the Korean offshore waters were ranging from 4 to 22 ng/g lipid wt, for animals collected from the Yellow Sea and from 9 to 12 ng/g lipid wt. for squids collected from the East Sea (Won et al., 2009).

### HCH

Among the POPs analyzed, HCH showed the lowest concentrations (Table 1, Fig. 2). The mean values ranged 1-5 ng/g lipid wt. These low values are most probably due to the fact that HCHs have high vapor pressure and volatilize easily from the contamination source for wider atmospheric distribution (Loganathan & Kannan, 1994). HCHs were determined in the mantle of the Japanese common squid collected from the offshore waters of Korea. The concentrations were ranged between 3 - 11 ng/g in samples having mantle lengths ranged from 22.6 to 25.4 cm (Won et al., 2009). Obviously, these values are remarkably higher than those detected in the present study.

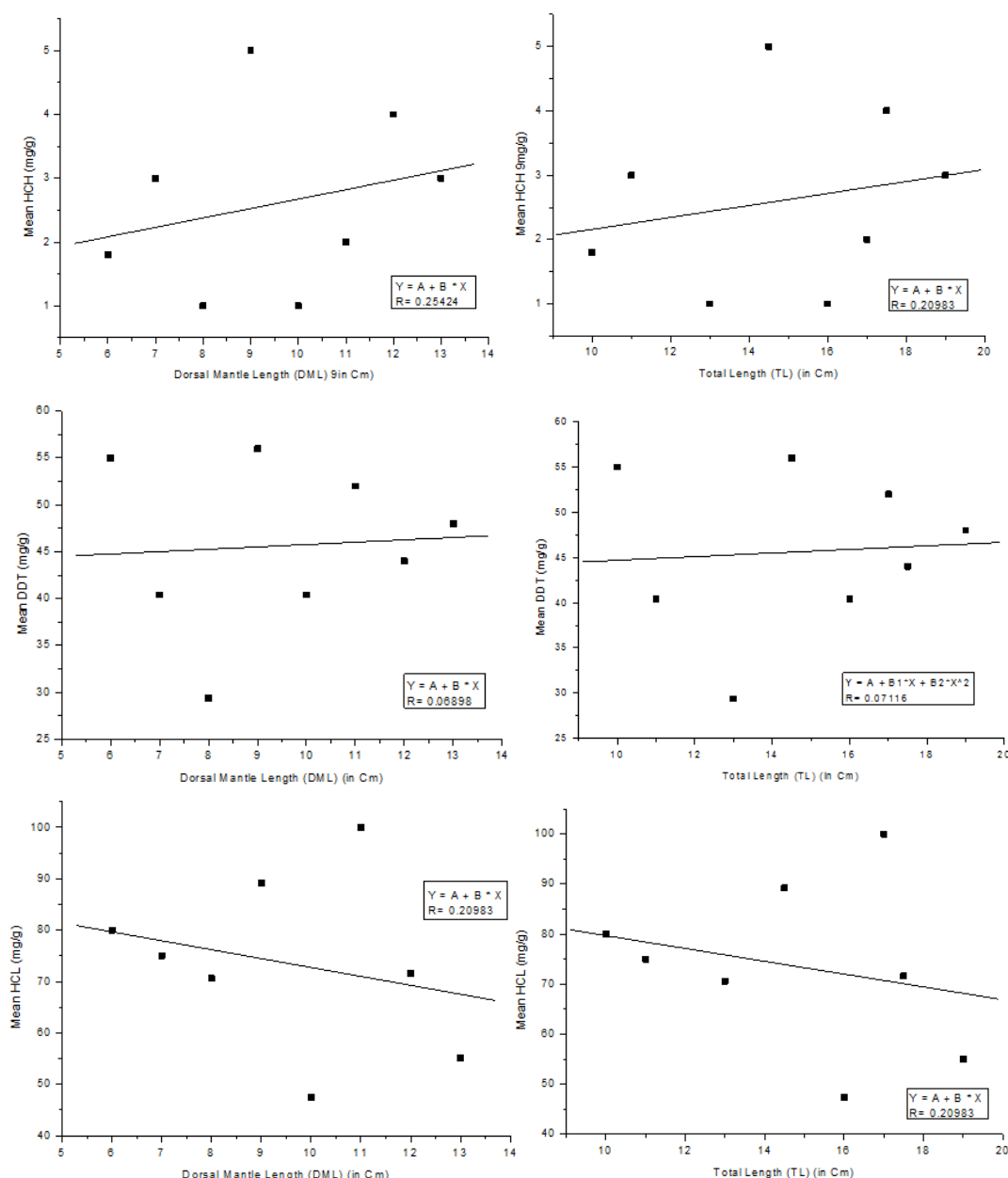
Several studies have discussed the relationship between HCH and DDT. Beyer et al. (2000) assessed long-range transport potential of OCs via air and water, and found that a-HCH is 4–5 times more transportable than DDT breakdown product of (DDE). Hong et al. (2006) and Hong et al., (2008) declared that HCHs show an even distribution in the environment with little spatial variation in comparison to DDTs. Won et al. (2009) mentioned that the overall level of HCH compounds in squid were clearly lower than that of DDTs which has been frequently observed in various marine organisms (Lee et al., 1997; Kajiwara et al., 2002; Monirith et al., 2003; Kajiwara et al., 2004).

