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Spectroscopic investigations and molecular docking study of 3-(1*H*-imidazol-1-yl)-1-phenylpropan-1-one, a potential precursor to bioactive agents



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ABSTRACT

The optimized molecular structure, vibrational wavenumbers, corresponding vibrational assignments of 3-(1H-imidazol-1-yl)-1-phenylpropan-1-one have been investigated theoretically and experimentally. The calculated geometrical parameters of the title compound are in agreement with the reported XRD data. The HOMO and LUMO analysis is used to determine the charge transfer within the molecule. The stability of the molecule arising from hyper-conjugative interaction and charge delocalization has been analyzed using NBO analysis. Molecular electrostatic potential was performed by the DFT method and from the MEP plot, it is evident that the negative charge covers the carbonyl group and the nitrogen atom N₃ of the imidazole ring and the positive region is over the remaining portions of the molecule. The first and second hyperpolarizabilities are calculated and the first hyperpolarizability of the title compound is derivatives are good object for further studies in nonlinear optics. The docked ligand title compound forms a stable complex with *plasmodium falciparum* and gives a binding affinity value of -5.5 kcal/mol and the preliminary results suggest that the compound might exhibit antimalarial activity against *plasmodium falciparum*.

1. Introduction

An increased incidence of both community-acquired and nosocomial invasive fungal infections has been witnessed over the past few decades. Nosocomial invasive and systemic fungal infections are considered one of the major causes of morbidity and mortality, particularly in patients with weakened or compromised immune systems such as patients with cancer or AIDS and in organ transplant cases [1,2]. In addition, *Candida* species represent the main agent for nosocomial fungal infections worldwide with *Candida albicans* being the cause for the majority of invasive candidiases with about 30–40% of mortality [3]. Antifungal agents

* Corresponding author. E-mail address: cyphyp@rediffmail.com (C.Yohannan Panicker). having azole moiety constitute the mainstay for treatment of fungal infections owing to their safety profile and good therapeutic index [4]. Azoles are nitrogen-containing five membered heterocyclic compounds with electron-rich property. Thereby, they can easily bind with the enzymes and receptors in organisms through weak interactions such as coordination bonds, hydrogen bonds, iondipole, π - π stacking as well as van der Waals force *etc.*, leading to diverse biological activities. The antifungal activity of azoles is attributed to the co-ordination binding of the heterocyclic nitrogen atom to the heme iron atom in the binding site of 14α -demethylase enzyme causing depletion of ergosterol. The ability of azoles to interact with the heme of many host cytochrome P450 enzymes, particularly mammalian CYP3A4, is the reason for the appearance of systemic azole toxicity [5,6]. Imidazole is a 1,3-diaza five membered heterocyclic azole. A plethora of imidazole-bearing antifungals have been used to treat fungal diseases due to their low toxicity and broad antimicrobial spectrum including yeasts and fungal strains. Accordingly, researchers were encouraged to develop new antifungal agents incorporating imidazole moiety [7]. N-Alkylated imidazole moiety is a common feature in many antifungal agents used in clinics, namely clotrimazole, miconazole, econazole and ketoconazole [8]. Therefore, much more attention has been paid in the investigations of such kind of antifungal imidazoles. The clinically available antifungal agents suffer from the development of drug resistance, undesirable side effects and high risk of toxicity [9]. This necessitates continuous efforts to develop new potent antifungal agents with fewer side effects in order to address these therapeutic issues. The title compound is an azole-containing compound bearing *N*-alkylated imidazole moiety and is the building block of a number of anti-*Candida* agents [10–12].

Spectroscopic studies of certain imidazole derivatives are already reported [13–15]. The present work describes the natural bond orbital analysis, molecular frontier orbital analysis, nonlinear optical properties and molecular potential surface study of the title compound. The experimentally observed infrared and Raman bands have been assigned with the help of the calculated vibrational frequencies and potential energy distribution analysis. In addition, a molecular docking study of the title compound is also reported due to the diverse biological activities of the imidazole derivatives [16–18].

2. Experimental details

A pure sample of the title compound was prepared via the reaction of 3-(dimethylamino)-1-phenylpropan-1-one hydrochloride with imidazole in water following the previously reported procedure of Attia et al. [19].

The FT-IR spectrum (Fig. 1) was recorded using KBr pellets on a DR/Jasco FT-IR 6300 spectrometer with a spectral resolution of 2 cm⁻¹. The FT-Raman spectrum (Fig. 2) was obtained on a Bruker RFS 100/s, Germany, and for excitation of the spectrum, the emission of Nd:YAG laser was used, excitation wavelength was 1064 nm, maximal power was 150 mW and measurement was carried out on solid sample.



Fig. 1. FT-IR spectrum of 3-(1H-imidazol-1-yl)-1-phenylpropan-1-one.



Fig. 2. FT-Raman spectrum of 3-(1H-imidazol-1-yl)-1-phenylpropan-1-one.

3. Computational details

All calculations have been performed with the Gaussian09 software [20] using the density functional theoretical method [21]. The theoretical calculations were performed with the hybrid B3LYP functional, i.e. a combination of the Becke's three parameter exchange functional [22] and Lee-Yang-Parr correlation functional [23,24]. The DFT calculations were reported to provide excellent vibrational frequencies of organic compounds if the calculated frequencies are scaled to compensate for the approximate treatment of electron correlation, for basis set deficiencies and for the anharmonicity [25]. The optimized structure parameters (Fig. 3) and frequencies were calculated using the DFT/B3LYP method at 6 - 311 ++ G(d) (5D, 7F) basis set level. The absence of imaginary frequencies confirmed that the stationary points correspond to minima on the potential energy surface. The vibrational assignments of the title compound are done with the help of Guassview program [26] and GAR2PED software [27]. The optimized geometrical parameters with the XRD data are given in Table 1.



Fig. 3. Optimized geometry of 3-(1H-imidazol-1-yl)-1-phenylpropan-1-one.

