Course number: CLS212	Course title: Medical Microbiology
Level/semester:	3 rd level /2 nd semester (Non-CLS Students)
	Thereof lecture hours: 3 hrs
Credit hours: 4 hrs	Thereof practical hours: 1hrs
Language:	English language
Aims and goals/ skills of the course:	This is a general medical microbiology course intended for students outside the department of Clinical Laboratory Sciences. The structure of the course is based on presenting the fundamentals of microbiology to include structures, morphology and classification of bacteria, viruses, fungi and parasites. The students will be introduced to the pathogenesis of the various infectious agents. The course will also cover some topics related to community health, including the modes and sources of infections as well as prevention of these infections.

	<u>CLS 212:</u>	Lectures Outline
	Weeks ^{1.}	Subjects General introduction Historical Background & classification of microorganisms
	2.	Introduction to Viruses Structure and morphology of viruses Classification / Replication / Pathogenicity
	3.	Introduction to Fungi Structure and morphology of fungi Classification / Fungal diseases
	4.	Introduction to Parasites Classification /General characteristics of protozoa Medically important protozoa General characteristics of helminths/Medically important helminths
Content of the course:	5.	Introduction to Bacteria Classification / Morphology Bacterial Structures / Bacterial replication
	6.	Bacterial growth / Growth curve Factors affecting growth
	7.	Antimicrobial agents
	8.	Microbial control / Principles Physical and chemical methods
	9.	Microbial control – Chemical agents Hospital acquired infections
	10.	Pathogenicity of infectious diseases Normal microbial flora
	11.	Upper respiratory tract infections
	12.	Lower respiratory tract infections
	13.	Wound and skin infections
	14.	Sexually transmitted diseases
	15.	Food borne diseases, Water borne diseases
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	written & Practical examination	
	1 st Mid Term Examination: Written	15
	2 nd Mid Term Examination: Written	15
Examination:	Mid Term Examination: Practical	10
Assessments:	Final Practical Examination:	20
	Final Theoretical Examination:	40
	 Total:	100
	Final Theoretical Examination:	40

Course number:CLS 221	Course title: Basic Anatomy & Physiology		
Level/semester: e.g. 2 ^{nc}	Semester/ 3 rd Level		
	Thereof lecture hours: 4		
Credit hours:	Thereof practical hours: 0		
Language: English			
Aims and goals/ skills of the course:	To acquaint the student with a comprehensive knowledge about the anatomy and physiology of the human body that help him to build his background information in the basic sciences that will enable him to understand the deviation from the normal in the different specialities that he/she may be directed to later on.		

1. Introduction to Anatomy and Physiology: - Body orientation, planes and sections, surface anatomy, body cavities and terminology, organ systems: an overview		Weeks	Subjects
 Body orientation, planes and sections, surface anatomy, body cavities and terminology, organ systems: an overview Body fluids and its distribution, mechanism of transport across the cell membrane, structure of the cell membrane. Cell and Tissues: Cell and Tissues: Structure and function of cell, Cell division Body fluids with and Body Membrane: Structure and function of sell, Cell division Basic structure and function of sell, Appendages Classification of body membranes State structure and function of skin, Appendages Classification of body membranes Stateletal system: Functions, bone markings and classification of bones, structures of bones, Applied aspects Nervous System: Cranial and spinal acrost cost of bones and spinal cord. CSF Central Nervous System: Cranial and spinal nerves autonomic Nervous tissue – Neuroglia, neuros classification and their microscopic structure physiology – Nerve impulses Central Nervous System: Cranial and spinal nerves autonomic Nervous System- Introduction, Applied aspects Peripheral Nervous System: Cranial and spinal nerves autonomic Nervous System: - Heart structure, External features Chambers, valves, pulmonary and systemic circulator Major arteries and veins of the body, special circulation Blood pressure, Pulse Applied aspects Physiology - Mechanics of respiratory gases and their insport, regulation of respiratory gases and their tansport, regulation of respiratory gases and their tansport, regulation of respiration - Applied aspects Organs of alimentary canal, mouth, pharynx, oesophagus, stomach, small and large			
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deferens, Accessory glands,External genitalia			
- Nerve and Muscle Physiology:			 Nerve and Muscle Physiology:
- Structure of the nerve, generation of action potential			
(nerve impulse), conduction of nerve impulse, types of nerves			(nerve impulse), conduction of nerve impulse, types of nerves
- Structure, types of muscles, mechanism of muscle			
			conduction, types of muscle contraction and concept of
13. Blood Physiology:		13.	
 Composition of the blood Function of the different blood cells and the plasma 			
- The process of haematopoesis			

Examination:	e.g. written examination, presentation First Mid Term Examination: 30 Second Mid Term Examination: 30 Final Theoretical Examination: 40	
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4 th level / 2 nd Semester
Thereof lecture hours: 2
Thereof practical hours: 2
The overall objective of this course is to prepare the students for the advanced courses in CLS and enables the student to identify microscopically the normal tissues and organs of the human body.
Weeks Subjects 1. Introduction 2-3. Epithelial tissue:(definition, features, classification & function) 4. Connective tissue (definition, features, cells and fibers) Connective tissue types: (loose, dense, reticular, adipose and elastic) 5. Supportive connective tissue, Cartilage (general features, functions and types: hyaline, elastic, white fibrocartilage), Bone (general features, functions, types: spongy, compact and development) 6. Muscular tissue (general features and types) 7. Nervous tissue, Central nervous system (parts), types and structure of nerves and ganglia, Nerve endings, Synapse ,Neuroglia 8. Circulatory system, Blood circulation, Heart structure and function Arteries and veins (types, structure and comparison of both) Comparison between capillaries and sinisoids Lymph circulatory system: lymph vessels 9. Lymphatic organs, Lymph nodes: structure of the lung, Pleura 11-12. Digestive system, Oral cavity: lip, tongue (papillae and its function) Digestive tube: general structure of esophagus, stomach, small and large intestine, appendix 13. Digestive glands: Salivary gland, liver, gall bladder and pancreas 14. Urinary system, Parts, function and structure of kidney and urinary passage (ureter, urinary bladder and urethra in males and females)
written & practical examinationFirst Mid Term Examination:15Second Mid Term Examination:15Student activities, sharing, Quiz10Final Practical Examination:20Final Theoretical Examination:40

Course number: CLS 223	Course title: General Pathology		
Level/semester: 4			
	Thereof lecture hours: 2 hour		
Credit hours: 2 hours	Thereof practical hours:0 hour		
Language: English			
Aims and goals/ skills of the course:	To recognize the causes of cellular stress and the general pathologic responses to cellular stress, the circulatory disturbances, the basic immune reactions, the main immunopathologic processes and disorders of growth and neoplasia.		

	1. TOPICS to be COVERED/ WEEK	Contact hours	W
	Introduction,Define pathology,Define disease , study of disease Predisposing and determining factors,Pathological anatomy (lesion) Manifestation of the disease,Pathological physiology,Diagnosis	4	
	Causes of cell damage (stress) Cellular response to stress Degeneration Necrosis and gangrene	2	v
	<u>Inflammation</u> (definition, nomenclature and causes) Pathogenesis, local vascular changes of acute inflammation The inflammation exudates, inflammatory cells and chemical mediators and sequelae of acute inflammation special types of acute inflammation types of chronic inflammation general effects of inflammation	4	
	Infectionfactors influencing establishment of infectionfactors influencing the course of infectionmechanism by which disease is produced: bacterial, viral andopportunistic infectionsHealingwound healingfactors influencing wound healingcomplicationshealing in special situations e.g. kidney, liver muscle, nervoustissue and bone	2	
Content of the course:	!st Mid Term Exam	1	W
	<u>Circulatory Disturbances</u> hyperemia congestion obstruction causes: thrombosis, embolism effect of vascular obstruction leading to oedema heamorrhage	4	
	Disturbances of growthHyperplasia, hypertrophy, atrophy, dysplasia and metaplasiaNeoplasiadefinition and classificationsgeneral features of benign and malignant tumorseffects and complication of benign and malignant tumorstumor of epithelial, connective, muscular and nervous tissuescarcinogenesisbrief discussion on diagnosis of tumors and treatment	4	W
	2 nd Mid Term Exam	1	W
	Immunology specific and non-specific immunity immune response, antigen, antibodies, compliment, humoral cell mediated responseDiseases of the immune system hypersensitivity reactions auto-immune disease, tolerance immune deficiency diseases	4	WW
	Assignment	2	W
	Final Theoretical examination		W