Compared with other POPs, the low bioaccumulation of HCHs is due to their physicochemical properties such as high biodegradability and low lipophilicity (Loganathan & Kannan, 1994). Tanabe et al. (1984) reported that the bioconcentration factor of HCHs between squid and seawater in the western North Pacific ecosystem was 300–400 times lower than those of DDTs.

### **Relationship between POPs concentrations and body-length and/or mantle length**

Results from the statistical analysis showed no significant relationship between body length and/or mantle length and the concentrations of the studied POPs (Fig. 3). These findings are in agreement with the results obtained by Al-Shwafi, et al. (2009). They concluded that there is no relationship between body length and DDTs concentration in the muscle of the *S. pharaonis* collected from both the Red sea of Yemen and the Gulf of Aden.

Similarly, this agrees with the results of Ueno et al. (2003) on the liver Japanese common squid *Todarodes pacificus* and that of Sato et al. (2000), who reported no significant correlation between body-length and a-HCH and PCBs concentrations in the Japanese common squid on dry weight basis.



**Fig. 3.** Variations of metal concentrations (mg/kg wet weight) in the cuttlefish *Sepia pharaonis* in relation to the mantle length and the dorsal mantle length.

#### 4. CONCLUSIONS

POPs such as, DDTs, CHLs and HCHs were determined in the mantle of the common cuttlefish, *Sepia pharaonis*, collected from the Arabian Gulf water of the Kingdom of Saudi Arabia. Peak levels of DDT and CHL were detected but HCH was low. The present data were compared to the international permissible and/or acceptable levels recommended by the organizations concerned with human health. From the human health point of view, the concentrations of POPs in the *S. pharaonis* show situation of no hazard effect for the consumers. In the study, no significant relationship was detected between the total body length and/ or dorsal mantle length and POPs concentrations.

Finally, it is obligatory to enact repressive laws and activate them to control industrial waste discharge and to avoid dispersal of these persistent toxic contaminants into the Arab countries environment. Additional studies should be carried out to monitor the levels of organochlorine pollutants in edible fish and shellfish inhabiting the marine water of the Arabian Gulf.