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Bond lengths (DFT/XI	<u>RD) (Å)</u>		
01–C15	1.2179/1.2152	N2-C19	1.4587/1.4576
N2-C22	1.3672/1.3445	N2-C26	1.3812/1.3612
N3-C22	1.3141/1.3098	N3-C24	1.3747/1.3622
C4-H5	1.0841/0.9300	C4-C6	1.3927/1.3842
C4-C14	1.4018/1.3909	C6-H7	1.0850/0.9300
C6–C8	1.3938/1.3832	C8-H9	1.0854/0.9300
C8-C10	1.3963/1.3762	C10-H11	1.0851/0.9300
C10-C12	1.3892/1.3812	C12-H13	1.0838/0.9300
C12-C14	1.4026/1.3937	C14–C15	1.4976/1.4960
C15-C16	1.5242/1.5052	C16–H17	1.0957/0.9700
C16-H18	1.0958/0.9700	C16-C19	1.5312/1.5142
C19–H20	1.0908/0.9700	C19–H21	1.0903/0.9700
C22-H23	1.0812/0.9300	C24-H25	1.0801/0.9300
C24-C26	1.3718/1.3493	C26-H27	1.0790/0.9300
Bond angles (DFT/XR	D) (1)		
C19-N2-C22	$\frac{-7.0}{126.8/127.0}$	C19-N2-C26	1269/1269
$C_{22} = N_{2} = C_{26}$	106 2/106 0	C22-N3-C24	105 2/104 1
H5-C4-C6	1190/1198	H5-C4-C14	120.6/119.8
C6-C4-C14	120 4/120 5	C4-C6-H7	1199/1199
C4 - C6 - C8	120.1/120.3	H7-C6-C8	120 1/119 9
C6-C8-H9	120.0/120.2	C6-C8-C10	120.1/119.8
H9_C8_C10	120.0/120.1	C8_C10_H11	120.0/119.0
C8_C10_C12	120.0/120.1	H_{11} C_{10} C_{12}	1100/1100
C10_C12_H13	121.0/110.7	C10-C12-C14	1205/1207
H13_C12_C14	121.0/115.7	$C_{10} - C_{12} - C_{14}$	110 1/118 6
$CA_{-}C1A_{-}C15$	122 7/122 5	$C_{12} = C_{14} = C_{12}$	1183/1180
$01 \ C15 \ C14$	122.7/122.5	$01 \ 015 \ 016$	120.2/120.9
C14 $C15$ $C16$	121.0/120.0	C15 C16 H17	120.2/120.8
C14 - C15 - C10	110.7/110.0	C15 - C16 - C10	112 4/112 0
	109.4/109.0		112.4/115.0
	100.0/107.0	H1/-C10-C19	110.1/109.0
H18-C10-C19	109.8/109.0	N2-C19-C16	112.1/111.4
NZ-C19-H20	109.0/109.3	NZ-CI9-HZI	108.1/109.3
10-19-120	110.7/109.5	10 - 19 - 121	110.4/109.5
H20-C19-H21	106.4/108.0	N2-C22-N3	112.4/112.9
N2-C22-H23	121.9/123.6	N3-C22-H23	125.//123.6
N3-C24-H25	121.4/124.6	N3-C24-C26	110.5/110.9
H25-C24-C26	128.1/124.6	N2-C26-C24	105.8/106.1
N2-C26-H27	121.9/126.9	C24–C26–H27	132.3/126.9
Torsion angles (DFT/)	<u>(RD) (°)</u>	C4 C2 C2 C10	0.0/ 1.0
C14-C4C6C8	0.0/0.3	C4-C6-C8-C10	0.0/-1.3
C6–C8–C10–C12	-0.0/0.8	C8-C10-C12-C14	0.0/0.8
C6–C4–C14–C12	-0.0/1.2	C6–C4–C14–C15	179.9/178.0
C10–C12–C14–C4	-0.0/-1.7	C10–C12–C14–C15	-180.0/-177.5
C4-C14-C15-O1	-179.5/-175.7	C12-C14-C15-O1	0.5/5.1
C4-C14-C15-C16	0.6/5.2	C12-C14-C15-C16	-179.5/-173.9
01-C15-C16-C19	0.1/-1.7	C14-C15-C16-C19	-179.9/177.3
C22-N2-C19-C16	-97.4/-87.0	C26-N2-C19-C16	78.2/89.2
C15-C16-C19-N2	179.5/179.6	C24-N3-C22-N2	-0.2/-0.1
C26-N2-C22-N3	0.3/0.2	C19-N2-C22-N3	176.6/177.0
C22-N3-C24-C26	0.0/-0.1	N3-C24-C26-N2	0.2/0.1
<u>C22-N2-C26-C24</u>	-0.3/-0.2	<u>C19–N2–C26–C24</u>	-176.6/-177.1

4. Results and discussion

4.1. IR and raman spectra

The calculated (scaled) wave numbers, observed IR, Raman bands and assignments are given in Table 2 and in the following discussion, the phenyl ring is designated as PhI and imidazole ring as PhII. The bands observed at 1675 cm⁻¹ in the IR spectrum, 1672 cm⁻¹ in the Raman spectrum and at 1671 cm⁻¹ (DFT) are assigned as the stretching modes of the carbonyl (C=O) group. According to literature, the stretching mode of the C=O group is expected in the range 1850–1550 cm⁻¹ [28]. The in-plane and out-of-plane bending modes of the C=O group are reported in the ranges 725 ± 95 and 595 ± 120 cm⁻¹ [29]. For the title compound, the C=O deformation bands are theoretically assigned at 616, 560 cm⁻¹ (DFT).

The stretching and deformation modes of the CH_2 groups appear in the regions 3020–2875, 1480-725 cm⁻¹, respectively [29–31]. The CH₂ stretching modes are observed at 2965, 2921 cm⁻¹ in the IR spectrum, 2999, 2965, 2944, 2918 cm⁻¹ in the Raman spectrum and theoretically in the range 3008–2919 cm⁻¹. The deformation modes of the CH₂ groups are assigned at 1444, 1415, 1366, 1294, 1269, 1252 cm⁻¹ in the IR spectrum, 1444, 1415, 1370, 1351, 1270 cm⁻¹ in the Raman spectrum and at 1441, 1418 (scissoring), 1362, 1355 (wagging), 1296, 1267 (twisting), 1254, 1132 (rocking) cm⁻¹ theoretically as expected [29–31].

