

Isolation and Purification of Proteins

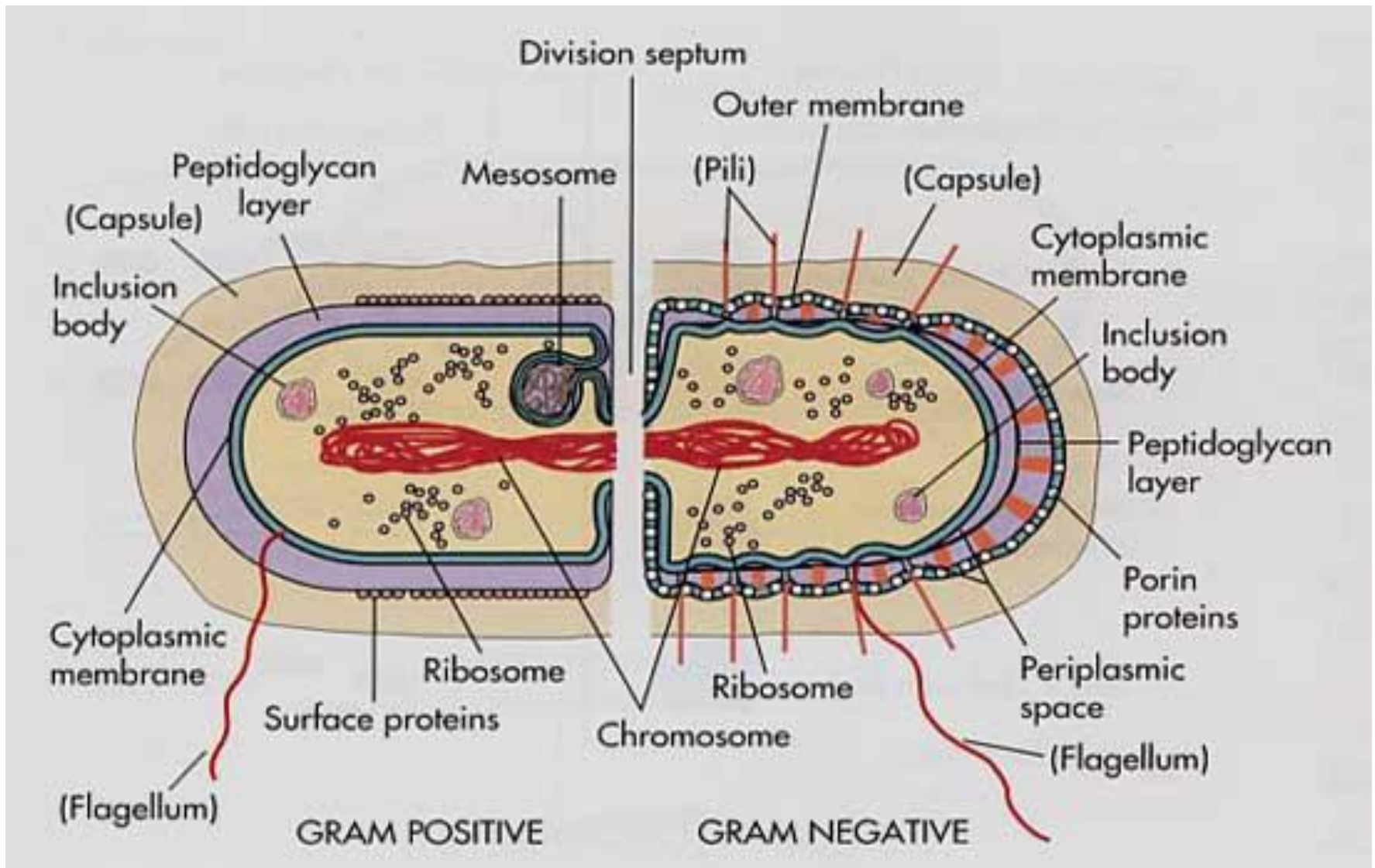
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Objectives of this Lecture

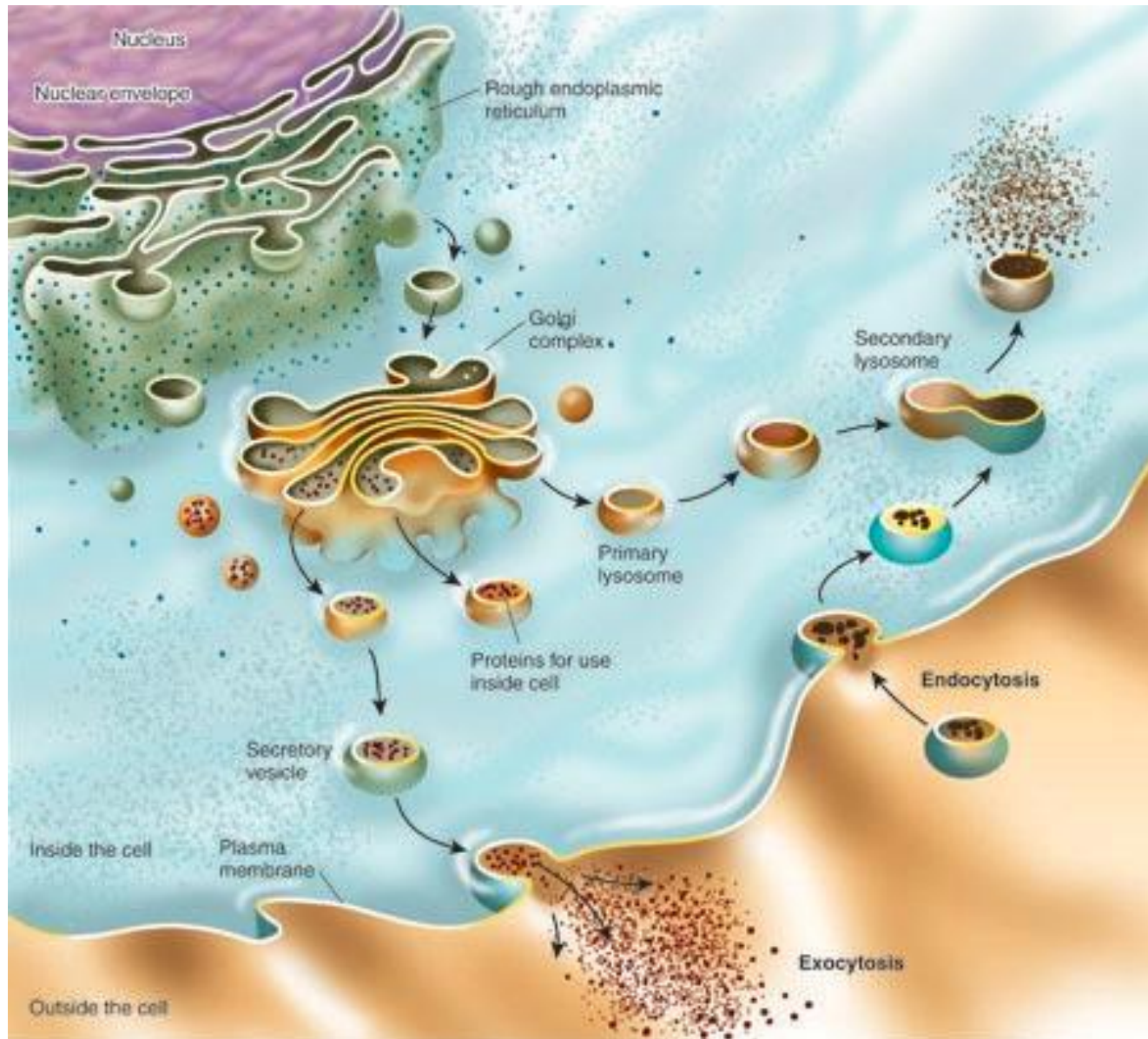
By the end of this lecture you will be able to:

1. Describe most common methods of protein isolation and purification
2. Compare between different methods of protein purification
3. Construct a purification algorithm based on your knowledge in protein purification

Intracellular and Extracellular



Intracellular and Extracellular

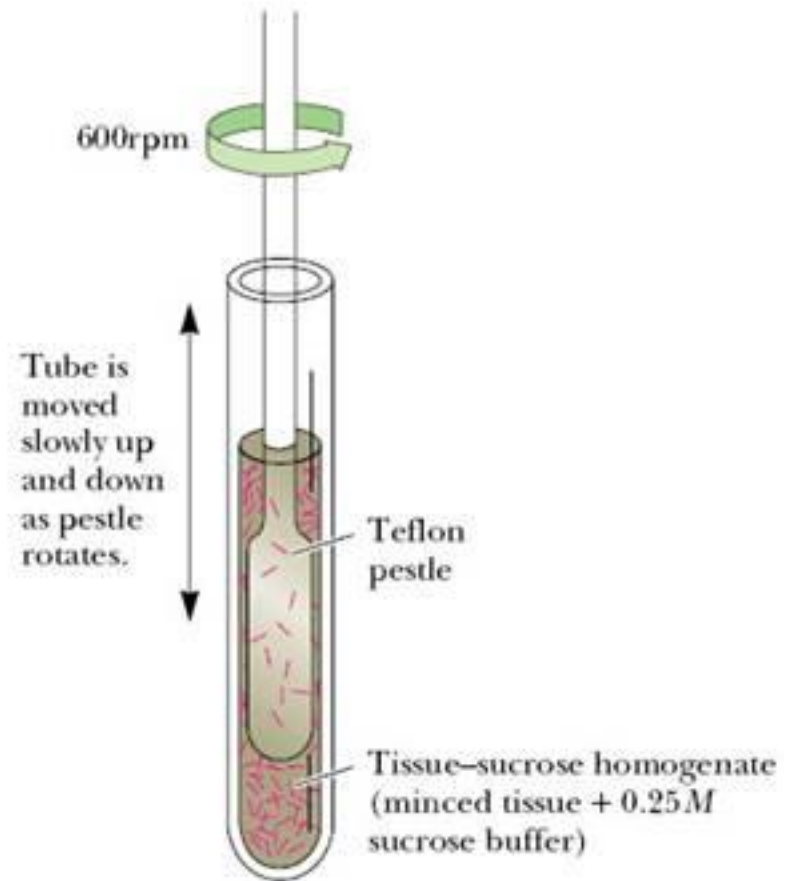


Isolation of Extracellular Proteins

- No need for cell disruption
- Secreted soluble proteins can be collected in the cell supernatant after centrifugation
- Membrane-bound proteins might be released from the cell simply using detergents

Isolation of Intracellular Proteins

- Needs cell disruption:
 - Detergents lysis
 - Enzymatic lysis
 - Osmotic lysis
 - Freeze-thaw cycles
 - Ultrasonication
 - Homogenization



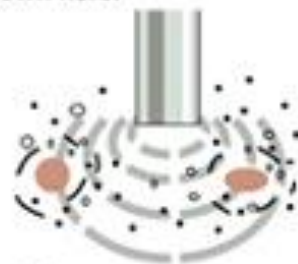
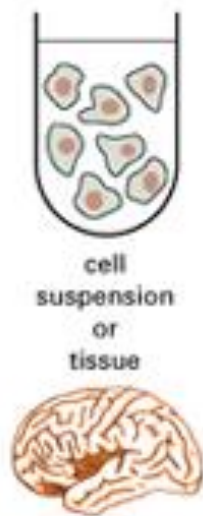
Isolation of Intracellular Proteins

BREAKING CELLS AND TISSUES

The first step in the purification of most proteins is to disrupt tissues and cells in a controlled fashion.

Using gentle mechanical procedures, called homogenization, the plasma membranes of cells can be ruptured so that the cell contents are released. Four commonly used procedures are shown here.

The resulting thick soup (called a homogenate or an extract) contains large and small molecules from the cytosol, such as enzymes, ribosomes, and metabolites, as well as all the membrane-bounded organelles.



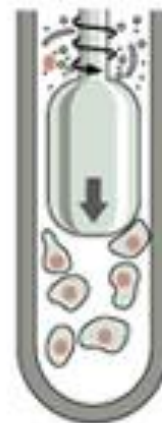
- ① break cells with high frequency sound



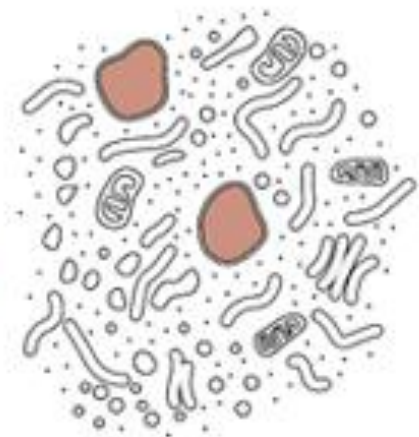
- ② use a mild detergent to make holes in the plasma membrane



- ③ force cells through a small hole using high pressure



- ④ shear cells between a close-fitting rotating plunger and the thick walls of a glass vessel



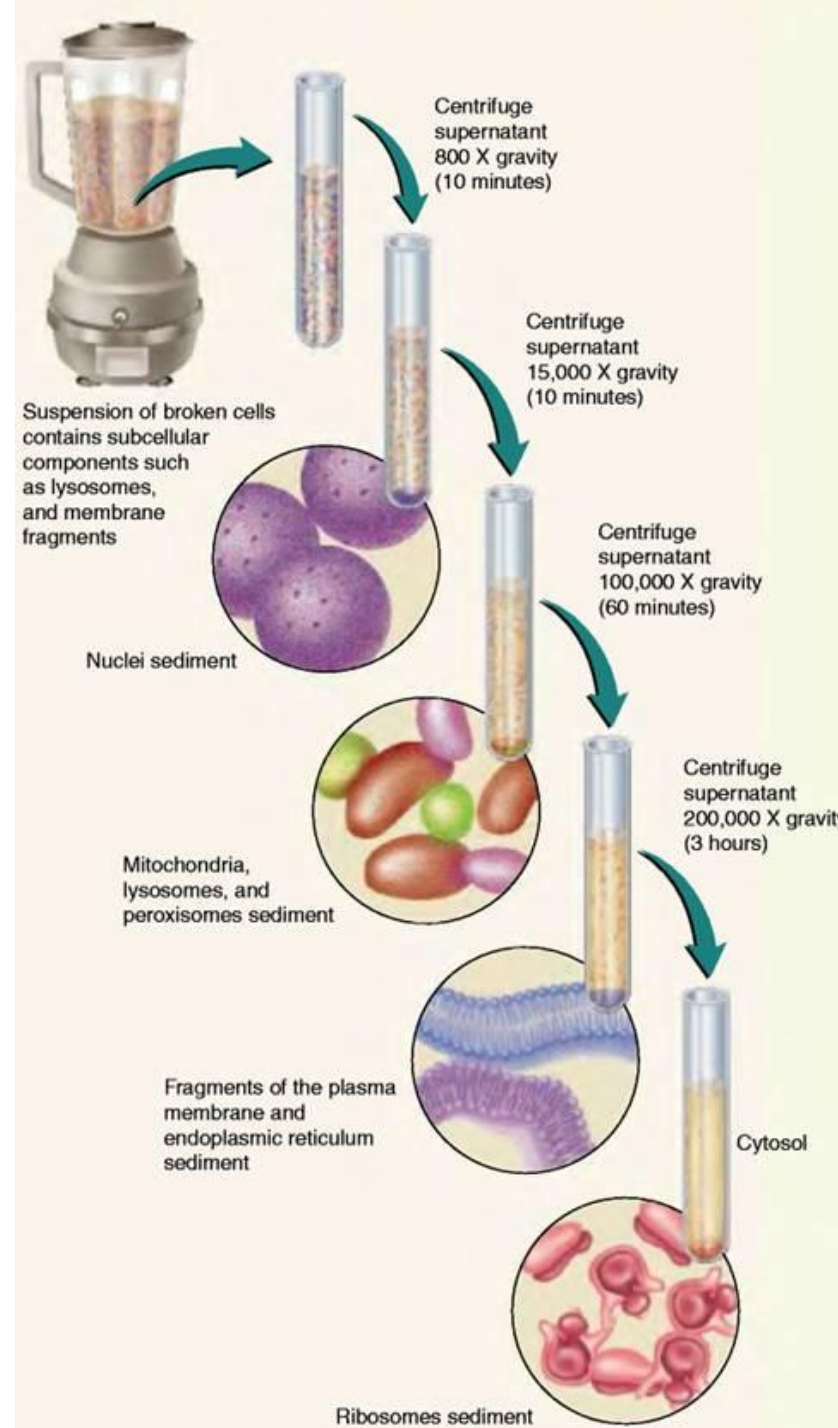
When carefully applied, homogenization leaves most of the membrane-bounded organelles intact.