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## REFERENCES

1. Al-Saleh I., Coskun S., El-Doush I., Billedo G., Mashhour A., Jaroudi K., Al-Shahrani A., Al-Mayman H., Mohamed G. 2009. Outcome of in-vitro fertilization treatment and DDT levels in serum and follicular fluid. Medical science monitor, 15(11): 320-33.
2. Al Shwafi N., Al-trabeen K. and Rasheed M. 2009. Organochlorine Pesticides and Polychlorinated Biphenyls Carcinogens Residual in some Fish and Shell Fish of Yemen. Jordan Journal of Biological Sciences, 2(1): 23-28.
3. Anderlini V. C., Al-Harmi L. Delappe B. W. Risebrough R. W. Walker W. Simoneit B. R. T. and Newton A. S. 1981. Distribution of hydrocarbons in the oyster, *Pinctada margaritifera*, along the coast of Kuwait. Marine Pollution Bulletin, 12: 57-62.
4. Aoyama T. and Nguyan T. 1989. Stock assessment of cuttlefish off the coast of the people's Democratic Republic of Yemen. Shimonosaki University of Fisheries, 37(203): 61-112.
5. Beyer A., Mackay D., Matthies M., Wania F. and Webster E. 2000. Assessing longrange transport potential of persistent organic pollutants. Environmental Science and Technology, 34: 699-703.
6. Butty J. S. and Holdway D. A., 1997. Assessment of the octopus *Octopus pallidus*, as a potential bioindicator of xenobiotic challenge: baseline studies. Marine Pollution Bulletin, 34: 564-570.
7. Danis B., Bustamante P., Cotret O., Teyssie J. L., Fowler S. W. and Warnau M., 2005. Bioaccumulation of PCBs in the cuttlefish *Sepia officinalis* from seawater, sediment and food pathways. Environmental Pollution, 134: 113-122.
8. Decreto Ministeriale 19 maggio, 2000. Maximum limit of pesticide residue in foods (Recepimento delle Direttive n. 97/41/CE, n. 1999/65/CE e n. 1999/71/CE).
9. G.U. 207 del 05/09/2000 (in Italian).
10. Depeng L., Leyuan, S., Xili, L. and Xiance, Y., 1996. A successful control programme for falciparum malaria in Xinyang, China. Transactions of the Royal Society of Tropical Medicine and Hygiene, 90: 100-102.
11. Douabul A.A.Z., Al-Saad H.T., Al-Obaidy S.Z., Al-Rekabi H.N., 1987. Residues of organochlorine pesticides in fish from the Arabian Gulf. Water, Air, and Soil Pollution 35, 187-194.
12. Ehrhardt M. 1987. Lipophilic organic material: an apparatus for extracting solids used for their concentration from seawater. ICES Techniques in Marine Environmental Sciences 4: 1-14.
13. FAO, 1983. Compilation of Legal Limits for Hazardous Substances in Fish and Fishery Products, FAO Fishery Circular No. 464, 5-100
14. FAO/WHO. 1970. Pesticide residues in food. Report of the 1969 Joint Meeting of the FAO Working Party of experts on Pesticide Residues and the WHO Expert Committee on Pesticide Residues. FAO Agricultural Studies, No. 84; WHO Technical Report Series, No. 458.
15. FAO/WHO. 1971. Pesticide residues in food. Report of the 1970 Joint Meeting of the FAO Working Party of experts on Pesticide Residues and the WHO Expert Committee on Pesticide Residues. FAO Agricultural Studies, No. 87; WHO Technical Report Series, No. 474.
16. FAO/WHO. 1987. Pesticide residues in food - 1987. Report of the Joint Meeting of the FAO Panel of Experts on Pesticide Residues in Food and the Environment and the WHO Expert Group on Pesticide Residues. FAO Plant Production and Protection Paper 84.
17. Fowler S. W. 2002. Agrochemicals. In: Price, A.R.G. (Ed.), The Gulf Ecosystem: Health and Sustainability. Bakhuis Publishers, Leiden, pp. 193-217.
18. Graham J. P., Mastic L. C. and Boyle P. R. 1994. Morphometric variation in *Loligo forebsi* and *L. vulgaris*: regional, seasonal, sex, maturity and worker differences. Fisheries Research, 21: 127-148.
19. Jones K. C. and de Voogt P. 1999. Persistent organic pollutants (POPs): state of the science. Environmental Pollution, 100: 209-221.
20. Kajiwaru N., Watanabe M., Tanabe S., Nakamatsu K., Amano M. and Miyazaki N., 2002. Specific accumulation and temporal trends of organochlorine contaminants in Dall's porpoises (*Phocoenoides dalli*) from Japanese coastal waters. Marine Pollution Bulletin, 44: 1089-1099.
21. Kajiwaru N., Ueno D., Takahashi A., Baba N. and Tanabe S. 2004. Polybrominated diphenyl ethers and organochlorines in archived northern fur seal samples from the Pacific Coast of Japan, 1972-1988. Environmental Science and Technology 38, 3804-3809.
22. Kelce W. D., Stone C. R., Laws S. C., Gray L. E., Kemppainen J. A. and Wilson E. M. 1995. The persistent DDT metabolite p,p'-DDE is a potent androgen receptor antagonist. Nature, 375:581-5.
23. Koueta N. and Boucaud-Camou, E. 2001. Basic growth relations during rearing of early juvenile cuttlefish *Sepia officinalis* L. (Mollusca Cephalopoda). Journal of Experimental Marine Biology and Ecology, 265: 75-87.
24. Lee J. S., Tanabe S., Takemoto N. and Kubodera T., 1997. Organochlorine residues in deep-sea organisms from Suruga Bay, Japan. Marine Pollution Bulletin 34, 250-258.
25. Loganathan B.G. and Kannan K., 1994. Global organochlorine contamination trends: an overview. Ambio, 23: 187-191.
26. MAFFJ (The Ministry of Agriculture, Forestry and Fisheries of Japan). 2001. The Status of Stockpiled and Buried Pesticides. Available from <<http://www.maff.go.jp/work/press010607-2.pdf>> (in Japanese).