	First and second midterm examinations (50%), Final theory examination
Examination:	(40%)and Assignments (10%)

Course number: CLS 231	Course title: Clinical Analytical Chemistry			
Level/semester: 4th				
	Thereof lecture hours: 2 hour			
Credit hours: 3 hours	Thereof practical hours: 1 hour			
Language: English				
Aims and goals/ skills of the course:	This course involves a review of func- analysis as applied to biological fluids. Th of different types of titri-metric analysis in and precipitation titrations are studied.	e theoretica	al and pract	ical aspects
Content of the course:	Topics	Planned Contact Hours	Actual Contact Hours	
	General Introduction/The analytical process (2 weeks)	4	4	
	Review of Fundamental concepts (1 week)	2	2	
	Acid Base theories and Ionisation of acids and bases (2 weeks)	2	2	
	Acid Base titrations (1 week)	4	4	
	Buffers and the pH scale (1 week)	2	2	
	Complexo metric Reactions and Titrations (1 week)	2	2	
	Precipation Reactions and Titrations(1 week)	2	2	
	Potentiometric Electrodes (1 week)	2	2	
	Redox titrations (1 week)	2	2	
	Gravimetric analysis of calculations (1 week)	2	2	
	Introduction to instrumental analysis	4	4	

Level/semester 4 th level / 2 th Semester Credit hours: Thereof lecture hours: 3 Thereof practical hours: 1 Language: English Aims and goals/ skills of the course: This course illustrates structures and functions of bimolecules which include carbohydrates, proteins, lipids, nucleic acids, enzymes and hormones. In the practical part of this course, students study the detection and quantitative determination of some of these biomolecules. Weeks Subjects 1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Wittern Examination: First Mid Term Examination: Examination: 15 Second Mid Term Examination: 5 5 Laboratory Reports: 5 Laboratory Reports: 5	Course number: CLS 232	nber: CLS 232 Course title: Clinical Biochemistry (1)			
Credit hours: Thereof practical hours: 1 Language: English Aims and goals/ skills of the course: This course illustrates structures and functions of bimolecules which include carbohydrates, proteins, lipids, nucleic acids, enzymes and hormones. In the practical part of this course, students study the detection and quantitative determination of some of these biomolecules. Weeks Subjects 1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8.9. Carbohydrates 10,11. Faty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15. Geordendite Examination: 5 Laboratory Reports: 5 Final Practical examination: 5 Laboratory Reports:					
Thereof practical hours: 1 Language: English Aims and goals/ skills of the course: This course illustrates structures and functions of bimolecules which include carbohydrates, proteins, lipids, nucleic acids, enzymes and hormones. In the practical part of this course, students study the detection and quantitative determination of some of these biomolecules. Weeks Subjects 1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8.9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination; First Mid Term Examination: 15 Second Mid Term Examination: 15 Courses: 5 Final Practical Examination: 15 Courses: 5 Final Practical Examination: 20		Thereof lec	ture hours: 3		
Language: English Aims and goals/ skills of the course: This course illustrates structures and functions of bimolecules which include carbohydrates, proteins, lipids, nucleic acids, enzymes and hormones. In the practical part of this course, students study the detection and quantitative determination of some of these biomolecules. Weeks Subjects 1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8.9. Carbohydrates 10,11. Faty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 5 Final Practical Examination: 15	Credit hours:				
Aims and goals/ skills This course illustrates structures and functions of bimolecules which include carbohydrates, proteins, lipids, nucleic acids, enzymes and hormones. In the practical part of this course, students study the detection and quantitative determination of some of these biomolecules. Weeks Subjects 1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10.11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Second Mid Term Examination: 15 Suizzes: 5 Laboratory Reports: 5 Second Mid Term Examination: 20		Thereof pra	actical hours: 1		
Aims and goals/ skills of the course: carbohydrates, proteins, lipids, nucleic acids, enzymes and hormones. In the practical part of this course, students study the detection and quantitative determination of some of these biomolecules. Weeks Subjects 1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination. Second Mid Term Examination. 15 Quizzes: 5. Second Mid Term Examination: 5. Second Mid Term Examination: 5. Laboratory Reports: 5 Laboratory Reports:	Language: Er	nglish			
1. Acids, bases, pH scale, buffer system 2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Laboratory Reports: 5 Laboratory Reports: 5		In the practical part of this course, students study the detection and			
2. Chemical bonds 3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Cuizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		Weeks	Subjects		
3. Amino acids: structure, physical and chemical properties 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		1.	Acids, bases, pH scale, buffer system		
Content of the course: 4. Peptide bonds, proteins: types, structure, functions 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination: 15 Second Mid Term Examination: 15 Second Mid Term Examination: 15 Second Mid Term Examination: 15 Final Practical Examination: 20		2.	Chemical bonds		
Content of the course: 5. Protein purification 6. Nucleotides and nucleic acids 7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: First Mid Term Examination: 15 Second Mid Term Examination: 15. Second Mid Term Examination: 16. Second Mid Term Examination: Second Mid Term Examination: 17. Second Mid Term Examination: Second Mid Term Exam		3.			
Content of the course: 6. Nucleotides and nucleic acids 6. Fazymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		4.			
7. Enzymes 8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Second Mid Term Examination: 15 Laboratory Reports: 5 Laboratory Reports: 5 Final Practical Examination: 20	Content of the course:	5.	Protein purification		
8,9. Carbohydrates 10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		6.	Nucleotides and nucleic acids		
10,11. Fatty acids and lipids 12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		7.	Enzymes		
12. Hormones 13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		8,9.	Carbohydrates		
13. Vitamins Written & practical examination, First Mid Term Examination: 15 Second Mid Term Examination: 15 Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		10,11.	Fatty acids and lipids		
Written & practical examination, First Mid Term Examination: Second Mid Term Examination: Quizzes: 5 Laboratory Reports: 5 Final Practical Examination: 20		12.	Hormones		
First Mid Term Examination:15Second Mid Term Examination:15Quizzes:5Laboratory Reports:5Final Practical Examination:20		13.	Vitamins		
First Mid Term Examination:15Second Mid Term Examination:15Quizzes:5Laboratory Reports:5Final Practical Examination:20		Written & nr	ractical examination		
Examination:Quizzes:5Laboratory Reports:5Final Practical Examination:20	Examination:	First Mid Terr	m Examination: 15		
Final Practical Examination: 20		Quizzes:	5		
Final Theoretical Examination: 40					

Course number: Course title: CLS 241: Haematology					
Level/semester: level 4- second semester					
	Thereof lecture hours: 2				
Credit hours:	Thereof practical hours: 2				
Language: English					
Aims and goals/ skills of the course:	a. To recognize the normal cellular, properties and functions of haemopoietic cells b.To understand the mechanisms of pathophysiology of different haemopoietic diseases c. to recognize different forms of red blood cell abnormalities, white blood cell abnormalities and platelet and clotting factors abnormalities d- understanding and on hand training of clinical laboratory techniques use for diagnosis of hematlogical disoders.				
Content of the course:	Introduction to haematology, production of blood cells Erythropoiesis and description of normal red blood cells Granulocytes, Monocytes, and reticuloendothelial system Lymphocytes and Benign disorders of white cells Red cell disorders: Anaemia- general concepts Iron: Physiology and deficiency, iron Overload and sideroblastic anaemia Mid Term Exam (Theory and Practical) Megalohlastic anaemia: Vitamin B12, and Folate deficiency Haemoglobiopathies-sickle cell disease Haemoglobiopathies-Thalassaemia leukaemia-General concepts Acute and chronic leukaemias Myeloproliferative disorders Bleeding disorders				
Examination:	 Mid term theoretical exam: Multiple choice questions Mid term practical exam Written assignment Final practical exam Final theoretical exam 				

Course number:CLS 311	Course title: Basic Microbiology		
Level/semester:	5 th level /2 nd semester		
	Thereof lecture hours: 3 hrs		
Credit hours: 4 hrs	Thereof practical hours: 1hrs		
Language:	English language		
Aims and goals/ skills of the course:	This course provides the students with basic theoretical and practical aspects of various groups of microorganisms to include bacteriology, virology, mycology, and parasitology as well as basic concepts of immunology and epidemiology. It also introduces the basic concepts of disinfectants, antiseptics, preservatives, Ames test, methods of sterilization, aseptic techniques and general microbial control.		