Purification of Proteins

- Differential centrifugation
- Differential salt precipitation
- Differential solvent precipitation
- Preparative electrophoresis
- Column chromatography

**More than one approach
may be required**

Differential Centrifugation



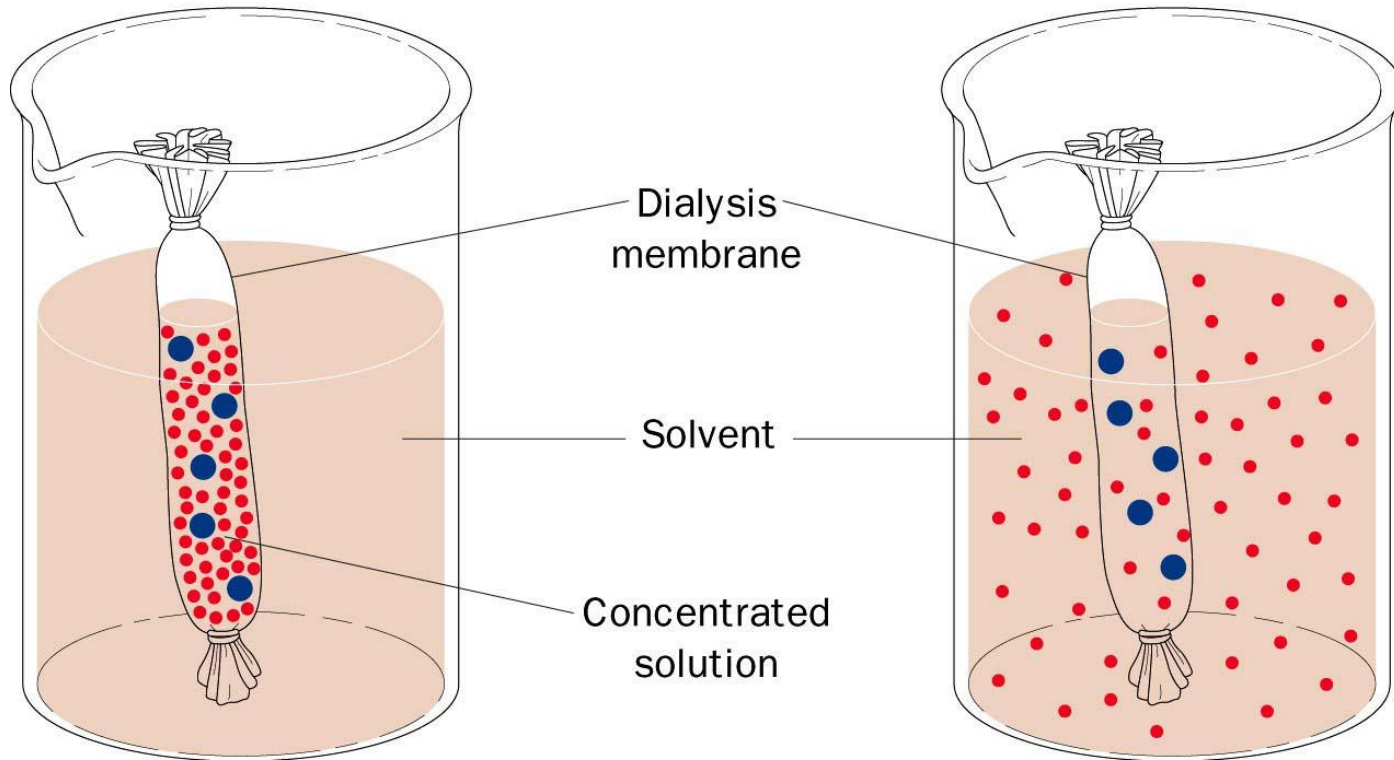
Differential Centrifugation

- **Separation of proteins (or any material) on the basis of their size, mass, and density**
- **It is a function of the size of the protein and the speed of centrifugation**
- **It gives us a rough separation. It is basically fractionating!**

Dialysis

(a) At start of dialysis

(b) At equilibrium



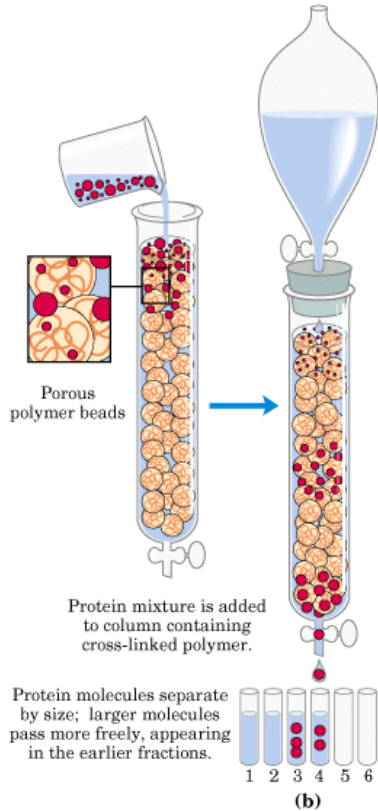
Dialysis

- **Based on osmotic pressure**
- **It allows you to get rid of most salt ions**
- **It is impossible to remove the salts completely**
 - *Why?*