For phenyl rings, the C–H stretching modes are expected above 3000 cm^{-1} [29]. In the title compound, the phenyl CH streching modes are theoretically assigned in the range 3079-3042 cm⁻ Experimentally, these bands are observed at 3060 cm⁻¹ in the IR and at 3068 cm⁻¹ in Raman spectrum. The phenyl ring stretching modes are assigned at 1575 cm⁻¹ (IR), 1580 cm⁻¹ (Raman) and in the range 1579–1308 cm⁻¹ (DFT). The sixth phenyl ring stretching mode, the ring breathing vibration appears as a weak band near 1000 cm⁻¹ in mono-, 1-3-di- and 1,3,5-tri substituted benzenes [29,32]. In the present case, the PED analysis gives ring breathing mode at 1009 cm^{-1} for the phenyl ring [29]. The C–H deformation modes of the phenyl ring, in-plane and out-of-plane modes are expected above and below 1000 cm⁻¹, respectively [29]. The inplane CH deformation bands of the phenyl ring are observed at 1063, 1013 cm⁻¹ in the IR spectrum, 1285, 1161, 1139, 1066 cm⁻¹ in the Raman spectrum and theoretically in the range 1283–1010 cm⁻¹. The CH out-of-plane deformation modes are assigned at 907, 825 cm⁻¹ in the IR spectrum, 905, 827 cm⁻¹ in the Raman spectrum and theoretically in the range 969-727 cm⁻¹. The ring substituent deformation modes are also identified and assigned (Table 2) and most of the modes are not pure but contains significant contributions from other modes.

For the title compound, the imidazole ring stretching modes are observed at 1329, 1092, 997 cm^{-1} in the IR spectrum, 1328 cm^{-1} in the Raman spectrum and theoretically at 1479, 1472, 1331, 1097, 998 cm⁻¹. This is in agreement with the work of Almajan et al. [33]. Sandhyarani et al. reported the C–N stretching mode at 1319 cm⁻¹ [34]. Benzon et al. reported the imidazole ring, C-N stretching modes at 1247, 1129, 938 cm⁻¹ theoretically, 1248, 1135, 926 cm⁻¹ in the Raman spectrum and 924 cm^{-1} in the IR spectrum [13]. The C–N stretching modes are reported at 1268, 1220, 1151 cm⁻¹ theoreticaly for benzimidazolium salts by Malek et al. [35]. The C=N stretching modes were reported in the range $1535-1666 \text{ cm}^{-1}$ [36], 1592 cm^{-1} experimentally and theoretically at 1584 cm^{-1} [37]. For an imidazole derivative, the C=N stretching mode is theoeretically assigned at 1464 cm⁻¹, 1462 cm⁻¹ in the IR spectrum and at 1464 cm⁻¹ in the Raman spectrum [13]. The C_{19} -N₂ mode is observed at 1212 cm⁻¹ in the IR spectrum, 1209 cm⁻¹ in the Raman spectrum and theoretically at 1214 cm^{-1} for the title compound.

4.2. Nonlinear optical properties (NLO)

Hyperpolarizabilities are very sensitive to the employed basis set and level of theoretical approach [38–40] because the electron correlation can change the value of hyperpolarizability. Urea is one of the prototypical molecules used in the study of the nonlinear properties of molecular systems. Therefore, it has been frequently used as a threshold value for comparative purposes. The calculations of the total molecular dipole moment, linear polarizability and first order hyperpolarizability from the Gaussian output have been previously explained in detailed [41] and DFT has been extensively used an effective method to investigate the organic NLO material [42]. The polar properties of the title compound were calculated by the density functional theory using the B3LYP method with 6 - 311 ++ G(d) (5D, 7F) basis set using the Gaussian09 program prackage. The calculated first hyperpolarizability of the title compound is 2.209×10^{-30} e.s.u, which is 16.99 times that of

 Table 2

 Calculated (scaled) wavenumbers, observed IR, Raman bands and assignments of the title compound.