CLS 311: Lectures Outline

Weeks	Subjects
1.	General introduction to Microbiology
	Historical background and Classification of
	Microorganisms
2.	Introduction to Viruses:
	Classification, morphology and structure, Replication a
	Pathogenicity
3.	Introduction to Fungi:
	Classification, morphology and structure, Replication
	Pathogenicity
4.	Introduction to Parasites:
	Classification, General Characteristics of parasites and
	Medically important Parasites
5.	Introduction to Bacteria:
	Classification, Morphology and Structures
6.	Bacterial Structures
7.	Microbial Growth requirements:
	Nutritional requirements, Physical requirements,
_	Types of bacteria according to energy production
8.	Bacterial Growth:
_	Growth curve, Constant and synchronous growth
9.	Bacterial Metabolism: Catabolic Pathways and regulation
10.	Bacterial Genetics:
	Mutation and mutagenic agents ,&Ames test
4.4	Gene transfer ,PCR ,& genetic engineering
11.	Bacterial Genetics continued
12.	Microbial Control:
	Principles, Disinfectants, antiseptics, and preservatives
	Physical and chemical methods of sterilization Aseptic
13.	techniques Microbial control continued…
	Pathogenicity of Infectious Diseases
4.4	Pathogenicity of infectious Diseases
14. 15.	Normal Microbial flora

Content of the course:

written and practical examination			
1 st Mid Term Examination: Written	15		
2 nd Mid Term Examination: Written	15		
Mid Term Examination: Practical	10		
Final Practical Examination:	20		
Final Theoretical Examination:	40		
Total:	100		
	1 st Mid Term Examination: Written 2 nd Mid Term Examination: Written Mid Term Examination: Practical Final Practical Examination: Final Theoretical Examination:	1 st Mid Term Examination: Written 15 2 nd Mid Term Examination: Written 15 Mid Term Examination: Practical 10 Final Practical Examination: 20 Final Theoretical Examination: 40	

Course number:CLS 31	12 Course title: Clinical Mycology				
Level/semester 6 th level / 2 nd Semester					
	Thereof lecture hours: 2				
Credit hours:	Thereof practical hours: 1				
Language: English					
Aims and goals/ skills of the course:	In this course the students will learn about the fungi (molds and yeasts) of medical importance and the diseases they cause. The classification, structure physiology, and cultural characteristics of fungi will be discussed. Emphasis in this course will be on the fungal diseases and their clinical presentation, pathogenesis, modes of transmission, laboratory diagnosis, prevention and control.				

	Weeks	Subjects
	1.	Introduction to Mycology. Structure and morphology of fungi
	2.	Fungal classification and taxonomy
	3.	Superficial mycosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	4.	Dermatophytosis : Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	5.	Mycetoma: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	6.	Chromoblastomycosis, Phaeohyphomycosis, Sporotrichosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	7.	Zygomycosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
Content of the course:	8.	Lobomycosis, Rhinosporidiosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	9.	Aspergillosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	10.	Candidiasis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	11.	Cryptococcosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	12.	Trichosporonosis, Geotrichosis Pneumocystosis: Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	13.	Primary Systemic Fungal Infection: Blastomycosis, Histoplasmosis - Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
	14.	Primary Systemic Fungal Infection Coccidioidomycosis, and paracoccidioidomycosis - Definition, symptoms, epidemiology, etiology, laboratory diagnosis and treatment
Examination:	written, practical, presentation examination,Mid Term Examination:15Continuous evaluation5Laboratory Notebook:10Final Practical Examination:30Final Theoretical Examination:40	

Course number:CLS 321	Course title: General Pathology			
Level/semester	5 th level / 2 nd Semester			
	Thereof lecture hours: 2			
Credit hours:	Thereof practical hours: 1			
Language: En	glish			
Aims and goals/ skills of the course:	Upon completion of this course, the students will have the appropriate knowledge about the causes of cellular stress and the general pathologic responses to cellular stress, the circulatory disturbance, the basic immune reactions, the main immunopathologic processes, Disorders of growth and neoplasia.			

	Weeks	Subjects
	1.	Introduction
		- Define pathology
	2.	- Define disease, study of disease
	2.	Etiology Predisposing and determining factors
		- Pathological anatomy (lesion)
		- Manifestation of the disease
		 Pathological physiology Diagnosis
	3.	Causes of cell damage (stress)
		Cellular response to stress
		Degeneration
	4-5.	Necrosis and gangrene Inflammation (definition, nomenclature and causes)
	+-0.	Pathogenesis, local vascular changes of acute inflammation
		The inflammation exudates, inflammatory cells and
		chemical mediators and sequelae of acute inflammation
		 special types of acute inflammation types of chronic inflammation
		general effects of inflammation
	6.	Infection
		 factors influencing establishment of infection factors influencing the course of infection
		- mechanism by which disease is produced: bacterial,
		viral and opportunistic infections
	7.	Healing
		 wound healing factors influencing wound healing
		- complications
		 healing in special situations e.g. kidney, liver muscle,
Content of the course:	8-9.	nervous tissue and bone Circulatory Disturbances
Content of the course.	0.01	- hyperemia
		- congestion
		 obstruction causes: thrombosis, embolism
		 effect of vascular obstruction leading to oedema
		- heamorrhage
	10-11.	Immunology specific and non-specific immunity
		- immune response, antigen, antibodies, compliment,
		humoral cell mediated response
		Diseases of the immune system - hypersensitivity reactions
		- auto-immune disease, tolerance
		- immune deficiency diseases
	12.	Disturbances of growth - hyperplasia
		- hypertrophy
		- atrophy
		- dysplasia - metaplasia
	13-14.	Neoplasia
		 definition and classifications
		 general features of benign and malignant tumors efforts and complication of bonign and malignant
		 effects and complication of benign and malignant tumors
		- tumor of epithelial, connective, muscular and nervous
		tissues
		 carcinogenesis brief discussion on diagnosis of tumors and treatment

	Written & practical examination:
Examination:	First Mid Term Examination:15Second Mid Term Examination:15Laboratory Quizzes:5Mid Term Practical Examination:5Final Practical Examination:20Final Theoretical Examination:40

Course number: CLS 3	Course title: Histological Techniques	
Level/semester	6 th level / 2 nd Semester	
	Thereof lecture hours: 1	
Credit hours:	Thereof practical hours: 1	
Language: English		
Aims and goals/ skills of he course:	This course will provide the student with the basic knowledge in the techniques of tissue preparation, staining and examination under the light microscope. Knowledge will be focused mainly on the paraffin technique; however, other histological methods will be covered. Also, the student will be instructed how to use the light microscope properly. An idea about other types of microscope will be provided.	