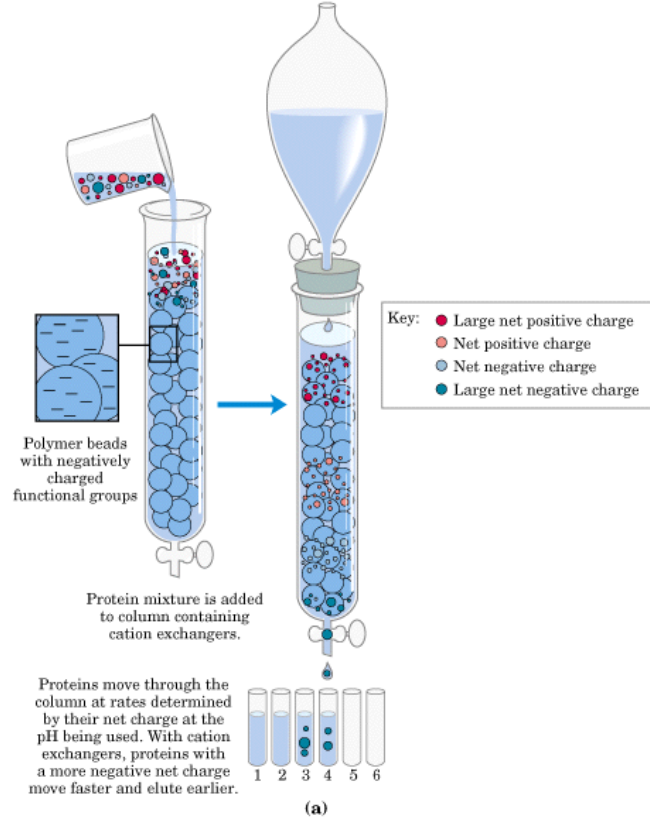
Column Chromatography

- Most common and best approach to purifying larger amounts of proteins
- Achieves Highest level of purity and largest amount
- Requires low effort
- Lowest likelihood to damage the protein product
- Standard method for pharmaceutical industry

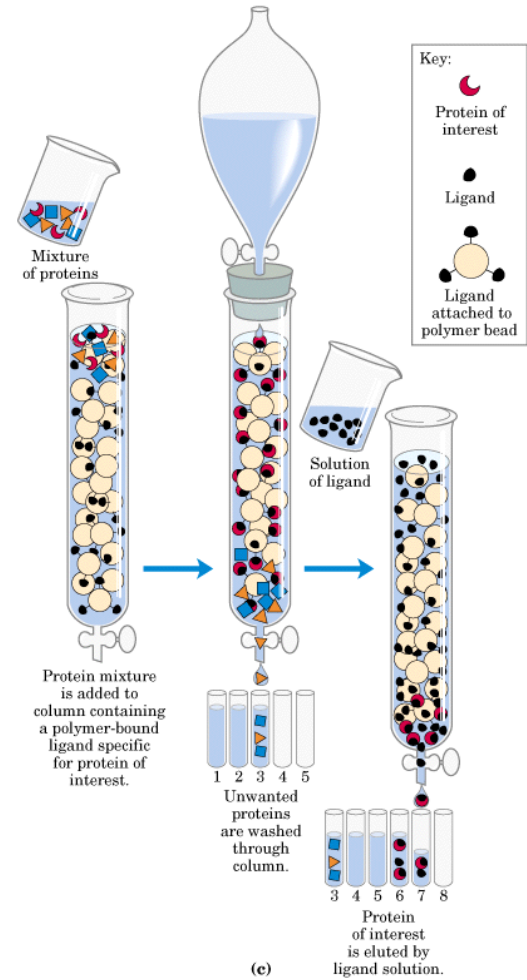
Gel Filtration



Ion Exchange

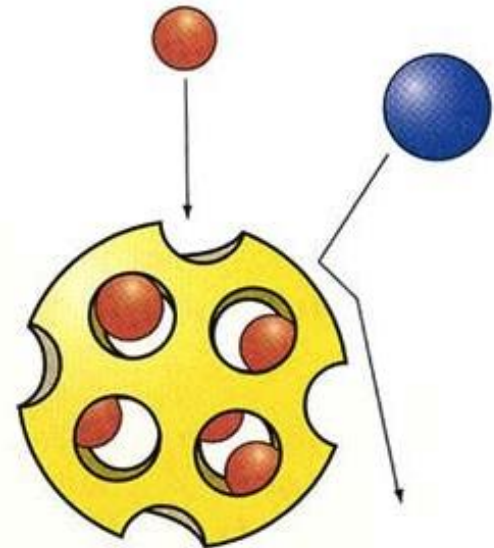


Affinity



Gel-Filtration Chromatography

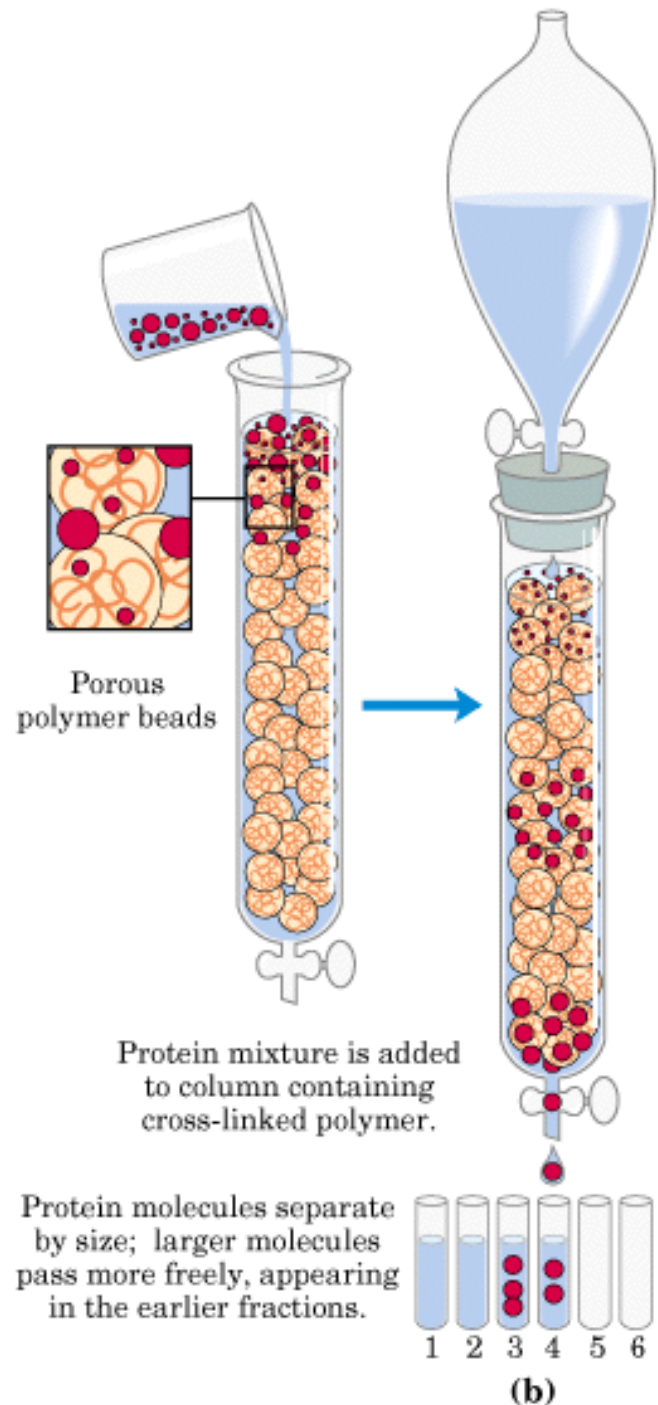
- AKA Size-Exclusion (or **molecular exclusion**) Chromatography
- Molecules are separated according to differences in their **size** as they pass through a hydrophilic polymer
- Polymer beads composed of cross-linked dextran (dextrose) which is highly and uniformly porous (like Swiss cheese)
- Large proteins come out first (can't fit in pores), small proteins come out last (get stuck in the pores)



Gel-Filtration Chromatography

You need to have a way to know where your protein is:

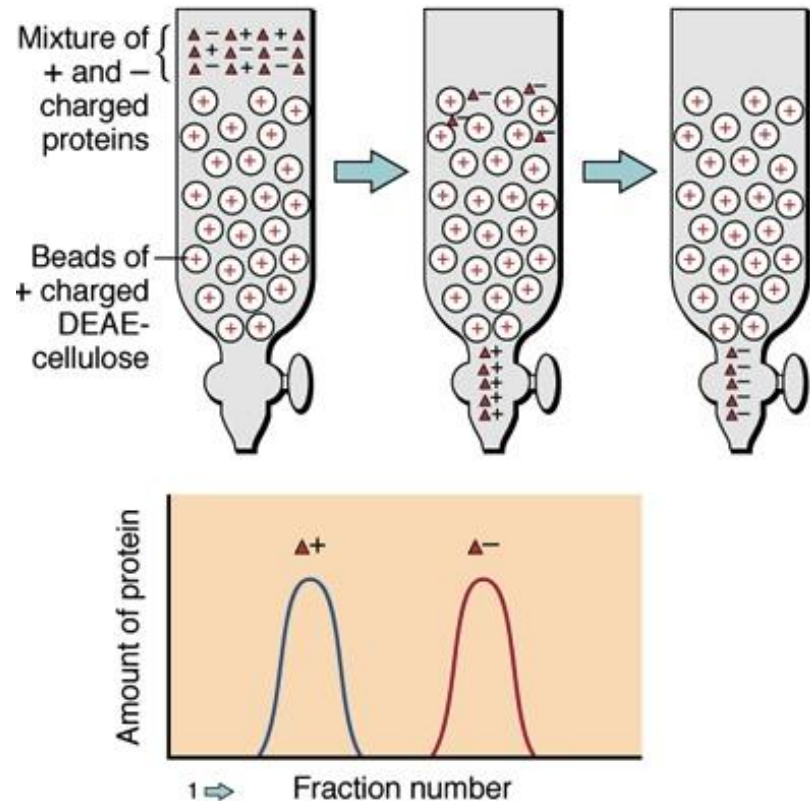
1. Molecular weight marker
2. Add a target to your protein e.g. if you want to purify an enzyme add a substrate to the tubes and see where does it appear



Ion-Exchange Chromatography

- Principle is to separate on basis of charge “adsorption”
- Highest resolving power
- Highest loading capacity
- Widespread applicability
- Most frequent chromatographic technique for protein purification

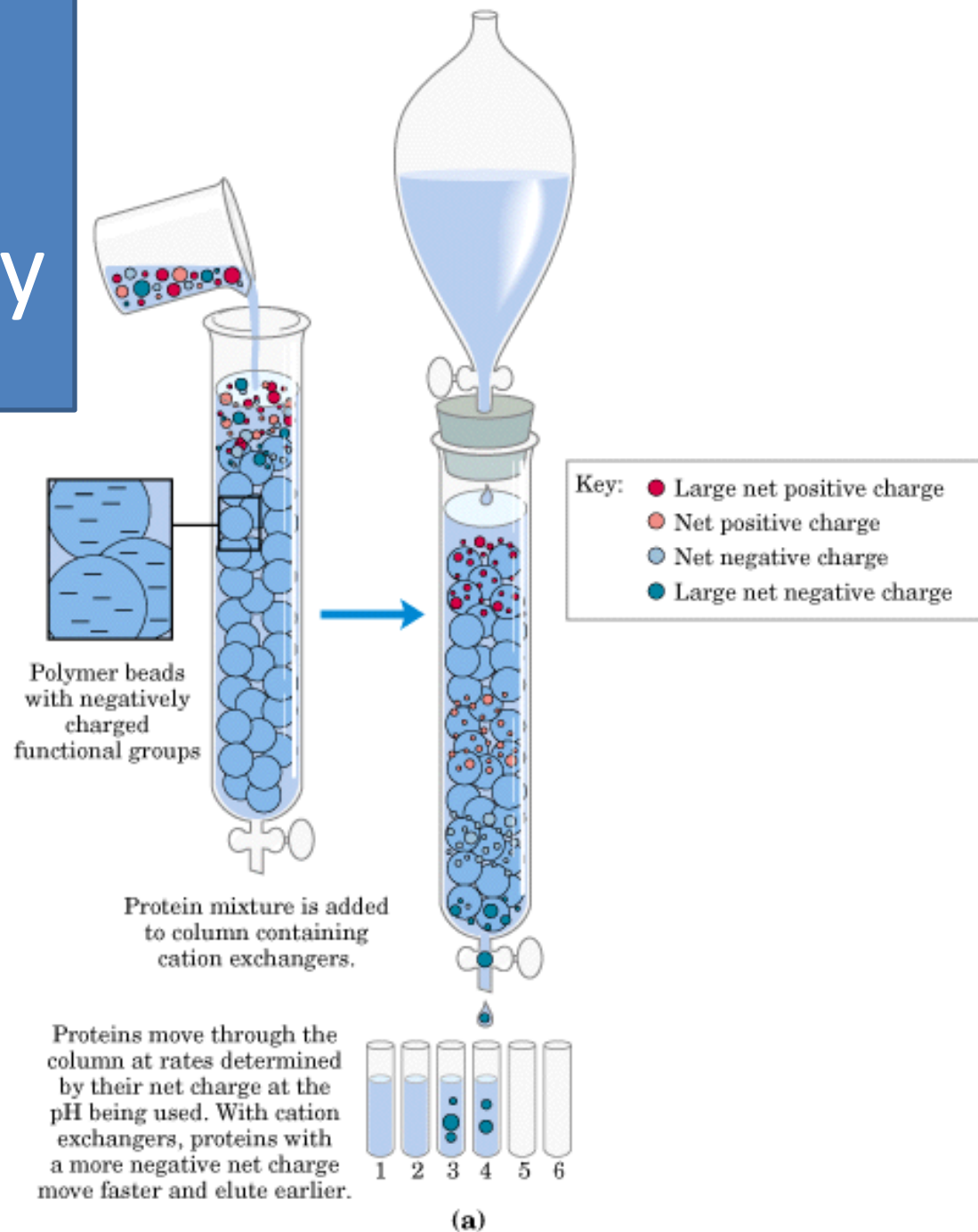
This column represents cation exchange chromatography



Ion-Exchange Chromatography

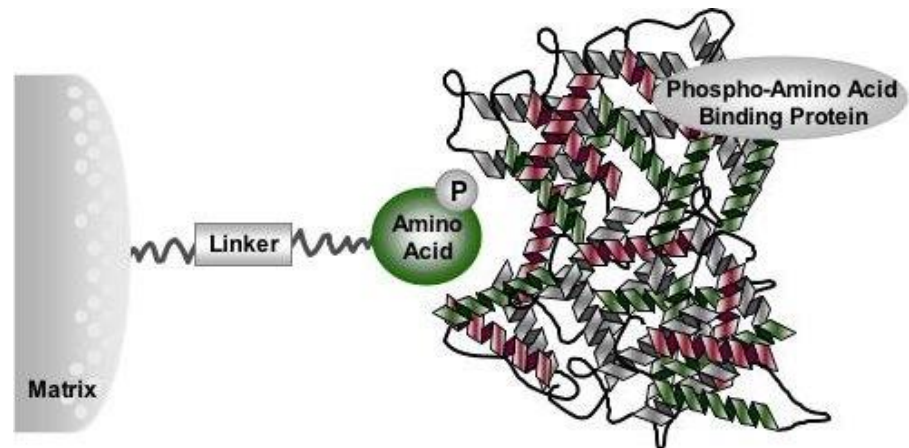
The ion exchange process is based on concentration of the counter ion