<u>B3LYP/6 – 311</u>	<u>++</u> <u>G(d) (5D,7F)</u>		IR	Raman	Assignments ^a
υ(cm ⁻¹)	IR	R _A	$\underline{u(cm^{-1})}$	$\underline{v(cm^{-1})}$	-
3134	3.71	107.96	3131	3133	υCHII(98)
3110	3.34	94.90	3111	3112	UCHII(99)
3108	5.33	42.83	_	_	UCHI(98)
3073	12.13	169.13	_	3068	vCHI(94)
3062	25.38	76.37	3060	_	υCHI(93)
3052	8.37	123.15	-	-	vCHI(96)
3042	0.23	46.53	-	-	υCHI(94)
3008	8.79 14.17	27.14	- 2965	2999	$UCH_2(97)$
2947	3.44	33.54	_	2944	$\nu CH_2(96)$
2919	12.08	84.19	2921	2918	υCH ₂ (99)
1671	193.14	75.13	1675	1672	$\nu C = O(80)$
1579	28.50 12.56	123.96	15/5	1580	VPH(57), 0CH(18) VPh(65), 0CH(19)
1479	34.42	3.71	_	_	υCNII(32), υCCII(25), δCHII(29)
1472	32.62	10.74	_	-	υCCII(24), δCHII(23), υCNII(17), δCH ₂ (14)
1469	9.48	3.68	-	-	δCHI(15), υPhI(55)
1441	11.99	5.76	1444	1444	$\delta CH_2(76)$ $\delta CHI(21) , DDI(60)$
1418	4.60	8.62	1415	1415	δCH ₂ (86)
1362	53.40	8.02	1366	1370	δCH ₂ (53), υCNII(23)
1355	43.31	5.53	-	1351	δCH ₂ (53), υCNII(20)
1331	1.30	45.05	1329	1328	υCNII(59), υCCII(13), δCHII(10)
1308	9.75 48.76	3.98 2.48	 1294	_	$\delta CHI(15), UPDI(63)$ $\delta CH_2(47), UPDI(27)$
1283	9.63	0.64	-	1285	ν PhI(15), δ CH ₂ (17), δ CHI(48)
1267	19.87	9.66	1269	1270	δCH ₂ (41), υCNII(13), δCHII(18)
1254	12.49	12.13	1252	-	δCHII(31), δCH ₂ (23), υCN(12), υCNII(14)
1214	26.11	5.69	1212	1209	$\delta CHII(35), \cup CNII(11), \cup CN(47)$
1159	53.86	8.23	-	1161	δCHI(69)
1142	3.36	6.61	_	1139	δCHI(77), υPhI(17)
1132	1.36	2.09	-	-	δCH ₂ (65), υCNII(22)
1097	18.87	5.96	1092	-	υCNII(64), δCHII(27)
1067	28.16	1.52	1063	1066	δCHII(23), σCHI(35) δCHII(60) μCCII(28)
1034	7.34	13.79	1036	1034	$\nu CC(55), \delta CH_2(19)$
1010	7.56	6.68	1013	-	δCHI(46), υCNII(11), δCH ₂ (14)
1009	9.93	23.06	-	1005	ν PhI(35), δ PhII(10), δ CH ₂ (15)
998 979	8.14 5.49	3.32 41.86	997	977	$DCNII(44), 0CH_2(36), 0PIII(15) \delta PbI(66), DPbI(19)$
969	0.23	0.04	_	_	γ CHI(81), τ PhI(14)
950	57.24	2.35	-	-	ν CC(45), $\delta_{C} = O(11)$, ν PhI(10)
948	0.60	0.02	-	-	γCHI(89)
905 883	1.26	0.09	907	905	γCHI(82) δDbII(82)
824	0.43	0.04	825	827	γ CHI(97)
817	2.59	1.52	_	-	γCHII(81), τPhII(13)
804	0.49	14.82	-	803	δCH ₂ (25), υCC(39), δPhII(10)
781	1.66	0.34	_	_	$\delta CH_2(46), \gamma C = O(13), \tau PhI(10)$
727	48.08	0.28	_	_	τ PhI(30), γ CHI(40), δ CH ₂ (16)
694	1.98	1.50	_	705	δPhI(29), δPhII(14), υCC(30)
689	34.08	1.58	691	-	γCHII(96)
674 647	38.80	0.01	671	670 642	τ PhI(62), γ CHI(30)
616	37.10	3 11	- 620	617	$\delta_c = O(41) \delta PhI(14) \pi PhII(20)$
611	3.47	0.37	_	_	τ PhII(66), δ PhI(11), γ CN(10)
607	1.14	5.69	-	-	δPhI(72)
560	6.70	0.06	562	565	$\gamma C = O(38), \gamma CC(11), \tau PhI(17)$
470 412	4.53	0.12	4/4 411	416	$\tau PhI(60) \ \gamma CC(19) \ \gamma C = O(10)$
396	0.01	0.01	_	395	$\tau PhI(83)$
379	0.52	1.74	-	-	δ CH ₂ (24), δ _C = O(17), δPhI(24)
325	1.08	0.49	-	-	δCN(71)
274 216	5.53 6.68	1.50 1.80	_	_	$\delta CC(46), \delta CH_2(20)$ $\delta CH_2(33), \delta CC(34)$
153	0.00	2.66	_	157	$\gamma CC(31), \tau PhI(31), \tau CH_2(15), \gamma C = O(10)$
138	0.87	0.58	-	_	γCN(41), δCC(38), δCH ₂ (12)
92	0.58	0.11	-	-	$\tau CN(27), \tau CH_2(38), \tau C = O(18)$
61 50	6.35	0.41	-	-	$\tau C = O(30), \tau CH_2(40), \tau CN(12)$
50	2.23	3.02	-	-	$\gamma \text{CIN}(55), 0 \text{CH}_2(40), 0 \text{CC}(10)$

Table 2 (continued)

<u>B3LYP/6 – 311 +-</u>	+ <u>G(d) (5D,7F)</u>		IR	Raman	Assignments ^a
$\underline{v(cm^{-1})}$	IR	R _A	$\underline{v(cm^{-1})}$	$\nu(cm^{-1})$	-
30	0.41	4.20	_	_	$\tau C = O(46), \ \tau CH_2(40)$
28	<u>1.80</u>	<u>7.14</u>	1	<u>-</u>	<u>τCN(50), τCH₂(37)</u>

^a υ-stretching; δ-in-plane deformation; γ-out-of-plane deformation; τ-torsion; PhI-Phenyl ring; PhII-imidazole ring; potential energy distribution (%) is given in brackets in the assignment column.

standard NLO material urea $(0.13 \times 10^{-30} \text{ e.s.u})$ [43]. The average second hyperpolarizability is $<\gamma> = (\gamma_{XXXX} + \gamma_{YYYY} + \gamma_{ZZZZ} + 2\gamma_{XXYY} + 2\gamma_{XXZZ} + 2\gamma_{YYZZ})/5$. The theoretical second order hyperpolarizability was calculated using the Gaussian09 software and is equal to -11.614×10^{-37} e.s.u [44]. Accordingly, we conclude that the title compound is an attractive object for future studies of nonlinear optical properties.

4.3. Natural bond orbital analysis (NBO)

The natural bond orbitals (NBO) calculations were performed using the NBO 3.1 program [45] as implemented in the Gaussian09 package at the DFT/B3LYP level in order to understand various second-order interactions between the filled orbitals of one subsystem and vacant orbitals of another subsystem, which is a measure of the intermolecular delocalization or hyper conjugation. In NBO analysis, large stabilization energy E(2) value shows the intensive interaction between electron-donors and electronacceptors, and greater the extent of conjugation of the whole system, the possible intensive interaction are given in Table 3. The second-order perturbation theory analysis of Fock-matrix in NBO basis shows strong intra-molecular hyper-conjugative interactions are formed by orbital overlap between n(O), n(N) and $\sigma^*(C-N)$, $\pi^*(C-N)$, $\sigma^*(C-O)$, $\pi^*(C-O)$, bond orbitals which result in intramolecular charge transfer causing stabilization of the system. The important intra-molecular hyper-conjugative interactions are $n_2(O_1) \rightarrow \sigma^*(C_{15}-C_{16})$, $n_1(N_2) \rightarrow \pi^*(N_3-C_{22})$ and $n_1(N_3) \rightarrow \sigma^*(N_2-C_{22})$ with stabilization energies 21.23, 47.50 and 9.31 kJ/mol and electron densities 0.05940e, 0.38144e, and 0.04196e. The NBO analysis also describes the bonding in terms of the natural hybrid orbital $n_2(O_1)$, which occupy a higher energy orbital (-0.26945 a.u.) with considerable p-character (100%) and low occupation number (1.88831) and the other $n_1(O_1)$ occupy a lower energy orbital (-0.69024 a.u.) with p-character (43.37%) and high occupation number (1.97778). Thus, a very close to pure p-type lone pair orbital participates in the electron donation to the $n_2(O_1) \rightarrow \sigma^*(C_{15}-C_{16})$, $n_1(N_2) \rightarrow \pi^*(N_3-C_{22})$ and $n_1(N_3) \rightarrow \sigma^*(N_2-C_{22})$ interactions in the compound. The results are tabulated in Table 4.