27. McConnell L. L., Kucklick J. R., Bidleman T. F., Ivanov G. P. and Chernyak S. M. 1996. Air–water gas exchange of organochlorine compounds in Lake Baikal, Russia. *Environmental Science and Technology*, 30: 2975-2983.
28. Metcalf R. L. 1977. *Pesticides in aquatic environment* (M.A. Q. Khan, Ed.), Plenum Press, New York, p 127.
29. Miramand P. and Bentley D., 1992. Concentration and distribution of heavy metals in tissues of two cephalopods, *Eledone cirrhosa* and *Sepia officinalis*, from the French coast of the English Channel. *Marine Biology*, 114, 407-414.
30. Monirith I., Ueno D., Takahashi S., Nakata H., Sudaryanto A., Subramanian A., Karuppiah S., Ismail A., Muchtar M., Zheng J., Richardson B.J., Prudente M., Hue N.D., Tana T.S., Tkalin A.V. and Tanabe S. 2003. Asia-Pacific mussel watch: monitoring contamination of persistent organochlorine compounds in coastal waters of Asian countries. *Marine Pollution Bulletin*, 46: 281-300.
31. Nakata H., Tanabe S., Tatsukawa R., Amano M., Miyazaki N. and Petrov E. A. 1995. Persistent organochlorine residues and their accumulation kinetics in Baikal Seal (*Phoca sibirica*) from Lake Baikal, Russia. *Environmental Science and Technology*, 29: 2877-2885.
32. Naso B., Perrone D., Ferrante M. C., Bilancione M., Lucisano A. 2005. Persistent organic pollutants in edible marine species from the Gulf of Naples, Southern Italy. *Science of the Total Environment*, 343: 83-95
33. Navarro J. C. and Villanueva R. 2001. Lipid and fatty acid composition of early stages of cephalopods: an approach to their lipid requirements. *Aquaculture*, 183, 161-177.
34. Ratcliffe D. A. 1967. Decrease in eggshell weight in certain birds of pray. *Nature*, 215: 208-10.
35. Sato K., Kajiwaru N. and Hashimoto S. 2000. Accumulative characteristics of organochlorine compounds (OCs) in squids. *Bulletin of the Japanese Society of Scientific Fisheries* 66 (4): 658-665.
36. Silas E. G., Sarvesan R., Nair K. P., Sastri Y. A., Sreenivasan P. V., Meiyappan M. M., Vidyasagar K., Rao K. S. and Rao B. N. 1985. Some aspects of the biology of cuttlefishes. *Cephalopod bionomics fisheries and resources of the exclusive economic zone on India*. Central marine Fisheries Research Inst. Cochin, India, 37: 49-70.
37. Skaare J. U., Bernhoft A., Derocher A., Gabrielsen G. W., Gokbyr G. W., Henriksen E, et al. 2000. Organochlorines in top predators at Svalbard-occurrence, levels and effects. *Toxicology Letters*, 112/113: 103-9.
38. Stockholm Convention on Persistent Organic Pollutants (POPs), Annex B (Restriction), Part II, para. 1-7. Treaty text available online at [www.chem.unep.ch/pops](http://www.chem.unep.ch/pops).
39. Storelli M. M., Barone G., Marcotrigiano G. O., 2005. Cadmium in cephalopod molluscs: implications for public health. *The Journal of Food Protection*, 68: 577-580.
40. Tanabe S., Tanaka H. and Tatsukawa R., 1984. Polychlorobiphenyls, DDT and hexachlorocyclohexane isomers in the Western North Pacific Ecosystem. *Archives of Environment Contamination Toxicology* 13, 731-738.