This column represents anion exchange chromatography



Affinity Chromatography

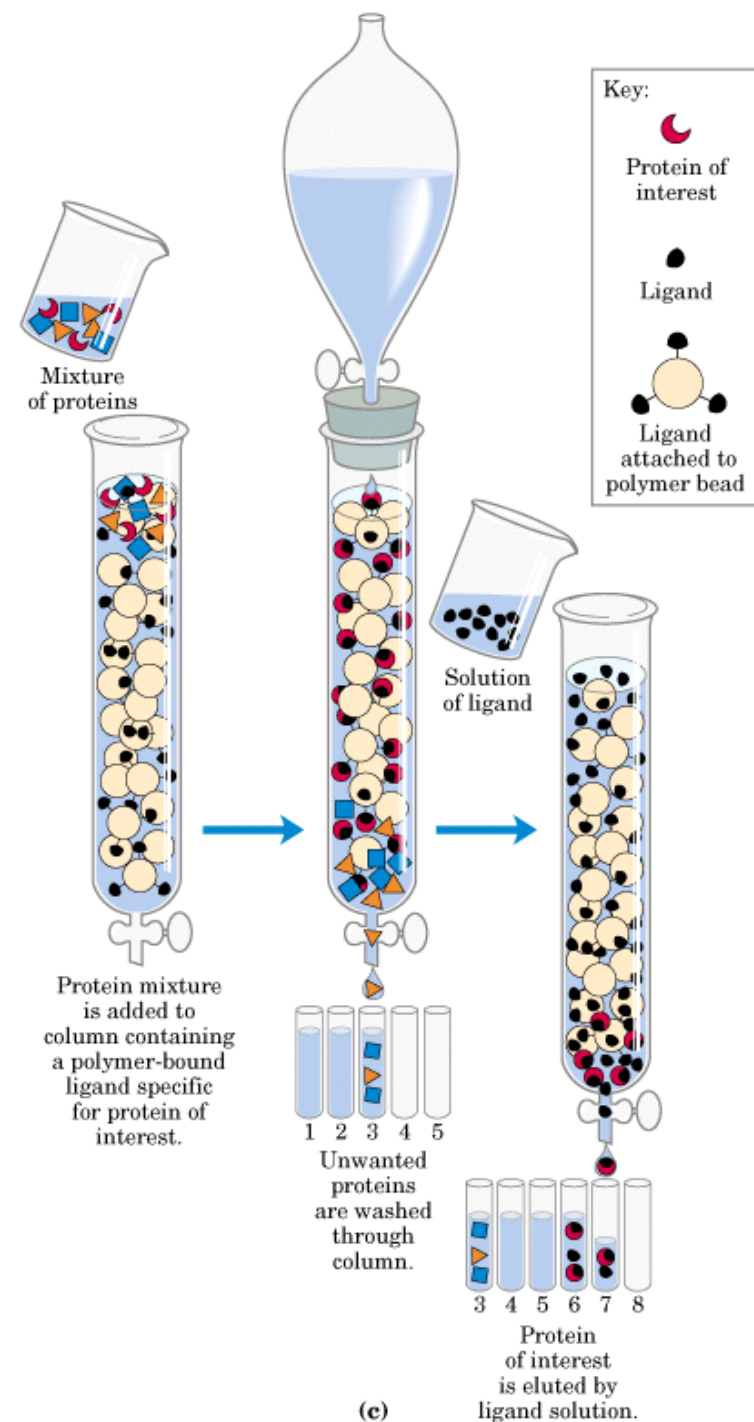
- Adsorptive separation in which the molecule to be purified specifically and reversibly binds (adsorbs) to a complementary binding substance (a ligand) immobilized on an insoluble support (a matrix or resin)
- Purification is 1000X or better from a single step (highest of all methods)
- Preferred method if possible



Affinity Chromatography

How can I get my protein out of the column?

The protein is not covalently bound to the ligand



Isoelectric Focusing

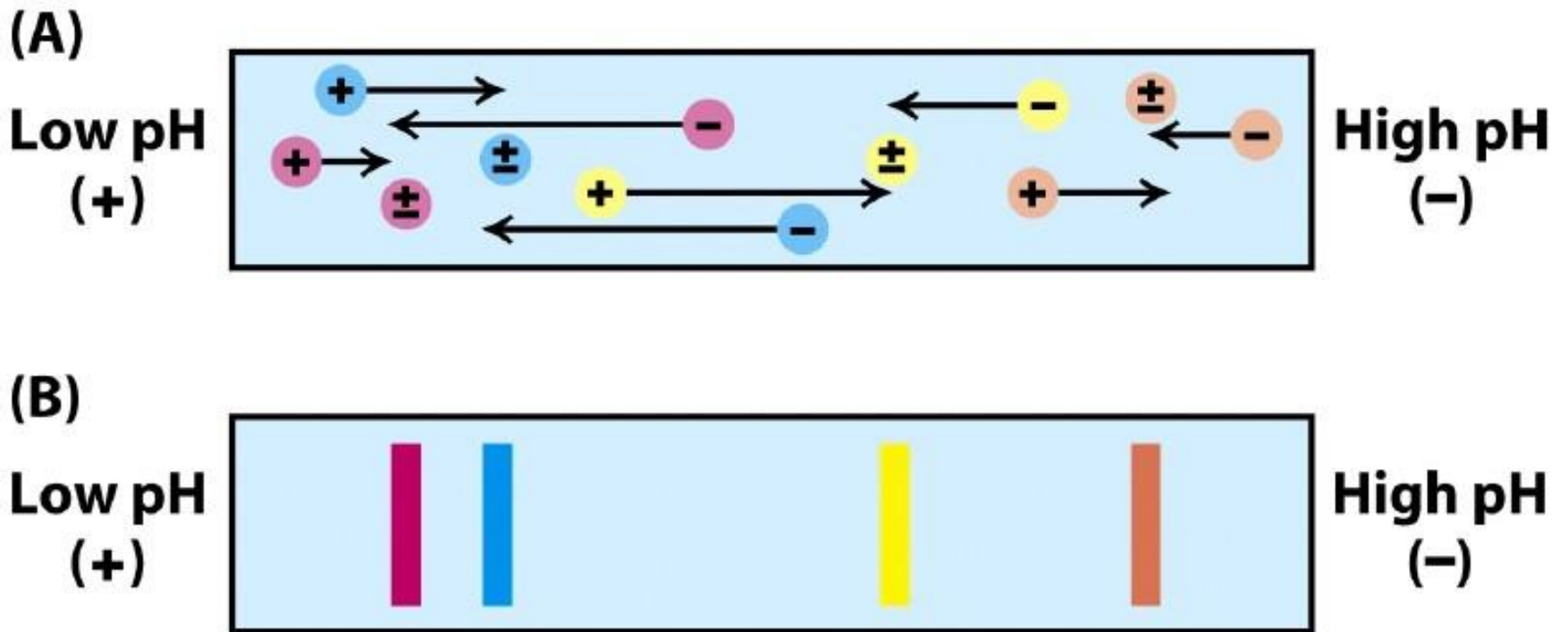


Figure 3-11
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Polyacrylamide Gel Electrophoresis