4.4. Frontier molecular orbital analysis

The HOMO (highest occupied molecular orbital) and LUMO (lowest unoccupied molecular orbital) energy gap of the title compound has been calculated at the B3LYP/6 -311 ++ G(d) (5D,7F) level. These orbitals determine the way the molecule interacts with other species. Fig. 4 shows the distributions and energy levels of HOMO and LUMO orbitals computed at the B3LYP level for the title compound. The HOMO is mainly localized on the imidazole ring, CH₂ and carbonyl groups. The LUMO is mainly delocalized in the CH₂ group, carbonyl group and the phenyl ring. This shows a charge transfer from the imidazole ring to the phenyl ring through the methylene and carbonyl groups. The value of the energy

Table 3

Second-order perturbation theory analysis of Fock matrix in NBO basis corresponding to the intra-molecular bonds of the title compound.

Donor(i)	Type	ED/e	Acceptor(j)	Туре	<u>ED/e</u>	<u>E(2)</u> ^a	<u>E(j)-E(i)</u> ^b	<u>F(ij)</u> ^c
N2-C22	σ	1.98634	N2-C19	σ*	0.02693	1.76	1.14	0.040
_		-	N2-C26	σ^*	0.02244	2.04	1.26	0.045
N3-C22	π	1.86756	C24-C26	π^*	0.01866	21.54	0.33	0.078
C14–C15	σ	1.98023	C4–C6	σ^*	0.01523	1.99	1.24	0.044
-	-	-	C4-C14	σ^*	0.02284	2.13	1.23	0.046
_	-	-	C10-C12	σ*	0.01446	2.25	1.25	0.047
_	-	_	C12-C14	σ^*	0.02166	1.84	1.23	0.042
_	_	_	C16-C19	σ*	0.01883	1.29	1.03	0.033
C15-C16	σ	1.98354	N2-C19	σ^*	0.02693	2.42	0.99	0.044
_	-	-	C12-C14	σ^*	0.02166	2.12	1.21	0.045
C16-C19	σ	1.98194	C14–C15	σ^*	0.06484	2.44	1.05	0.046
C24-C26	σ	1.98579	N2-C19	σ*	0.02693	5.14	1.04	0.065
_	π	1.86234	N3-C22	π^*	0.38144	14.95	0.28	0.061
LPO1	σ	1.97778	C14-C15	σ^*	0.06484	1.86	1.12	0.041
_	-	-	C15-C16	σ*	0.05940	1.50	1.07	0.036
_	π	1.88831	C14–C15	σ*	0.06484	19.61	0.70	0.106
_		_	C15-C16	σ^*	0.05940	21.23	0.65	0.106
LPN2	σ	1.55450	N3-C22	π^*	0.38144	47.50	0.28	0.104
_		-	C16-C19	σ^*	0.01883	6.41	0.62	0.063
_		-	C24–C26	π^*	0.30939	31.60	0.29	0.089
LPN3	σ	1.92739	N2-C22	σ*	0.04196	9.31	0.82	0.078
<u>-</u>		<u>-</u>	C24–C26	σ*	0.01866	5.61	0.95	0.066

^a E(2) means energy of hyper-conjugative interactions (stabilization energy in kJ/mol).

^b Energy difference (a.u) between donor and acceptor i and j NBO orbitals.

 $^{c}\;$ F(i,j) is the Fock matrix elements (a.u) between i and j NBO orbitals.

Table	24
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NBO results showing the formation of Lewis and non-Lewis orbitals.

Bond(A-B)	<u>ED/e</u> ^a	EDA%	EDB%	NBO	<u>s%</u>	<u>p%</u>
σN2-C22	1.98634	64.49	35.51	0.8031(sp ^{2.00})N+	33.38	66.62
-	-0.79862	-	-	0.5959(sp ^{2.32})C	30.12	69.88
πN3-C22	1.86756	56.87	43.13	0.741(sp ^{1.00})N+	0.00	100.0
_	-0.28527	-	-	0.6567(sp ^{1.00})C	0.00	100.0
σC14-C15	1.98023	52.11	47.89	0.7219(sp ^{2.25})C+	30.80	69.20
_	-0.66455	_	_	0.6920(sp ^{1.80})C	35.64	64.36
σC15-C16	1.98354	48.02	51.98	0.6930(sp ^{1.90})C+	34.49	65.51
_	-0.64285	_	_	0.7210(sp ^{2.81})C	26.23	73.77
σC16-C19	1.98194	50.65	49.35	0.7117(sp ^{2.63})C+	27.51	72.49
_	-0.62329	_	_	0.7025(sp ^{2.55})C	28.18	71.82
σC24-C26	1.98579	48.64	51.36	0.6974(sp ^{1.73})C+	36.56	63.44
_	-0.69421	_	_	0.7167(sp ^{1.63})C	38.02	61.98
πC24-C26	1.86234	48.30	51.70	0.6950(sp ^{1.00})C+	0.00	100.0
_	-0.25205	_	_	0.7190(sp ^{1.00})C	0.00	100.0
n101	1.97778	_	_	sp ^{0.76}	56.63	43.37
_	-0.69024			•		
n2O1	1.88831	_	_	sp ^{1.00}	0.00	100.00
_	-0.26945			L.		
n1N2	1.55450	_	_	sp ^{99.99}	0.09	99.01
_	-0.24958					
n1N3	1.92739	_	_	sp ^{2.00}	33.26	66.74
-	-0.34085	<u>-</u>	<i>_</i>	<u>-</u>	<u>-</u>	_

^a ED/e in a.u.