	Weeks	Subjects
	Weeks	Subjects
	1.	Introduction to Microtechniques
		Lab tools, rules & safety
	2.	Different histological methods (paraffin, celloidin, cryostat)
		Fresh unfixed tissue; teased, squashed, smear, impression,
		frozen techniques
		Tissue processing (manual, automated)
	3.	Upon obtaining the specimen
		Narcotization
		Steps of tissue preparation
	4.	Fixation (purpose, effects, methods, types)
	5.	Dehydration and clearing
	6.	Infiltration and embedding
		Types of wax, other embedding media
Content of the course:	7.	Sectioning: 1) Microtome, Types, components,
		techniques, and care
		2) Microtome knife; types, sharpening3) Sectioning
		Affixation and mounting
	8.	Staining: Definitions
		Theory, mechanism, Types,
	9.	Principle of light microscope,
		Theory of magnification & resolution
		Components, Definitions
	10.	Dark field microscope,
		Fluorescent microscope
	11.	Polarized microscope
		Phase contrast microscope
	12.	Discussion
	13.	Revision
		tical examination,
Examination:	First Mid Term E Second Mid Terr	
	Final Practical E	xamination 30
	Final Written Exa	amination 40

	Course number: CLS 3		e title: General Pathophysiology	
	Level/semester	vel/semester 5 th level / 2 nd Semester		
		Thereof lecture hours: 2		
	Credit hours:	Thereof pra	actical hours: 0	
	Language: English			
Aims and goals/ skills of the course:		At the completion of this course the student will know the normal structure and functions of the body systems studied. Student should be able to understand the pathophysiology of the most common diseases, its manifestations and the diagnostic methods used.		
		Weeks	Subjects	
		1,2,3,4.	 Cardiovascular System Physiology and Manifestations Congenital Heart Disease, Valvular Disease, Rheumatic disease Vascular Disease, Atherosclerosis, Ischemic Heart disease Hypertension, Hypotension, Heart Failure 	
		5,6,7,8.	Respiratory System Physiology and Manifestations Pneumonia, Tuberculosis Obstructive Disease Respiratory Failure 	
Con	tent of the course:	9.	Female Genital System - Physiology and Manifestations - Causes of Abnormal Uterine Bleeding, Toxemia	
		10,11,12.	Urinary System	
		13.	Digestive System - Physiology and Manifestations - Common diseases of the digestive system	
		14,15.	Endocrine System - Physiology and Manifestations - Common endocrine diseases	
		Written examination,		
Exar	nination:	First Mid Term Examination:20Second Mid Term Examination:20Presentation20Final Theoretical Examination:40		

Course number: CLS 32	24 Course ti	tle: Principles of Electron Microscopy	
Level/semester	6 th level / 2 nd	^d Semester	
	Thereof lecture hours: 1 Thereof practical hours: 1		
Credit hours:			
Language: E	nglish		
This course will provide the student with the basic knowledge in the theory of magnification and resolution, knowledge about the structure of the electron microscope and its operation, techniques of tissue processing; preparation, and staining. Knowledge will be focused mainly on the avoidance of the artifacts. Students will be informed about some special techniques, scanning electron microscope and dark room.			
	Weeks	Subjects	
	1.	Introduction to electron microscopic unit	
		Rules & safety and administration	
	2.	Lab orientation, Component of the unit	
		Unit precautions	
		Equipments, Lab tools	
		machines, rules and principles	
	3.	Theory of magnification and resolution	
	4.	Structure of the electron microscope; electron gun,	
Content of the course:		electromagnetic lens, comparison between LM & EM	
	5.	Operation of the EM	
	6.	Processing of a specimen to TEM	
		Obtaining a specimen, fixation, dehydration and clearing	
	7.	Infiltration, embedding and sectioning	
	8.	Artifacts, definition, types, avoidance	
	9.	,chatter, knife marks	
	10.	Spécial techniques, perfusion fixation, decalcification	
	11.	Scanning electron microscope, SEM versus TEM	
	12.	Dark room	
	Written & prac	tical examination	
Examination:	First Mid Term Second Mid Ter Laboratory Quiz Final Practical E Final Theoretica	rm Examination: 15 zzes: 10 Examination: 20	

Course number: CLS 3	er: CLS 331 Course title: Clinical Biochemistry (II)		
Level/semester	5 th level / 2 nd Semester		
	Thereof lecture hours: 3		
Credit hours:	Thereof practical hours: 1		
Language:	English		
ms and goals/ skills of e course:	Course work involves a study of the metabolic activity of animal tissue and its regulation. Major catabolic pathways related to carbohydrate, amino acids, and fatty acids are discussed in detail. the study includes a look at the bioenergetics of cells as well as the digestion and absorption of food material.		

	Weeke	Subjecto
	Weeks	Subjects
	1.	General introduction - Metabolic pathways
		- Catabolism and Anabolism
		- Regulation of metabolic activity
	2.	Digestion and Absorption
		- Photometry
		- Role of vitamins in metabolism
	3.	Bioenergetics
		- The role and chemistry of ATP
		 Standard free energy change of biochemical reactions
	4.	Glycolysis
		- Regulatory enzymes of glycolysis
Content of the course:	5.	The citric acid cycle
Content of the course.		- Regulation of carbohydrate catabolism
	6.	The Electron Transport chain & oxidative
		phospharylation
		- The energetic value of glucose
	7.	The pentose phosphate shunt and gluconeogenesis
	8.	Glycogen metabolism
		- The glycogen storage diseases
	9.	Oxidative degradation of amino acids
		- The synthesis of urea
	10.	Introduction to Inborn Errors of Metabolism
	11.	Oxidation of fatty acids in animal tissue
		- B – oxidation pathway
	12,13.	Hormones and their metabolic regulatory role
	Written & pr	actical examination
		n Examination: 15
Examination:		erm Examination: 15
LaboratoryWork and Reports: 15 Final Practical Examination: 15		
		cal Examination: 40

Course number: CLS 3	332 Course title: Instrumental Analysis		
Level/semester 5 th level / 2 nd Semester			
	Thereof lecture hours: 3		
Credit hours:	Thereof practical hours: 1		
Language:	English		
ns and goals/ skills of e course:	The knowledge of the principles of instrumentation is essential for a clinical laboratory technologist which can be utilized data. A good understanding of the principles of instrumentation along with the fundamentals of electronics applicable to laboratory		

instruments of the future.

instruments is offered in the course. This will prepare the laboratory technologist to adopt analytical procedures to the

	Weeks	Subjects
	1.	Introduction
		- Electromagnetic radiation, Absorption spectroscopy
		- Spectrophotometric techniques, Application of Beer's
		law, Visible absorption spectrophotometer,
		 Components and applications
	2.	Atomic absorption spectrophotometer
		- Principle, basic components and their function
		- Flamless atomic absorption spectrophotometer
		Advantages, interferences
	3.	Emission spectroscopy
		- Luminescence, Fluorometers
		- Fluorometers v/s conventional spectrophotometers
		 Instrumentation, Possible disadvantages, solutions Special application of fluorometers
	4.	Light scattering phenomenon
	7.	- Turbidity and nephlometery
		 Nephlomter v/s spectrophotometer
	5.	Flame photometry
	0.	- Internal standard in flame photometry
		- Advantages/disadvantages
	6.	Osmometry
		- Osmosis, colligative propertiesOsmolality/osmolarity
		- Osmometers, Freezing point osmometers
		- Vapor presure osmometers
	7.	Ion selective electrodes
		 Types of electrodes, principle and function
		 Reference electrodes, Glass pH electrodes
		- Precipitate electrodes, Solid state electrodes
Content of the course:		- PCO2 and PO electrodes
Content of the course.	8.	Electrophoretic techniques
		- Introduction, Buffers, support materials
		 Endosmosis, Factors affecting the migration rate Clinical applications of electrophoresis
	9.	Chromatographic techniques
	0.	- General principles, Physical basis of separation
		- Adsorption (liquid- solid)
	10.	Partition (liquid- liquid)
		- Gel filtration (steric exclusion)
		 Ion exchange chromatography
	11.	Thin layer chromatography
		High performance liquid chromatography
	10	- Gas chromatography
	12.	Automated procedures
		 Basic concepts, operation, trouble shooting Automatic disponsors (dilutors), Types of automated
		equipment, Continuos flow analyzers Discrete
		analysers
	13.	Single channel instruments
		- Discontinious operation
		 (Batch processing, single specimen)
		- Multichannel instruments
		- Small instrumentssix channels
		- Large instrumentsnon discretory, discretory
		- Contrifugal analysers

Examination:	Written & practical examinationFirst Mid Term Examination:15Second Mid Term Examination:15Laboratory Work and Reports:15Final Practical Examination:15Final Theoretical Examination:40