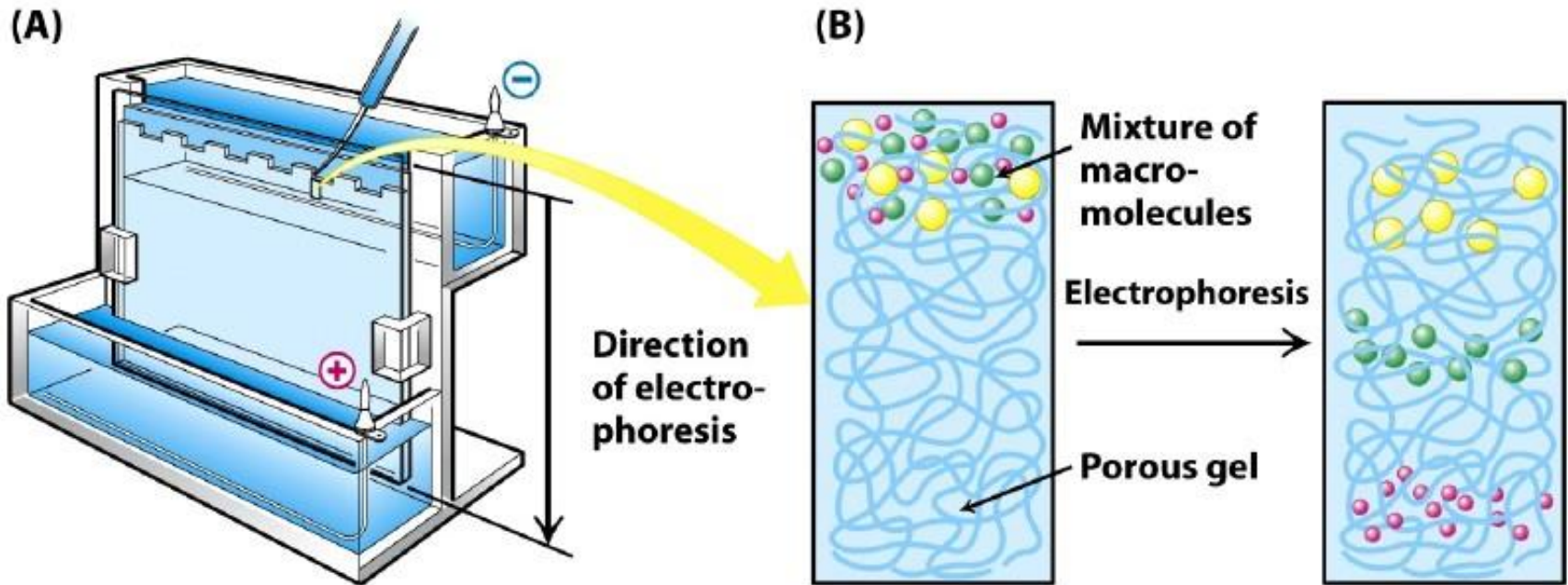


Figure 3-7
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Electrophoresis separates proteins based on their size using electrical current

Polyacrylamide Gel Electrophoresis

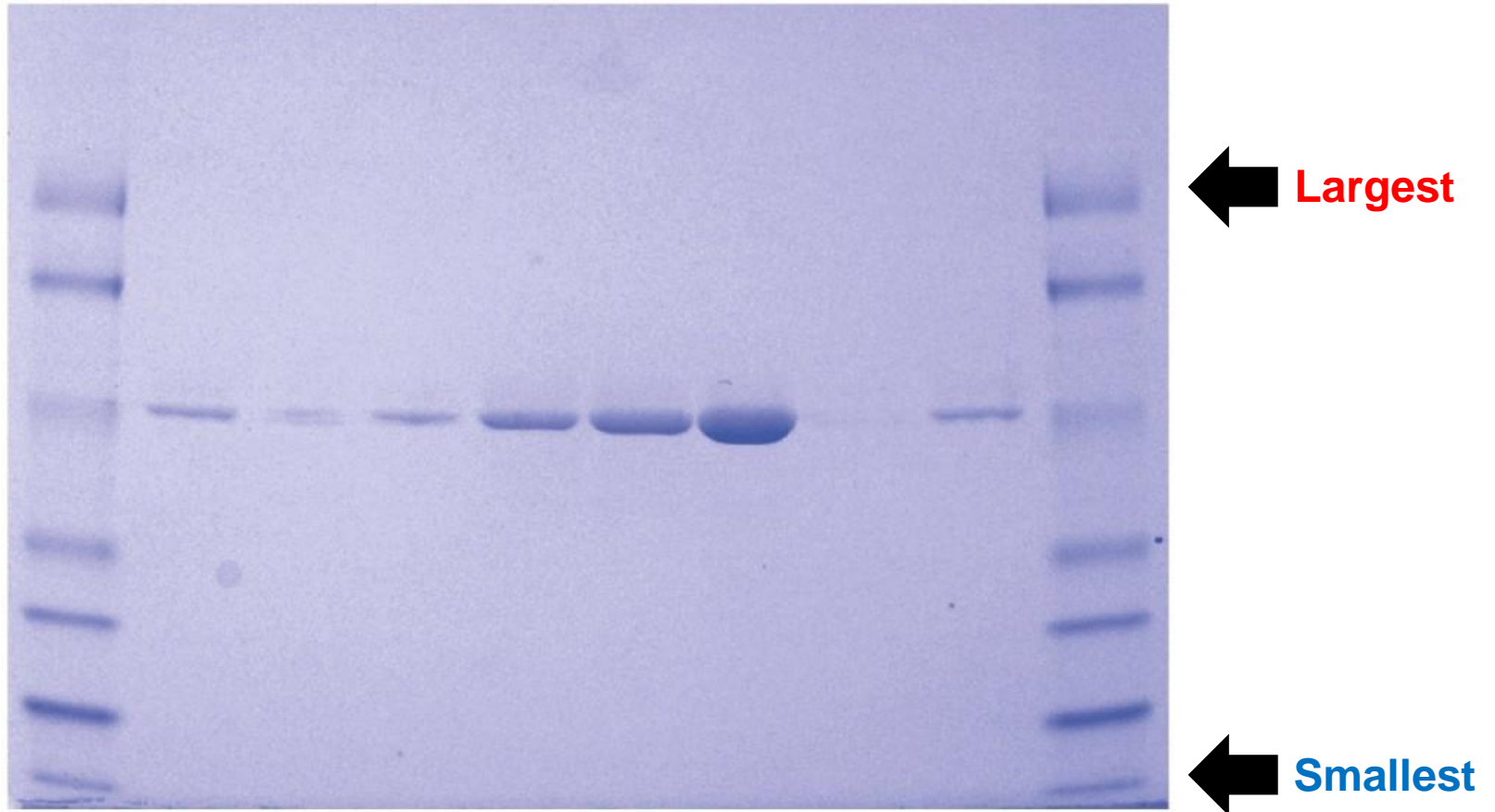
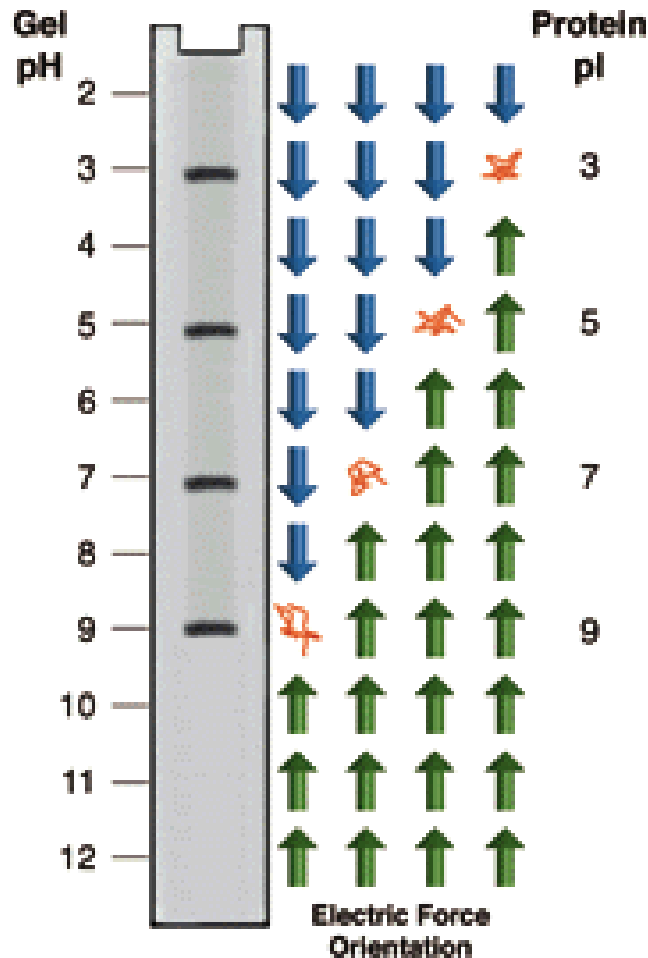