Fig. 4. HOMO-LUMO plots of 3-(1H-imidazol-1-yl)-1-phenylpropan-1-one.

separation between the HOMO and LUMO is 3.222 eV. The lowering of the HOMO-LUMO band gap is essentially a consequence of the large stabilization of the LUMO due to the strong electron-acceptor ability of the electron-acceptor group. Using HOMO and LUMO orbital energies, the ionization energy and electron affinity can be expressed as: I = -E_{HOMO}, A = -E_{LUMO} [46]. The hardness η and chemical potential μ are given the following relations $\eta = (I-A)/2$ and $\mu = -(I + A)/2$, where I and A are the first ionization potential and electron affinity of the chemical species [46]. For the title compound, the E_{HOMO} = -8.662 eV, E_{LUMO} = -5.440 eV, Energy gap = HOMO-LUMO = 3.222 eV, Ionization potential I = 8.662 eV, Electron affinity A = 5.440 eV, global hardness $\eta = 1.611$ eV,

chemical potential $\mu = -7.051$ eV and the global electrophiliciy = $\mu^2/2\eta = 15.430$ eV. It could be concluded that the chemical potential of the title compound is negative and it indicates that the compound is stable.

4.5. Molecular electrostatic potential (MEP)

Molecular electrostatic potential at a point in the space around a molecule gives an indication of the net electrostatic effect produced at that point by the total charge distribution of the molecule and correlates with dipole moments, electronegativity, partial charges and chemical reactivity of the molecules [47]. It provides a visual method to understand the relative polarity of the molecule. The different values of the electrostatic potential at the MEP surface are represented by different colours: red, blue and green represent the regions of most negative, most positive and zero electrostatic potential, respectively. The negative electrostatic potential corresponds to an attraction of the proton by the aggregate electron density in the molecule (shades of red), while the positive electrostatic potential corresponds to the repulsion of the proton by the atomic nuclei (shade of blue). The negative (red and yellow) regions of the MEP were related to the electrophilic reactivity and the positive (blue) regions to the nucleophilic reactivity. From the MEP plot (Fig. 5), it is evident that the negative charge covers the carbonyl group and the nitrogen atom N₃ of the imidazole ring and the positive region is over the remaining portions of the molecule.

4.6. Molecular docking

In addition to the well-documented antifungal activity of imidazole derivatives, several imidazole and fused imidazole structures are associated with a wide range of biological activities including antiviral [48], anticancer [49,50], antitrypanosomal [51], antimalarial [52–54], antihistaminic [55] and anticoagulant [56] activities. Thus, it was of interest to study the molecular docking of the title molecule as an inhibitor of plasmodium falciparum, the causative agent of malaria. High resolution crystal structure of plasmodium falciparum was downloaded from the protein data bank website (PDB ID: 3PBL) and all molecular docking calculations were performed on AutoDock-Vina software [57]. The 3D crystal structure of plasmodium falciparum was obtained from the Protein Data Bank and the protein was prepared for docking by removing the co-crystallized ligands, water and co-factors and the Auto Dock Tools (ADT) graphical user interface was used to calculate the Kollman charges and polar hydrogens. The ligand was prepared for docking by minimizing its energy at the B3LYP/6 - 311 ++ G(d) (5D,7F) level of theory and the partial charges were calculated by Geistenger method. The active site of the enzyme was defined to



Fig. 5. MEP plot of 3-(1H-imidazol-1-yl)-1-phenylpropan-1-one.

include the residues of the active site within the grid size of 40 Å \times 40 Å \times 40 Å and the most popular algorithm, Lamarckian Genetic Algorithm (LGA) available in Autodock was employed for docking and the docking protocol was tested by extracting cocrystallized inhibitor from the protein and then docking the same. The docking protocol predicted the same conformation as was present in the crystal structure with RMSD value well within the reliable range of 2 Å [58]. Amongst the docked conformations. one which binds well at the active site was analyzed for detailed interactions in Discover Studio Visualizer 4.0 software. The ligand binds at the active site of the substrate (Figs. 6 and 7) by weak noncovalent interactions. The amino acids Phe202, Tyr167 shows π - π interaction with the phenyl ring and Asn165, Gln203 forms H-bond interaction with the C=O group and imidazole ring, respectively. The docked ligand title compound forms a stable complex with *plasmodium falciparum* and gives a binding affinity (Δ G in kcal/mol) value of -5.5 (Table 5). These preliminary results suggest that the compound might exhibit antimalarial activity against plasmodium falciparum (Fig. 8).

4.7. Geometrical parameters

In the title compound, the carbon-carbon bond lengths (DFT/ XRD) in the phenyl ring lie in the range 1.3927-1.4026/ 1.3762-1.3937 Å and these values are somewhere in between the normal values for a single (1.54 Å) and a double (1.33 Å) bond [59]. The C_{22} -N₃ bond length (DFT/XRD) = 1.3141/1.3098 Å shows typical double bond characteristics. However, the C_{24} -N₃ = 1.3747/ 1.3622, $C_{26}-N_2 = 1.3812/1.3612$, $C_{22}-N_2 = 1.3672/1.3445$ bond lengths (DFT/XRD) are shorter than the normal C-N single bond length of about 1.48 Å, while the reported corresponding values are 1.407, 1.395, 1.3874 Å [60] and 1.4176, 1.4001, 1.391 Å [13]. At N₂ position, the bond angles (DFT/XRD) are C_{26} - N_2 - C_{22} = 106.2/106.0, $C_{26}{-}N_2{-}C_{19}{=}126.9/126.9$ and $C_{22}{-}N_2{-}C_{19}{=}126.8/127.0^\circ$ and this asymmetry in angles reveal the steric repulsion between the imidazole ring and adjacent CH₂ groups. The imidazole ring is tilted from the adjacent methylene groups, as is evident from the torsion angles, $C_{24}-C_{26}-N_2-C_{19} = -176.6$, $C_{26}-N_2-C_{19}-C_{16} = 78.2$, $N_3 - C_{22} - N_2 - C_{19} = 176.6$ and $C_{22} - N_2 - C_{19} - C_{16} = -97.4^{\circ}$.



Fig. 6. Schematic presentation of the interaction of dopamine D3 receptor and ligand.



Fig. 7. The docked protocol reproduced the co-crystallized conformation with H-bond (green), π -anion (brown), alkyl (pink) and H-bond receptor surface.(For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Table 5

The binding affinity values of different poses of the title compound predicted by AutodockVina.

Mode	Affinity (kcal/mol)	Distance from I	best mode (Å)
_	_	RMSD l.b.	RMSD u.b.
1	-5.5	0.000	0.000
2	-5.2	2.584	3.287
3	-5.1	26.737	28.337
4	-5.1	3.207	4.036
5	-5.0	2.719	3.638
6	-5.0	1.520	2.180
7	-5.0	22.820	26.244
8	-5.0	3.923	5.431
9	-5.0	2.749	4.733



Fig. 8. Pictorial representation of the ligand embedded in the active site of human dopamine D3 receptor.