Course number: CLS 333	Course title: . Clinical Biochemistry I	II				
Level/semester: 6						
L	Thereof lecture hours: 3 hour	Thereof lecture hours: 3 hour				
Credit hours: 4 hours	Thereof practical hours: 1 hour					
Language: English	<u> </u>					
Aims and goals/ skills of the course: This is a required course in biochemistry for preparing a generalist in medical technology program. The course will provide a useful application of a basic biochemistry along with physiological chemistry in developing an appreciation of the clinical significance of the test methodology, and interpretation of test results.				pplication of veloping an		
	Topics	Planned Contact Hours	Actual Contact Hours			
	Disorders of carbohydrate metabolism (2 weeks)	6	6			
	Disorders of aminoacid metabolism (1 weeks)	3	3			
	Protein in bodyfluids (1 weeks)	3	3			
	Disorders of lipid metabolism (1 weeks)	3	3			
	Liver function and Liver Diseases (1 week)	6	6			
Content of the course:	Biochemical tests of liver function (1 week)	3	3			
	Electrolytes of blood gases (1 week)	3	3			
	Renal function and Renal Diseases (2 week)	6	6			
	Biochemical Tests of Renal Function (1 week)	3	3			
	Clinical Enzymology (1 week)	3	3			
	Gastrointestinal Function and diseases (1 week)	3	3			
	Endocrine Function Tests (1 week)	3	3			
	Ovarian/Testicular Function Tests (1 week)	3	3			
Examination:	Midterm examination (15%), Reports (15%) and and Lab sessions (15%) and Final lab examina		en examinati	on (40%)		

Course number: CLS 41		e title: Clinical Bacteriology I
Level/semester: Level	7 / both sem	esters
	Thereof lec	ture hours: 2
Credit hours:	Thereof practical hours: 1	
Language: English		
Aims and goals/ skills of the course:	chemotherap susceptibility The second relationships, positive cocc bacilli, and th presentation,	t of this course deals with the theoretical and practical aspects o eutic agents, their modes of action, methods of evaluation testing, and mechanisms of microbial resistance. part of the course describes the mechanisms of host parasite followed by a detailed study of the structure and physiology of Gram i and Gram negative cocci, the aerobic spore and non-spore forming e Mycobacteria. The diseases caused by these organisms, the clinica pathogenesis, modes of transmission, laboratory diagnosis, antibiotic testing, prevention and control will be fully discussed.
	Weeks	Subjects
	1.	Chemotherapeutic agents / Antibiotics – Definition, Modes of action, methods of evaluation
	2.	Antibiotics cont mechanisms of resistance, Susceptibility testing
	3.	Staphylococci and Micrococci
	4.	Streptococci – The Beta haemolytic streptococci
	5.	Alpha haemolytic Viridans streptococci, &Streptococcus pneumoniae Non-haemolytic streptococci (Enterococci)
	6.	Neisseria , Moraxella, Kingella, Eikenella & Acinetobacter
Content of the course:	7.	Aerococcus, Peptococcus, Veillionella, and Gemella
	8.	Corynebacteria
	9.	Listeria, Erysipelothrix, Lactobacilli, Kurthia
	10.	Mycobacteria – M. tuberculosis and M. leprae
	11.	The Atypical Mycobacteria
	12.	Actinomycetes & Nocardia
	13	The aerobic spore bearers – Bacilli Bacillus anthracis, Bacillus cereus and other spore forming- bacilli
	14.	Fastidious gram negative rods: Legionella, Francisella ,,Gardnerella, Pasteurella and Calymmatobacterium granulomatis
Examination:	written exan	nination, home work reports, practical examination 30%

Course Description Prof.Mohd Zahedi Daud

Course number: CLS412	Course title: Medical Parasitology	
Level/semester:	Semester (7) Year (4)	
Credit hours: 4	The Lecture hours: 3 X 15 = 45 hours The Practical hours: 2 X 15 = 30 hours	
Language : English		

Aims and goals/ skills of the course:	 a) Students should be able to appreciate the parasites' life-cycle, their behavior in the environment, and the disease they caused. Basic biochemistry and host specificity, mechanisms of evasion of host's immune responses are discussed. b) Students are exposed to drugs commonly used against human parasites, and disease control strategies. The epidemiology of the disease and the control strategies are discussed in the context of the local environment. c) Students are to be competent public health workers, and should be able to advise the public in control strategies against parasitic diseases. d) Current issues in medical parasitology are introduced; topics cover new emerging diseases and the impact/implication these diseases on public health. e) Students are encouraged to do text-book-based or IT-based self-studies. g) Students do laboratory (practical work) sessions; they are exposed to permanent mount parasites or in fresh biological specimens. Studenst are also taught how to isolate parasites and to do simple diagnostic test techniques. They are trained to be competent laboratory personnel, able to isolate, process and stain, and identify human parasites of importance thus at the end of the day: Students are exposed to various species of human and animal parasites of public health importance, their life cycles and the discases they caused. The epidemiology of these parasitic isolate, mount and identify most of the common parasites at the species level. Parasites isolation and identify most of the common parasites at the species level.

Content of the course:	Teaching and learning processes are by formal lectures, tutorials, practical laboratory sessions, and supplemented with self-study and group discussions. Students are regularly assessed by lecture-room quizzers. Small group discussions, and informal meetings with the lecturer and laboratory staff further enhance the teaching and learning strategies. Course Content Introduction to Parasitenogy • general parasite morphology and biology • general parasite and the environment • parasite endidene wirromment • parasite endidene wirromment • intestinal protozoa eg Entamoeba histolytica • intestinal protozoa eg Entamoeba histolytica • intestinal romatodes ca Acaris lumbricoides • intestinal romatodes ca Castones eg flukes and tapeworms Parasites of the Digestive system • biolod nematodes eg Acaris lumbricoides • intestinal trematodes eg Russandium spp. • biolod nematodes eg Glustan eg Subset • biolod nematodes eg Glustan eg Subset • parasites of the Xueuo-Skeletal System • biolod nematodes eg Biasnodium spp. • nematodes eg Takina Soyne • nematodes eg Takina Soyne • protozoa eg Vestionson spp. • nematodes eg Treisonom spp. • nematodes eg Treisonom sopne • protozoa eg Subsitionson spp. • nematodes eg Treisonom sopne • protozoa eg Treisonoma hematobium Parasites of the Nervous System • protozoa eg Treisonoma hematobium Parasites of the Regraminus Netternani Parasites of the Seg Sophastan gondii • nematodes eg Jarvial stages of solit ransmitted helminths • cestodes eg Javahi de syste of Fachinococcus sp. • protozoa eg Subpassional desades eg Gruppeniolesis • outrof strategies Zoonotic Diseases (
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	Examination: Quizzes once every forthnight 6%
	Mid-Semester Examination 18%
	Final Semester Examination 40%
	Assignments (2 term papers) 6%
Examination:	Practical Examinations 30%
	Examinations papers consisted of MCQs, True/False, Fill in Blanks and Short Essays

Course number: CLS 413	Course title: Clinical Bacteriology (II)		
Level/semester 8 th level / 2 nd Semester			
	Thereof lecture hou	rs: 2	
Credit hours:	Thereof practical ho	ours: 1	
Language: English			
ims and goals/ skills of ne course:	Helicopacter, Borgetella, Brucella, Haemophilus, Legionella and related		

The Enterobacteriacea: The different genera making up the group 'Enterobacteriaceae'. Their characteristics and growth identifications. Enterobacteriacea cont Medical importance of members of this group of organisms, the disease they cause Enterobacteriaceae contPathogenesis, modes of transmission, antibiotic susceptibility, prevention and control PSEUDOMONADS and glucose non-fermentors G(-)rods The small Gram negative rods: Haemophilus, Brucella, and Bordetella, and HACEK – Characteristics and growth The small Gram negative rods cont diseases they cause,
clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The curved G(-) rods, Vibrios: characteristics, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control Microaerophilic ,Species of Campylobacter and Helicobacter: Characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Spirochaetes: Treponema species, characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Spirochaetes cont Species of Borrelia, Leptospira - characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The anaerobes: Peptococcus, Peptostreptococcus, The Lactobacilli, Propionibacteria Veillonella- characteristics, species of medical importance, diseases they, cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Clostridia: The pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Clostridia: The pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Clostridia cont The Bacteroides species; characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Clostirdia cont The Bacteroides species; characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control Mycoplasma, Chlamydia ,and the Rickettsia group::
antibiotic susceptibility, prevention and control The Clostridia: The pathogenic species, characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control The Clostirdia cont The Bacteroides species; characteristics, species of medical importance, diseases they cause, clinical presentation, pathogenesis, modes of transmission, antibiotic susceptibility, prevention and control
diseases they cause. <i>practical examination</i> Examination: 20 Practical Examination: 15 s & reports 5 trical Examination: 20 pretical Examination: 40

	Course number:CLS 414	 ⁴¹⁴ Course title: Immunology 8th level /2nd semester 	
	Level/semester:		
		Thereof lecture hours: 3 hrs	
	Credit hours: 4 hrs	Thereof practical hours: 2 hrs	
	Language:	English language	
t Aims and goals/ skills of the course:		This course is designed to give a basic understanding of heoretical and practical aspects of immunology. The first part of he course deals with the basic aspects of Immunology – Types of immunity, immune response, characterization of antigens, the numoral response (different classes of antibodies), the Complement system, the cellular response (different types of T cells and their functions, the cytokines), the immune regulation. The second part of the course covers the Clinical aspects of mmunology Immunopathology, immunity to microbial nfections, types of hypersensitivity reactions, auto-immune diseases,immunodeficiency diseases, transplantation, tissue yping and rejection.	