Figure 3-9
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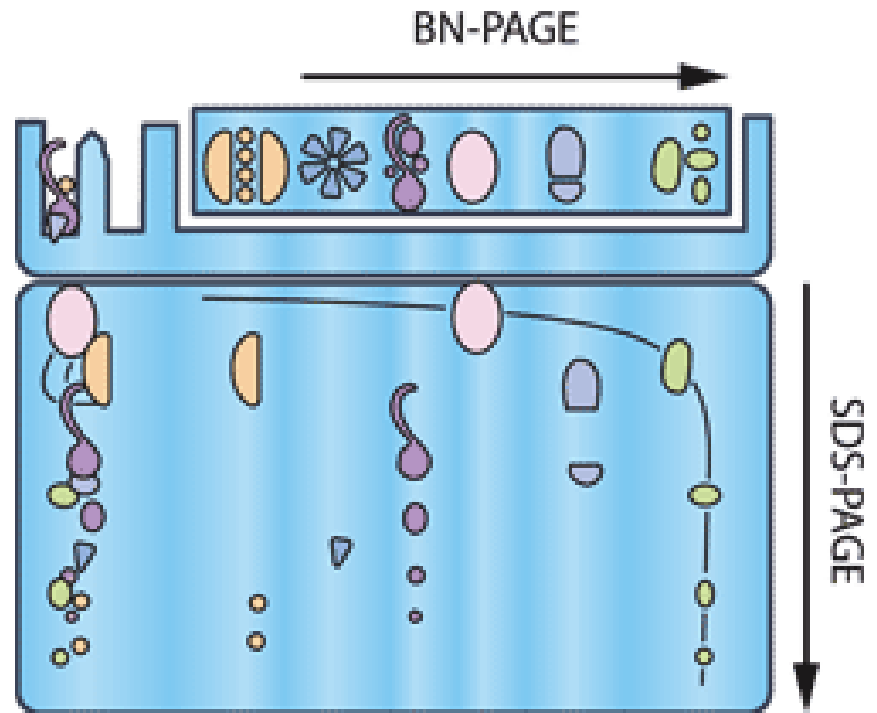
Preparative electrophoresis

2D Polyacrylamide Gel Electrophoresis (2D PAGE)

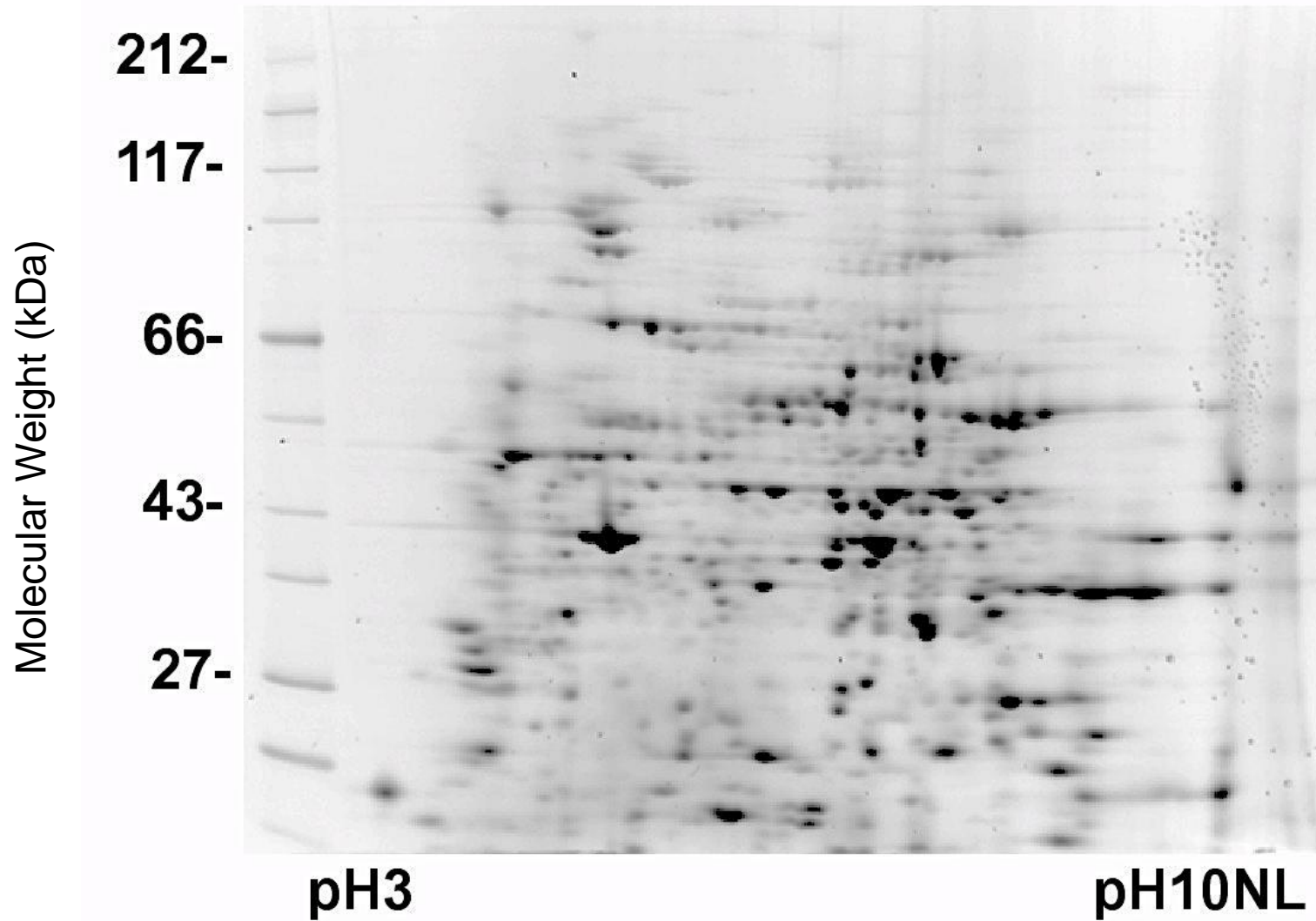
Step 1: Isoelectric Focusing (IEF)



Step 2: SDS-PAGE



2-D PAGE



Now you are able to:

- ✓ Describe most common methods of protein isolation and purification
- ✓ Compare between different methods of protein purification
- ✓ Construct a purification algorithm based on your knowledge in protein purification