5. Conclusions

The vibrational spectroscopic studies of 3-(1*H*-imidazol-1-yl)-1phenylpropan-1-one were reported theoretically and experimentally. Potential energy distribution of normal modes of vibrations was done with the help of GAR2PED software. The HOMO is mainly localized on the imidazole ring, CH₂ and carbonyl groups and the LUMO is mainly delocalized in the CH₂ group, carbonyl group and the phenyl ring and the lowering of the HOMO-LUMO energy gap supports for the bioactivity of the molecule. MEP predicts the most reactive part in the molecule and from the MEP plot, it is evident that the negative charge covers the carbonyl group and the nitrogen atom N_3 of the imidazole ring and the positive region is over the remaining portions of the molecule. The first and second hyperpolarizability values are also reported and the title compound is a good object for further studies in nonlinear optics. From the molecular docking study, the ligand binds at the active site of the substrate by weak non-covalent interactions and the amino acids Phe202, Tyr167 shows π - π interaction with the phenyl ring and Asn165, Gln203 forms H–bond interaction with the C=O group and imidazole ring, respectively.

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References

- [1] S.K. Fridkin, W.R. Jarvis, Clin. Microbiol. Rev. 9 (1996) 499–511.
- [2] J.R. Wingard, H. Leather, Biol. Blood Marrow Transpl. 10 (2004) 73-90.
- [3] M.A. Pfaller, D.J. Diekema, Crit. Rev. Microbiol. 36 (2010) 1–53.
- [4] W. Wang, C. Sheng, X. Che, H. Ji, Z. Miao, J. Yao, W. Zhang, Arch. Pharm. Chem. Life Sci. 342 (2009) 732–739.
- [5] J.T. Slama, J.L. Hancock, T. Rho, L. Sambucetti, K.A. Bachmann, Biochem. Pharmacol. 55 (1998) 1881–1892.
- [6] H. Ji, W. Zhang, M. Zhang, M. Kudo, Y. Aoyama, Y. Yoshida, C. Sheng, Y. Song, S. Yang, Y. Zhou, J. Med. Chem. 46 (2003) 474–485.
- [7] B. Narasimhan, D. Sharma, P. Kumar, Med. Chem. Res. 20 (2011) 1119–1140.
 [8] D.J. Sheehan, C.A. Hitchcock, C.M. Sibley, Clin. Microbiol. Rev. 12 (1999) 40–79.
- [9] M.A. Hossain, M.A. Ghannoum, Exp. Opin. Invest. Drugs 9 (2000) 1797–1813.
- [10] M.N. Aboul-Enein, A.A. El-Azzouny, M.I. Attia, O.A. Saleh, A.L. Kansoh, Arch.
- Pharm. Chem. Life Sci. 344 (2011) 794–801. [11] M.I. Attia, A.A. Radwan, A.S. Zakaria, M.S. Almutairi, S.W. Ghoneim, Chem. Cent. J. 7 (2013) 168.
- [12] M.I. Attia, A.S. Zakaria, M.S. Almutairi, S.W. Ghoneim, Molecules 18 (2013) 12208–12221.
- [13] K.B. Benzon, H.T. Varghese, C.Y. Panicker, K. Pradhan, B.K. Tiwary, A.K. Nanda, C. Van Alsenoy, Spectrochim. Acta A 151 (2015) 965–979.
- [14] N. Sundaraganesan, S. Ilakiamani, P. Subramani, B.D. Joshua, Spectrochim. Acta A 67 (2007) 628–635.
- [15] K.B. Benzon, H.T. Varghese, C.Y. Panicker, K. Pradhan, B.K. Tiwary, A.K. Nanda, C. Van Alsenoy, Spectrochim. Acta A 146 (2015) 307–322.
- [16] S.A. Laufer, W. Zimmermann, K.J. Ruff, J. Med. Chem. 47 (2004) 6311–6325.
- [17] Ö. Uçucu, N.G. Karaburun, Í. Işkdağ, Farmaco 56 (2001) 285–290.
- [18] C.A. Hamilton, Pharmacol. Ther. 54 (1992) 231–248.
- [19] M.I. Attia, H.A. Ghabbour, A.A. El-Azzouny, O.A. Al-Deeb, M.S. Almutairi, H.-K. Fun, J. Chem. 2013 (2013) 1–5. Article ID 515309.
- [20] Gaussian 09, Revision B.01, M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Robb, J.R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G.A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H.P. Hratchian, A.F. Izmaylov, J. Bloino, G. Zheng, J.L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J.A. Montgomery, Jr., J.E. Peralta, F. Ogliaro, M. Bearpark, J.J. Heyd, E. Brothers, K.N. Kudin, V.N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J.C. Burant, S.S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J.M. Millam, M. Klene, J.E. Knox, J.B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R.E. Stratmann, O. Yazyev, A.J. Austin, R. Cammi, C. Pomelli, J.W. Ochterski, R.L. Martin, K. Morokuma, V.G. Zakrzewski, G.A. Voth, P. Salvador, J.D. Dannenberg, S. Dapprich, A.D. Daniels, O. Farkas, J.B. Foresman, J.V. Ortiz, J. Cioslowski, D.J.

Fox, Gaussian, Inc., Wallingford CT, 2010.