	<u>CLS 414:</u>	Lectures Outline
	Weeks	Subjects
	1.	Introduction: history and terminology Types of immunity (natural and acquired immunity)
	2.	Innate (natural) immunity, parameters and mechanism
	3.	Antigens, immunogenicity, types of antigens & mitogens
	4.	Organs and tissue of the immune system, primary and secondary lymphoid organs
	5.	Cells involved in immune responses - types and subtypes, sites of their developments - their functions
	6.	The immune response: humoral response, primary and secondary immune response , primary and secondary immune response, mechanism of antibody production
	7.	Antibodies: structure, classes, subclasses and characters of immunoglobulins as well as their functions and uses
	8.	Monoclonal antibodies, hybridoma technology, and their applications. Anti-idiotypic antibodies and their uses
	9.	Complement system: characters, mechanism of activation,
	10.	biological functions and techniques involving complement Major Histocompatibility Complex (MHC) antigens,Human leukocytes Antigens (HLA). Cellular distribution, structure and functions of MHC antigens.
Content of the course:	11.	The cellular immune response: mechanism of cytotoxicity reactions and measurement of cytotoxicity <i>in-vitro</i> , types and functions of cytokines. Treg cells
	12.	Immunoregulation and immunoprophylaxis Clinical Immunology: Immunity to microbial infections
	13.	Immunopathology: Types of hypersensitivity (Types 1,II, III, and IV)
	14.	Transplantation and Tissue typing, mechanism of tissue rejection
	15.	Autoimmune diseases and immunodeficiency diseases

	written examination			
	1 st Mid Term Examination: Written	10		
	2 nd Mid Term Examination: Written	10		
Examination: Assessments:	Mid Term Examination: Practical	10		
	Final Practical Examination:	20		
	Oral Examination:	10		
	Final Theoretical Examination:	40		
	Total:	100		

Course number: 415	Course title: Virology			
Level/semester: 8 th Se	econd semester			
	Thereof lecture hours: 2			
Credit hours: 2	Thereof practical hours: 0			
	Language: English			
Aims and goals/ skills of the course:	 To Recognize clinically important viruses and their classification, morphology and replication. To describe the diseases caused by these viruses, the clinical presentation, pathogenesis, modes of transmission, laboratory diagnosis, treatment and prevention and control. 			
Content of the course:	I. CONTENTS COVERED Introduction: history and terminology and structure of the virus Virus characters – physical and biochemical Basis of virus classification Virus replication cycle (virus-cell relationship) Virus pathogenesis and virus immunity Laboratory identification of viruses in clinical specimens: Direct diagnosis based on isolation, morphological and biochemical characters Laboratory diagnosis of viruses in clinical specimens: Serological identification, Molecular diagnosis based on identification of virus nucleic acid, using DNA probe, and PCR Hepatitis viruses (HAV, HEV, HDV and TTV) Characters, epidemiology, pathogenesis and laboratory diagnosis Hepatitis viruses (HBV & HCV) – Characters, epidemiology, pathogenesis and laboratory diagnosis Retroviruses: Human Immunodeficiency Virus (HIV): HTLV ₁ , HTLV ₁ characteristics, pathogenesis, epidemiology and laboratory diagnosis Herpes viruses: Types, characteristics, importance and laboratory Herpes viruses: Characteristics, pathogenesis and laboratory diagnosis Encephalitis viruses: Characteristics, pathogenesis and laboratory diagnosis, Riconaviruses Meningitis viruses: Characteristics, pathogenesis and laboratory diagnosis, RTI Viral vaccines an anti-virus therapy Rabies, mea			
Examination:	First midterm examination (30%), second midterm examination (25%), Final examination (40%). Oral exam.(5%)			

Course number: CLS 416	16 Course title: Environmental Microbiology		
Level/semester 9 th level / 2 nd Semester			
<u> </u>	Thereof lecture hours: 1		
Credit hours:	Thereof practical hours: 1		
Language: En	glish		
Aims and goals/ skills of the course: This course is designed to provide the students with the understanding of the vital activities that microorganisms in nature and in the broader dimension of organic activities total ecosystem. Students will learn about the useful and total ecosystem. Students will learn about the useful and total ecosystem of microorganisms in the food and dairy industries use of microorganisms in industrial processes, the treatment waste materials and microbial quality controls will a discussed.			

	Weeks	Subjects
	1.	Fundamentals of microbial ecology- Characteristics of microbial ecosystems, microorganisms and their ability to cause changes; Adaptation
	2.	Microbial symbiosis: Neutralism, mutualism, comensalism, antagonism, competition, parasitism and predation
	3.	Epidemiology of infectious diseases: Terminology, disease reservoirs, modes of transmission, and public health measures for the control of epidemics
	4.	Epidemiology of zoonotic diseases: Viral, bacterial, and parasitic zoonoses; prevention and control of zoonotic diseases
	5.	Air Microbiology – Indoor and outdoor air, airborne diseases and its transmission, control of microorganisms in the air
Content of the course:	6.	Water Microbiology – Water sources, microbial content of water, pollution of water
	7.	Environmental sanitation: Types of sewer lines, treatment of sewage, sewage transmitted diseases
	8.	Microorganisms involved in the spoilage of different types of food and milk
	9.	Microorganisms involved in the spoilage of food continued Food borne diseases: Infection, intoxication;
	10. 11.	Food borne diseases continued Infection, intoxication; prevention and control of food borne diseases Nosocomial infections: Predisposing factors, endogenous and exogenous infections, sources, control measures
	12.	Laboratory acquired infections: Classification of organisms into Risk groups, safety precautions when handling each Risk group, hazardous laboratory techniques and processes
	13.	Microbial production processes: Characteristics of microbial fermenters, different types of microbial processes – dairy, pharmaceutical, single cell protein
Examination:	Written & practical examinationFirst Mid Term Examination:15Second Mid Term Examination:15Laboratory Quizzes:5Mid Term Practical Examination:5Final Practical Examination:20Final Theoretical Examination:40	