41. Tehranifard A. and Dastan K. 2011. General morphological characteristics of the *Sepia Pharaonis* (cephalopoda) from Persian Gulf, Bushehr region. *International Conference on Biomedical Engineering and Technology*. IPCBEE vol.11 (2011) © (2011) IACSIT Press, Singapore.
42. Toft G., Edwards T. M., Baatrup E. and Guillette L. J. Jr. 2003. Disturbed sexual characteristics in male mosquito fish (*Gambusia holbrooki*) from a lake contaminated with endocrine disruptors. *Environ Health Perspect*, 111: 695-701.
43. Tolosa I., De Mora S. J., Fowler S. W., Villeneuve J.P., Bartocci J. and Cattini C. 2005 Aliphatic and aromatic hydrocarbons in marine biota and coastal sediments from the Gulf and the Gulf of Oman. *Marine Pollution Bulletin*, 50: 1619-1633.
44. Ueno D., Inoue S., Ikeda K., Tanaka H., Yamada H. and Tanabe S. 2003. Specific accumulation of polychlorinated biphenyls and organochlorine pesticides in Japanese common squid as a bioindicator. *Environmental Pollution*, 125: 227-235.
45. Wade T. L., Atlas E. L., Brooks J. M., Kennicutt II, M. C., Fox R. G., Sericano J. L., Garcia-Romero B. and Defreitas D. A. 1988. NOAA Gulf of Mexico Status and Trends program: Trace organic contaminant distribution in sediments and Oysters. *Estuaries*, 11: 171-179.
46. Wei D., Kameya T. and Urano K. 2007. Environmental management of pesticidal POPs in China: Past, present and future. *Environment International*, 33: 894-902.
47. Weisbrod A., Shea D., Leblanc G., Moore M. and Stegeman J. J. 2000. Organochlorine bioaccumulation and risk for whales in a Northwest Atlantic food web. *Marine Environmental Research*, 50: 431-441.
48. Weisbrod A., Shea D., Moore M. and Stegeman J. J. 2001. Species, tissue and gender-related organochlorine bioaccumulation in white-sided dolphins, pilot whales and their common prey in the Northwest Atlantic. *Marine Environmental Research*, 51, 29-50.
49. WHO (World Health Organization). 1999. Inventory of IPCS and Other WHO Pesticide Evaluations and Summary of Toxicological Evaluations Performed by the Joint Meeting on Pesticide Residues [JMPR] through 1999, 3rd Ed., Geneva, International Programme on Chemical Safety.
50. WHO (World Health Organization). 2001. Vector Control for Malaria and Other Mosquito-Borne Diseases, Geneva: World Health Organization, WHO Technical Report Series #857, 1995; and "Situation of Malaria Programs in the Americas," *Epidemiological Bulletin*, Vol. 22, No.1, March 2001, [wysiwyg://34/http/www.paho.org/English/SHA/be\\_v22n1-malaria.htm](http://www.paho.org/English/SHA/be_v22n1-malaria.htm)

51. Won J. H., Hong S. H. Shim W. J. Yim U. H. and Kim G. B. 2009. Persistent organochlorine pollutants in Korean offshore waters: Squid (*Todarodes pacificus*) as a biomonitor. Marine Pollution Bulletin. 58: 1229-1244.
52. Wong M. H., Leung A. O. W., Chan J. K. Y. and Choi M. P. K. 2005. A review on the usage of POP pesticides in China, with emphasis on DDT loadings in human milk. Chemosphere, 60(6): 740-752.