- [21] W. Kohn, L.J. Sham, Phys. Rev. A140 (1965) 1133-1138.
- [22] A.D. Becke, J. Chem, Phys. 98 (1993) 5648-5652.
- [23] B.K. Paul, N. Guchhait, Comput. Theor. Chem. 1012 (2013) 20-26.
- [24] C. Lee, W. Yang, R.G. Parr, Phys. Rev. B37 (1988) 785-789.
- [25] N.C. Handy, P.E. Masley, R.D. Amos, J.S. Andrews, C.W. Murray, G. Laming, Chem. Phys. Lett. 197 (1992) 506-515.
- [26] R. Dennington, T. Keith, J. Millam, GaussView, Version 5, Semichem Inc., Shawnee Mission, KS, 2009.
- [27] J.M.L. Martin, C.V. Alsenoy, GAR2PED, a Program to Obtain a Potential Energy Distribution from a Gaussian Archive Record, University of Antwerp, Belgium, 2007.
- [28] G. Socrates, Infrared Characteristic Group Frequencies, John Wiley and Sons, New York, 1981.
- [29] N.P.G. Roeges, A Guide to the Complete Interpretation of Infrared Spectra of Organic Structures, John Wiley and Sons Inc., New York, 1994.
- [30] N.B. Colthup, L.H. Daly, S.E. Wiberly, Introduction to Infrared and Raman Spectroscopy, Academic Press, New York, 1990.
- [31] R.M. Silverstein, F.X. Webster, Spectrometric Identification of Organic Compounds, 6th, John Wiley Asia, 2003.
- [32] G. Varsanyi, Assignments of Vibrational Spectra of Seven Hundred Benzene Derivatives, Wiley, New York, 1974.
- [33] G.L. Almajan, S.F. Barbuceanu, L. Farcasanau, C. Draghici, Rev. Chim. Buchar. 62 (2011) 386–390.
- [34] N. Sandhyarani, G. Skanth, S. Berchmanns, V. Yegnaraman, T. Pradeep, J. Colloid Interface Sci. 209 (1999) 154–161.
- [35] K. Malek, A. Puc, G. Schroeder, V.I. Rybachenko, L.M. Proniewich, Chem. Phys. 327 (2006) 439–451.
- [36] S. Genç, N. Dege, A. Çetin, A. Cansiz, M. Sekerci, M. Dincer, Acta Cryst. E60 (2004) o1580.
- [37] H. Tanak, Y. Köysal, Y. Ünver, M. Yavuz, Ş. Işik, K. Sanck, Mol. Phys. 108 (2010) 127–139.
- [38] A.E. Reed, L.A. Curtiss, F. Weinhold, Chem. Soc. 102 (1988) 899–926.
- [39] J.P. Foster, F. Weinhold, J. Am. Chem. Soc. 102 (1980) 7211-7218.
- [40] H. Sekino, R.J. Bartlett, J. Chem. Phys. 84 (1986) 2726–2733.
- [41] J. Henriksson, T. Saue, P. Norman, J. Chem. Phys. 128 (2008) 105–112.
- [42] J.P. Hermann, D. Ricard, J. Ducuing, Appl. Phys. Lett. 23 (1973) 178–180.
- [43] C. Adant, M. Dupuis, J.L. Bredas, Int. J. Quantum. Chem. 56 (2004) 497–507.
 [44] C. Hätting, O. Christiansen, P. Jørgensen, Chem. Phys. Lett. 282 (1998) 139–149.
- [45] E.D. Glendening, A.E. Reed, J.E. Carpenter, F. Weinhold, NBO Version 3.1, Gaussian Inc., Pittsburgh, PA, 2003.
- [46] R.J. Parr, R.G. Pearson, J. Am. Chem. Soc. 105 (1983) 7512–7516.
- [47] S. Sebastian, N. Sundaraganesan, Spectrochim. Acta A 75 (2010) 941–952.
- [48] C.A. Dvorak, R. Apodaca, A.J. Barbier, C.W. Berridge, S.J. Wilson, J.D. Boggs, W. Xiao, T.W. Lovenberg, N.I. Carruthers, J. Med. Chem. 48 (2005) 2229–2238.
- [49] J. Chen, S. Ahn, J. Wang, Y. Lu, J.T. Dalton, D.D. Miller, W. Li, J. Med. Chem. 55 (2012) 7285–7289.
- [50] R. Wang, C. Chen, X. Zhang, C. Zhang, Q. Zhong, G. Chen, Q. Zhang, S. Zheng, G. Wang, Q.-H. Chen, J. Med. Chem. 58 (2015) 4713–4726.
- [51] G. Andriani, E. Amata, J. Beatty, Z. Clements, B.J. Coffey, G. Courtemanche, W. Devine, J. Erath, C.E. Juda, Z. Wawrzak, J.T. Wood, G.I. Lepesheva, A. Rodriguez, M.P. Pollastri, J. Med. Chem. 56 (2013) 2556–2567.
- [52] L. Zhang, R. Sathunuru, D. Caridha, B. Pybus, M.T. O'Neil, M.P. Kozar, A.J. Lin, J. Med. Chem. 54 (2011) 6634–6646.
- [53] A.M. Sanz, F. Gómez-Contreras, P. Navarro, M. Sánchez-Moreno, S. Boutaleb-Charki, J. Campuzano, M. Pardo, A. Osuna, C. Cano, M.J.R. Yunta, L. Campayo, J. Med. Chem. 51 (2009) 1962–1966.
- [54] G.N. Vazquez, M.D.M.R. Vilchis, L.Y. Mulia, V. Melendez, L. Gerena, A.H. Campos, R. Castillo, F.H. Lui, Eur. J. Med. Chem. 41 (2006) 135–141.
- [55] M. Govoni, R.A. Bakker, I. van de Wetering, M.J. Smit, W.M.B.P. Menge, H. Timmerman, S. Elz, W. Schunack, R. Leurs, J. Med. Chem. 46 (2003) 5812–5824.
- [56] J.J. Hangeland, T.J. Friends, K.A. Rossi, J.M. Smallheer, C. Wang, Z. Sun, J.R. Corte, T. Fang, P.C. Wong, A.R. Rendina, F.A. Barbera, J.M. Bozarth, J.M. Luettgen, C.A. Watson, G. Zhang, A. Wei, V. Ramamurthy, P.E. Morin, G.S. Bisacchi, S. Subramaniam, P. Arunachalam, A. Mathur, D.A. Seiffert, R.R. Wexler, M.L. Quan, J. Med. Chem. 57 (2014) 9915–9932.
- [57] O. Trott, A.J. Olson, J. Comput. Chem. 31 (2010) 455-461.
- [58] B. Kramer, M. Rarey, T. Lengauer, Proteins Struct. Funct. Genet. 37 (1999) 228-241.
- [59] P. Sykes, A Guide Book to Mechanism in Organic Chemistry, sixth ed., Pearson Education, India, New Delhi, 2004.
- [60] Y.S. Mary, P.J. Jojo, C.Y. Panicker, C. Van Alsenoy, S. Ataei, I. Yildiz, Spectrochim. Acta A 122 (2014) 499–511.