Course number: CLS	Course title: Clinical Practice in Microbiology		
Level/semester	9 th level / 2 nd Semester		
	Thereof lecture hours: 1		
Credit hours:	Thereof practical hours: 2		
Language:	English		
ns and goals/ skills of course:	This final microbiology course involves lectures as well as practical sessions in a hospital Microbiology laboratory. The lectures will deal mainly with specimens received in the bacteriology laboratory, their collection, suitability, and processing. The students will be introduced to the safety measures implemented in the laboratories. They will learn about culture media preparation and sterilization, processing of specimens, isolation/ identification of organisms, and antibiotic susceptibility testing. The diagnostic techniques in the routine Clinical Immunology, Mycology and Virology laboratories will		

also be taught

	Weeks	Subjects
	1.	Bacteriological culture media Basic constituents / preparation / sterilisation Types of bacteriological culture media
	2.	Urine Specimens – Types of / collection Processing for chemical analysis
	3.	Bacteriology of urine specimens continued Pathogens isolated from urine specimens Processing of urine specimens for routine culture sterile pyuria
	4.	Blood culture /Indications for Timing and technique for blood collection Manual / Automated processing of blood culture
	5.	Sputum culture / Indication for / Collection Processing of sputum in routine respiratory tract infections
Content of the courses	δ.	Processing of sputum in pulmonary tuberculosis by conventional / automated methods Antimycobacterial susceptibility testing by conventional and automated methods
Content of the course:	7.	Stools specimens / indications for culture Pathogens isoloated from faecal specimens
	8.	Processing of stools for culture / Isolation and identification of different organisms
	9.	Preparation of stools specimen for examination of parasites
	10.	General bench specimens: CSF / Pus / Ear/ Conjunctiva / Genital / Nasal / Throat- indications for taking these specimens / collection / processing
	12.	Commonly used serological tests in clinical bacteriology Widal / CFT / ASO , Serological test of syphilis specific and non-specific: RPR,VDRL,TPHA, FTA ,CRP, Pregnancy test, TORCH, organ transplant matching(MHC)& FC
	13.	Rapid review in diagnosis of common viral infections. Tissue culture types- CPE CPE / EIA / Immunofluorescense
	14.	Rapid review in diagnosis of common fungal infections - Microscopic examination and culture of specimens in Mycology
F	Written & practical e Mid Term Examination Mid Term Examination	n(1): 15 n(2) 15
Examination:	Absentees & reports Final Practical Examin Final Theoretical Exan	

Course number:CLS 421	Course title: Pathophysiology
Level/semester:	7 th level /2 nd semester
	Thereof lecture hours: 2 hrs
Credit hours: 3hrs	Thereof practical hours: 1 hr
Language:	English language
Aims and goals/ skills of the course:	At the completion of this course the student will know the normal structure and functions of the body systems studied. Student should be able to understand the pathophysiology of the most common diseases, its manifestations and the diagnostic methods used.

	<u>CLS 421:</u> Weeks	Lectures Outline Subjects
	1,2,3,4.	 Cardiovascular System Physiology and Manifestations Congenital Heart Disease, Valvular Disease, Rheumatic disease Vascular Disease, Atherosclerosis, Ischemic Heart disease Hypertension, Hypotension, Heart Failure
Content of the course:	5,6,7,8.	Respiratory System Physiology and Manifestations Pneumonia, Tuberculosis Obstructive Disease Respiratory Failure
	9.	Female Genital System - Physiology and Manifestations - Causes of Abnormal Uterine Bleeding, Toxemia
	10,11,12.	Urinary System
	13.	Digestive System
	14,15.	Endocrine System - Physiology and Manifestations - Common endocrine diseases

Examination: Assessments:	written examination		
	1 st Mid Term Examination: Written	20	
	2 nd Mid Term Examination: Written	20	
	Final Practical Examination:	20	
	Final Theoretical Examination:	40	
	Total:	100	

Course number: CLS	6 422 Cou	rse title: Cytopathology	
Level/semester	8 th level / 2 ⁿ	^d Semester	
	Thereof le	ecture hours: 1	
Credit hours:	Thereof p	practical hours: 1	
Language:	English		
Aims and goals/ skills of he course:	theory and malignant examinatic aspirated r through th malignant trained in t	e will provide the student with the basic knowledge of the practical aspect in the diagnosis of tumour cells and non- conditions. This can be achieved by microscopic on of smears prepared from exfoliated cells or fine needle material. The course will enable the student to identify e microscope, the normal and malignant cells by which tumours can be diagnosed. The students also will be the different technical methods applied in smears n and self precautives from contacting infections.	
	Weeks	Subjects	
	1,2.	Introduction to cytology The anatomy & histology of the female genital tract	
	3.	The normal constituents of Pap-smear	
		The normal cytology of female genital tract and other cells of the body	
	3.	Hormonal Cytology	
	4,5,6.	Inflammation: Criteria and causes Diagnostic cytology of different inflammatory Smears	
	7.	Premalignant and Malignant criteria	
Content of the course:	8.	Diagnosis of malignancy in Pap smear "cytology of malignant cells"	
	9.	Introduction to non-gynaecology cytology and the diagnosis of neoplastic and non- neoplastic cells from many parts of the body	
	10.	Sputum, bronchial wash and bronchial brush cytology	
	11.	Body fluids cytology "cytology of serous effusion"	
	12.	Fine needle aspiration cytology (general information)	
	13.	Urine, oral and C.S.F cytology	

rm Examination: 15 Term Examination: 15 Quizzes: 5 ractical Examination: 5 cal Examination: 20

Course number: CLS 43 Level/semester: 8				
	-			
Credit hours: 2 hours	Thereof lecture hours: 1 hour Thereof practical hours: 1 credit hour (3 contact hours per week)			
Language: English				
Aims and goals/ skills of the course:	 This course deals with the theoretical and practical aspects of the chemic structure, mechanisms of action and kinetics of various enzymes. Design assay methodologies is included. The use of enzymes as clinical markers disease related to liver, kidneys, intestines, pancreas, bones & muscles is extensively studied. On successful completion of the course the student must : Understand the chemical specificity of the enzyme structure Be able to study the kinetics of an enzymatically calatalysed react Be able to set up a biochemical assay of enzyme activity Be able to differentially diagnose disease by analysing data of various serum enzyme levels. 			
	Topics	Planned Contact Hours per week	Actual Contact Hours per week	
	Arrangement of enzymes inside cells, catalytic function of enzymes (1week)	1	1	
	Chemical properties of enzymes, stereochemistry and the active site (1 week)	2	2	
	Classification of enzymes (1week)	1	1	
Content of the course:	Enzyme kinetics,Km Value and Effect of Temperature and pH (2 week)	2	2	
	Measurement of enzyme activity in biological fluids (2 weeks)	2	2	
	Enzyme inhibition and types of enzyme inhibitors	1	1	
	Mechanisms of enzyme action. The ES complex (1 week)	1	1	
	Transamination of amino acids and oxidation of the corresponding ketoacids (2 week)	2	2	
	CK, LD< ALP, ACP,ALT,AST,GST,G6DP,Lipase and Amylase as clinical markers of Disease (4 week)	4	4	
Examination:	Midterm examination (15%), Reports (15%) and (40%) and Lab sessions (15%) and Final lab exa			

Course number: CLS 432	2 Course title: Clinical Practice in Biochemistry,			
Level/semester: 8				
	Thereof lecture hours: 1 hour Thereof practical hours: 1 hour			
Credit hours: 2 hours				
Language: English	I			
Aims and goals/ skills of the course:	This course is designed to train students in routine procedures performed in a biochemistry laboratory. Students are encouraged to look up and read review articles and recent advances in the clinical lab practice and changes related to metabolic disorders. In addition, students are expected to become familiar with the up-to-date methodologies related to early diagnosis of such disorders.			
	1 Topics to be Covered Topic	No of Weeks	Contactho urs	
	1. Clinical Specimens safety precaution	1	1	
	2. Quality control	2	1	
	3. ELISA,EIA and RIA	3,4		
Content of the course:	4. Electrophoresis	5,6	2	
	5. Automated equipment	7,8	2	
	 Glucose, cholesterol, Tryglycerides, Urea,Creatinine 	9	1	
	 Sodium,potassium,chloride,CO2,bilirubin,Album in Serum and Urine Samples. 	10	1	
	8. Amylase, ALP, ACP, ALT and AST	11	1	
	9. Review,Problem Solving,Critique	12,13	2	
Examination:	Midterm examination (20%), Reports (20%) and Fina (40%) and Lab sessions (10%) and Final lab examin			

Course number: CLS	441 C	ourse title: Immunohaematology		
Level/semester	7 th level / 2	n ^d Semester		
	There	Thereof lecture hours: 2		
Credit hours:	There	Thereof practical hours: 2		
Language:	English	 English		
Aims and goals/ skills of the course:	proficien of antibo applicati compatit supply w student t situation Advance	The subject develops skills and knowledge required for proficiency in the safe supply of human blood products. A review of antibody antigen interaction will lead into the practical application of antibody screening, identification and compatibility testing. Quality assurance and safety of the blood supply will be recovered. Case study presentation will lead the student through problem solving incompatible reactions, situations they will likely encounter in a working laboratory. Advanced techniques and current developments in stem cell transplant and cord blood banking will also be discussed,		
	Weeks	Subjects		
	1.	Introduction to Immunohaematology		
	2.	ABO Blood Group System		
	3.	The Rh Blood Group System		
	4.	Other Blood Group System		
	5.	Antihuman Globulin (Coombs') Test		
	6.	Detection and Identification of Antibodies		
Content of the course:	7.	Cross Matching (Compatibility Testing)		
	8.	Transfusion Reactions and Complications		
	9.	Screening for Diseases Transmitted through Blood		
	10.	The Blood Donor and Collection of Blood		
	-			
	11.	Storage and Preservation of Blood and Components		
	11.	Storage and Preservation of Blood and Components		
	11. 12.	Storage and Preservation of Blood and Components Haemolytic Disease of the Newborn (HDN)		

	Written & practical examination
Examination:	Mid Term Examination:20Mid Term Practical Examination:20Final Practical Examination:20Final Theoretical Examination:40

Course number: CLS 44	Course title: Clinical Haematology Practices	
Level/semester: level 9, first semester and second semester		
	Thereof lecture hours: 1 hr	
Credit hours:	Thereof practical hours: 3 hours	
Language: English		

<u>Course description</u>		
The students will be introduced to and learn the working system		
of routine haematology laboratory and the Blood Bank. The need		
is for the students to follow the routine laboratory work starting		
from specimen collection right through to sending results to the		
clinicians. Some attention will be given to interpretation of results		
and identification and quick reporting of grossly abnormal life		
threatening results to the clinicians		
In the blood bank, the students will be encouraged in participating in the production of blood products (packed cells, platelets concentrates, fresh frozen plama etc .) and in the production of cryoprecipitate		
AIMS:		
 become familiar with the haematology laboratory; become proficient in basic diagnostic techniques used in a haematology laboratory and understand their use in classifying clinical pathology; be able to identify the structure and function of cellular blood components; understand normal haemostasis and techniques of its determination. become proficient in the morphological interpretation of a blood film; be able to relate changes in blood cell morphology to clinical pathology; develop an understanding of the requirements of a safe blood supply; become proficient in various techniques in antibody screening and identification; become knowledgeable in techniques of compatibility testing and be able to problem solve incompatible reactions; achieve a level of proficiency required for employment in a medical laboratory 		

	1-Haemolytic anaemias: General and Inherited membrane and		
	enzyme defects Bone marrow failure		
	2-Chronic myeloid leukaemia		
	3-Mye]odysplasia		
	4-Multiple myeloma		
	5-Lymphoma: Hodgkin lymphoma, Non-Hodgkin lymphoma		
	6-Disorders of haemostasis: vessel wall and platelets		
	7-Disorders of coagulation: Inherited and Acquired		
Content of the course:	8-Thrombosis and thrombophilia		
	9 -Haernatological aspects of disease: Renal, liver, endocrine		
	pregnancy		
	9-Blood transfusion and Stem cell transplantation		
	10-Blood bank policy		
	11-Complications of blood products		
	12- Case studies		
	13 Assessments of the course		
	Written & practical examination		
Examination:	Mid Term Examination:20Mid Term Practical Examination:20Final Practical Examination:20Final Theoretical Examination:40		

Course number: CLS	Course title: Laboratory Management	
Level/semester	9 th level / 2 nd Semester	
	Thereof lecture hours: 2	
Credit hours:	Thereof practical hours: 0	
Language:	English	
The course is on the different types of medical laboratories, management functions, in clinical laboratories, organization supervision involving laboratory design & space utilization, steps in clinical laboratory work flow, work schedule and wo load measurements, productivity. Emphasis on quality cont clinical laboratories (choice of quality control materials), preventive maintenance and safety measures in quality com monitoring, education of laboratory personnel, budgeting consideration in clinical laboratory		

	Weeks	Subjects	
	1.	Introduction: Clinical Laboratory Organization	
		- Status of laboratories (hospital, public health, reference,	
		specialty, private) The Management functions.	
		- Planning, organizing coordinating, directing, controlling.	
	2.	Organization and Supervision	
		- Laboratory design, space utilization	
		- System approach and organization theory	
	3.	Steps in Clinical Laboratory Workflow	
		- Collection and delivery of specimens	
		- Specimen receiving and entry	
		- Test performance	
		- Recording and reporting results	
		Procedural and Instrumentational Requirements	
		 Work Schedule Workload measurement 	
		 General consideration, Systems of measurement 	
	4.	Productivity	
		- Personnel requirements Choosing the type of	
		employee for the job, Problem solving	
		Personnel Relations	
		- General considerations	
		- Job description	
Content of the course:		- Evaluation of performance	
	5.	Communication in Clinical Laboratory	
		- Channels of communication	
		 Factors affecting communication 	
		 Intra-laboratory and inter-laboratory communication 	
	6.	Budget consideration in Clinical Laboratory	
		- Volume forecasting, labour cost, productivity index,	
		materials and services equipment purchase, budget	
		monitoring	
	7.	Material Management	
	8.	Quality Control in Clinical Laboratory	
		- Implementing Q.C. program	
		- Evaluation of a test methodology	
		- Choice of quality control materials	
	9.	Preventing maintenance in Quality Control Monitoring	
	-	Safety Measures in Quality Control Monitoring	
	10.	Quality Assurance in Clinical Laboratory	
	-	- Certification	
		- Proficiency	
		- Patient test management	
	11.	Accreditation	
	12.	Education of Laboratory Personnel	
		 Continuing education programs 	
	13.	Information Systems	
	Written exar		
Examination:		n Examination: 30	
		Ferm Examination: 30	
	Final Theoretical Examination: 40		

Course number: CLS 4	52 Course title: Indepedant Studies		
Level/semester: 9			
Credit hours: 2 hours	Thereof lecture hours: 2 hour		
	Thereof practical hours: 0 hour		
Language: English			
Aims and goals/ skills of the course:	Is to let the student think, work and present his latest research review in front of his classmates and teachers. This will give a good idea about the student level, his ability to give the lectures in a impressive manner and to assess the level of understanding of the given topic and its presentation Student ability to respond to the questions from teachers and students, the personality of the students and finally to evaluate their English language.		
Content of the course:	 In this course each student is given a virology topic to prepare it from different aspects such as History of the disease and virus and its relation to present situation. Clinical features and details of the disease and its diagnosis Pathogenesis of the virus and the characteristic pathologic features. Epidemeology and transmission of the disease, epidemic and pandemic. Laboratory diagnosis of the viruses by different sensitive and specific methods such as EIA,IF NT and PCR etc. People at risk of these viruses and the advice for them. Control of the virus: By isolation of the patients, giving treatments, analgesic, antipyretic, vaccine, specific immunoglobulin and antiviral therapy. 		
Examination:	Written report (60%), Presentation (40%)		

Course number: CLS 453 Course title: Medical Genetics				
Level/semester: 9				
	Thereof lecture hours: 2 hour			
Ore dit house 0 hours				
Credit hours: 3 hours	Thereof practical hours: 1 hour			
Language: English				
Aims and goals/ skills of the course:	This course is designed to give a basic understanding of the theoretical and practical aspects of medical genetics. In addition to molecular medical genetics, topics such as cytogenetics, DNA, RNA, mutation, mutation detectiontechnique, PCR, cloning, population genetics are also taught. In practical sessions, the commonly used techniques such as gel electrophoresis, DNA & RNA extractions, PCR, RFLP methods and Bioinformatics will be trained.			
Content of the course:	1 Topics to be Covered Topic	No of	Contactho	
		Weeks	urs	
	1. History of Genetics	1	1	
		1	1	
	2. The Cell	1	1	
	3. Cytogenetics (Chromosomes)	1	_	
	4. DNA structure and function	1	1	
	5. Mutation	1	1	
	6. Mutation detection techniques	1	1	
	7 DCD	1	1	
	7. PCR 8. Cloning	1	1	
	9. Population Genetics	1	1	
	10. Clinical Genetics and Genetic disorders	1	1	
	11. Chromosomal Disorders	1	1	
	12. Sickle Cell Anaemia & Thalassemia	1	1	
	First and second midterm examinations (40%), Fina	al practical	examination	
Examination:	(20%)and Final theory examination (40%)			